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Melioidosis: an important cause of pneumonia in residents of and travellers returned from endemic regions

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ABSTRACT: Melioidosis is endemic in South East Asia, Asia and northern Australia. Infection usually follows percutaneous inoculation or inhalation of the causative bacterium, *Burkholderia pseudomallei*, which is present in soil and surface water in the endemic region. While 20–36% of melioidosis cases have no evident predisposing risk factor, the vast majority of fatal cases have an identified risk factor, the most important of which are diabetes, alcoholism and chronic renal disease.

Half of all cases present with pneumonia, but there is great clinical diversity, from localised skin ulcers or abscesses without systemic illness to fulminant septic shock with multiple abscesses in the lungs, liver, spleen and kidneys. At least 10% of cases present with a chronic respiratory illness (sick >2 months) mimicking tuberculosis and often with upper lobe infiltrates and/or cavities on chest radiography. As with tuberculosis, latency with reactivation decades after infection can also occur, although this is rare.

Confirmation of diagnosis is by culture of *B. pseudomallei* from blood, sputum, throat swab or other samples. Microbiology laboratories need to be informed of the possibility of melioidosis, as those not familiar with it can misidentify the organism. Antibiotic therapy is initial intensive therapy with *i.v.* ceftazidime or meropenem or imipenem +/- cotrimoxazole for ≥10 days, followed by eradication therapy with cotrimoxazole +/- doxycycline +/- chloramphenicol (first 4 weeks only) for ≥3 months.

Melioidosis has been increasingly recognised in returning travellers in Europe and recently melioidosis and colonisation with *B. pseudomallei* have been documented in cystic fibrosis patients visiting or resident in endemic areas.

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The history of *Burkholderia pseudomallei* and melioidosis

In Rangoon, Burma, in 1912, WHITMORE and KRISHNASWAMI [1] described cases of a newly recognised septicaemic disease. Some fatal cases were morphia addicts and autopsy findings were characterised by widespread caseous consolidation of the lungs, classically with abscesses in the liver, spleen, kidney and subcutaneous tissues. This is the fulminant end of the spectrum of a disease now recognised as a major cause of fatal septicaemia in endemic tropical locations. The Gram-negative bacillus isolated from the lungs and other organs was similar to that causing glanders (*Burkholderia mallei*), but was motile. WHITMORE [2] also noted the clinical resemblance to glanders. The disease was named melioidosis by STANTON and FLETCHER [3], based on a derivation of the Greek *melis* (distemper of asses). After various earlier names, the causative bacterium was called *Pseudomonas pseudomallei* [4]. Due to 16s ribosomal ribonucleic acid sequences, deoxyribonucleic acid (DNA)-DNA homology values, the cellular lipid and fatty acid composition, and phenotypic characteristics within the *Pseudomonas* genus, seven species were moved to a new

genus, *Burkholderia* [5], in 1992. The new genus was named after the US microbiologist Walter Burkholder, who, in 1949, first described *B. cepacia*, formerly *P. cepacia*, as the plant pathogen causing bacterial rot of onions ("slippery skin") [6]. *B. cepacia* is the type species in the genus, which includes the organisms causing melioidosis (*B. pseudomallei*) and glanders (*B. mallei*). A number of additional related environmental bacterial species have subsequently been discovered and added to the *Burkholderia* genus.

***Burkholderia pseudomallei* in soil and water**

Although originally considered a zoonotic infection from rodents [7], zoonotic infection is actually very rare, with only three possible cases described from Australia [8]. It is now clear that humans and animals are infected by exposure to *B. pseudomallei* present in soil and surface water in endemic locations [9].

The interaction of *B. cepacia* and other species in the *Burkholderia* genus with environmental elements has relevance

for *B. pseudomallei* [10]. *B. cepacia* strains have an ecological niche in the rhizosphere of plants and there is great interest from the agricultural industry in using *B. cepacia* as a biological control agent [11]. *B. cepacia* can degrade some toxic compounds in pesticides and herbicides, and can kill many soil-borne plant pathogens. However, rapid mutation and adaptation is considered likely, given the presence of numerous insertion sequences within *B. cepacia* [11], including, for some strains, sequences identical to *B. pseudomallei* insertion sequences [12]. This justifies concern that widespread agricultural use of *B. cepacia* may be a hazard to human health and, most particularly, for patients with cystic fibrosis (CF) [11]. Recent molecular studies have confirmed, for the first time, that as with *B. pseudomallei*, human pathogenetic strains of *B. cepacia* from CF patients can be found in local agricultural soils [13].

Endemic melioidosis

Following the initial account in Burma, melioidosis was documented in humans and animals in Malaysia and Singapore from 1913 [7] and then in Vietnam from 1925 and Indonesia from 1929 [14, 15]. The majority of cases of melioidosis are from South East Asia and northern Australia.

Thailand has reported the greatest numbers [16–18], with an estimated 2,000–3,000 cases of melioidosis each year [19]. Melioidosis is also common in Malaysia [20–22] and Singapore [23–28]. Other countries in the region where melioidosis is recognised in humans and animals include China (especially Hong Kong), Taiwan and Brunei [15, 29–33]. Melioidosis is also likely to occur in Cambodia, Laos and the Philippines [15, 19, 34]. Melioidosis has been increasingly recognised in India [29, 35, 36], although reports that some of the “plague” scares of 1994 may have been cases of melioidosis [37] have been disproved [38, 39]. Cases have been reported from Sri Lanka, Bangladesh and Pakistan [15, 40]. A recent study from southern Vietnam confirmed that melioidosis is present but far less common than in north-east Thailand [41]. Cases of melioidosis have also been documented in Papua New Guinea [42] and Fiji, but the extent of endemicity in the Pacific islands remains to be defined.

The two locations where melioidosis is arguably the most important single bacterial pathogen for humans are some north-east provinces in Thailand and the top end of the Northern Territory of Australia. In north-east Thailand, 20% of community-acquired septicaemic cases are due to melioidosis, which accounts for 39% of fatal septicaemias [17] and 36% of fatal community-acquired pneumonias [43]. At the Royal Darwin Hospital in the Northern Territory, melioidosis is the most common cause of fatal community-acquired bacteraemic pneumonia [44]. Within Thailand, melioidosis occurs in other northern provinces and to a lesser extent in the south, and it is also well recognised in other regions in tropical northern Australia, such as northern Queensland [45–47] and the Kimberley region of northern Western Australia [48].

Global warming may well result in expansion of the endemic boundaries of melioidosis. In Australia it is predicted that temperature, overall rainfall and rainfall intensities will all increase in northern regions, possibly facilitating a southern extension of melioidosis [49].

Melioidosis introduced into nontropical regions

In addition to endemic melioidosis, there are several documented situations where melioidosis has become established in nontropical locations. In France, in the 1970s, cases of melioidosis occurred in animals in a Paris zoo and then

spread to other zoos and equestrian clubs in France [15]. As well as fatal animal and human cases, there was extensive soil contamination persisting for some years. *B. pseudomallei* was considered likely to have been introduced by importation of infected animals, possibly a panda donated to France by Mao-Tse-Tung or horses from Iran [15]. A cluster of cases occurred over a 25-yr period in south-west Western Australia (31°S), involving animal cases and one human infection in a farmer [50]. Ribotyping of the farm animal and human isolates and one isolate from the soil showed identical patterns [51]. This supports the suggestion of clonal introduction of *B. pseudomallei* into this temperate region, probably via an infected animal, with environmental contamination, local dissemination and persistence over 25 yrs.

Melioidosis in travellers to and returned from endemic regions

Melioidosis was an important cause of morbidity and mortality in foreign troops fighting in South East Asia. DANCE [15] noted that ≥ 100 cases occurred among French forces in Indochina between 1948–1954, and by 1973, 343 cases had been reported in American troops fighting in Vietnam [52]. Concerns of re-activation of latent infection in soldiers returned from Vietnam, with estimates from serology studies of ~225,000 potential cases, resulted in melioidosis being called the “Vietnamese time bomb” [53, 54]. However, while occasional cases of re-activation of *B. pseudomallei* still occur in Vietnam veterans [55, 56], it is rare in comparison to the numbers exposed.

Melioidosis remains a risk for travellers to endemic areas, especially if they have a recognised risk factor as discussed below. Adventure tours resulting in extensive exposure to wet season soils and surface water are a possible scenario for infection. This occurred in early 2002 in a young French tourist who died from fulminant melioidosis septicaemic pneumonia after exposure to muddy soils on a camping tour of a tropical Australian wilderness park. Imported melioidosis cases are seen each year in hospitals in southern Australia [44]. Some of these cases are acquired in tourists returning from northern Australia and others are acquired in South East Asia. Occasional imported cases are also reported from Europe and the USA [40, 57, 58]. While the vast majority of these cases are from recent acquisition of *B. pseudomallei*, as occurs with >90% of cases in northern Australia [59], occasional patients may have chronic melioidosis or have re-activation of disease from a latent focus many years after leaving an endemic area, as noted above in returned soldiers from Vietnam.

With increasing movement of people between countries and with expanding live animal exports from endemic regions such as northern Australia, emergence of new foci of melioidosis may well occur in addition to the diagnosis of imported cases in hospitals around the world in people returning from or travelling in endemic regions.

How does infection occur?

In most endemic regions there is a close association between rainfall and melioidosis. In north-east Thailand [60] and northern Australia [61], 75 and 85% of cases, respectively, occur in the wet season. Although the earliest animal studies had shown infection with *B. pseudomallei* through oral or nasal exposure and from ingestion, recent reviews consider that the majority of human cases are from percutaneous inoculation of *B. pseudomallei* following exposure to muddy soils or surface water in endemic locations [4, 14, 44]. Ingestion

[44] and sexual transmission [62] have been suggested as unusual modes of transmission of *B. pseudomallei*.

Presentations with melioidosis pneumonia following presumptive inoculating skin injuries have been documented in patients with soil-contaminated burns [63] and are common in tropical Australia [44, 61]. An animal model has also shown spread of *B. pseudomallei* to the lungs following burn inoculation [63]. This suggests haematogenous spread to the lung rather than inhalation or spread from the upper respiratory tract. This is analogous to post-primary tuberculosis, with disease from haematogenous spread localising in the upper lung zones. Support for this comes from the observation that septicaemic melioidosis pneumonia cases are often more systemically ill than suggested by initial chest radiography [44].

However, it is likely that under certain epidemiological conditions inhalation is the main mode of infection. This was suggested for soldiers exposed to dusts raised by helicopter rotor blades in Vietnam [64]. Melioidosis following near drowning is well documented, with the probable infecting event being aspiration [31, 65, 66]. Recently, it has been shown that intensity of rainfall is an independent predictor of melioidosis presenting as pneumonia and of a fatal outcome (B.J. Currie, unpublished data). It was postulated that this might be explained by heavy monsoon rainfall and winds resulting in a shift towards inhalation as the mode of infection with *B. pseudomallei*. The occurrence of a more fulminant disease with a high mortality when inhalation is the mode of infection rather than percutaneous inoculation is analogous to inhalational anthrax and plague [67, 68].

The natural history of infection with *Burkholderia pseudomallei*

Serology studies indicate that most infection with *B. pseudomallei* is asymptomatic [45, 69–71]. In north-east Thailand, a majority of the rural population is seropositive by indirect haemagglutination (IHA) [60], with most seroconversion occurring between 6 months and 4 yrs of age [71]. Although melioidosis occurs in all age groups, severe clinical disease, such as septicaemic pneumonia, is seen mostly in those with risk factors such as diabetes, renal disease and alcoholism [4, 46, 60, 61]. In addition to infection by inhalation, bacterial load on exposure (inoculating dose) and virulence of the infecting strain of *B. pseudomallei* are also likely to influence severity of disease. However, it has been noted that, despite the large bacterial load in severely ill patients with septicaemic pulmonary melioidosis, person-to-person transmission is extremely unusual [9, 72]. This, together with the rarity of fulminant melioidosis in healthy people, supports the primary importance of host risk factors for the development of melioidosis.

To date there is no definitive evidence for development of immunity from melioidosis with natural exposure to *B. pseudomallei* and re-infection has occurred with a different strain of *B. pseudomallei* following successful treatment of melioidosis [73, 74].

The incubation period for melioidosis is likely to be influenced by inoculating dose, mode of infection, host risk factors and differential virulence of infecting *B. pseudomallei* isolates [64, 75]. Rapid onset of melioidosis (even within 24 h of inoculation) has been seen in presumed aspiration following near drowning [31, 65], as well as in a small number of patients with heavy environmental exposure [59]. In 25 cases where a clear incubation period could be determined between the inoculating injury and the onset of symptoms, the incubation period was 1–21 days (mean 9 days) [59]. This is consistent with a series of nosocomial melioidosis cases from

Thailand, where the incubation period was 3–16 days (mean 9.5 days) [76].

Melioidosis presentations can be acute or chronic disease [59]. Chronic disease is defined as illness with symptoms for >2 months duration on presentation. Of 252 cases of culture-confirmed melioidosis, 222 (88%) presented with acute illness and 30 (12%) with chronic illness, with no fatalities in the latter group [59].

It has long been recognised that, analogous to tuberculosis, *B. pseudomallei* has the potential for re-activation, hence, the concern of the Vietnamese time bomb in returned soldiers (see above). Latent periods from exposure to *B. pseudomallei* in an endemic region to onset of melioidosis in a nonendemic region have been documented to be as long as 29 yrs [56]. However, documented cases of re-activated *B. pseudomallei* are very uncommon. The small numbers of such cases in the USA in comparison to the estimated 225,000 seropositive soldiers who returned from Vietnam [53, 54], suggest re-activation of *B. pseudomallei* is an infrequent event.

Most cases of melioidosis in endemic areas are recent infections presenting with acute illness [15, 17, 44–46]. In the Darwin study, only 3% (8 out of 252) of cases were considered to be re-activation of *B. pseudomallei* from a latent focus [59]. Most cases of re-activation present as pulmonary melioidosis. This is consistent with the latent focus usually being in the lungs, with *B. pseudomallei* probably dormant within pulmonary macrophages or other cells. Re-activation of melioidosis has been associated with influenza [55], other bacterial sepsis [77] and development of known melioidosis risk factors such as diabetes. It remains unknown what proportion of asymptomatic seropositive people actually have latent infection with the potential for re-activation.

The pathogenesis of melioidosis

There has been a resurgence of interest in *B. pseudomallei* and melioidosis pathogenesis in the last decade. Concerns of possible bio-terrorism using the bacteria or its virulence components in genetically engineered constructs [78] and of exposure of military personnel to *B. pseudomallei* have driven funding for recent work. There are excellent reviews of recent developments and the search for virulence factors and potential vaccines [79–83].

While it is clear from laboratory studies of isolates of *B. pseudomallei* from animals, humans and the environment that there is differing virulence amongst *B. pseudomallei* isolates [84–86], the importance of this variation in virulence in determining clinical aspects of melioidosis remains uncertain. Molecular typing showing clonality of isolates in animal and human clusters in Australia has revealed that the same outbreak strain can cause different clinical presentations, with host factors being most important in determining the severity of disease [8, 44, 87]. Furthermore, molecular studies both within and between endemic regions are required to ascertain if clinically significant strain variation occurs, both in relation to virulence and tissue or organ specificity.

Immune response and host risk factors for melioidosis

There are important host protective mechanisms against *B. pseudomallei* in cytokine responses, as well as potentially detrimental ones [88, 89]. However, the predominant association with fatal melioidosis is the presence of defined patient risk factors [44, 61]. The most important risk factors for melioidosis are diabetes, alcohol excess and renal disease, as summarised in table 1 [4, 16, 17, 46, 60, 61, 90, 91].

Table 1. – Summary of risk factors for melioidosis

	Thailand	Australia
Diabetes	23–60	37
Alcohol excess	12	39
Renal disease	20–27	10
Chronic lung disease		27
Thalassemia	7	Nil
Malignancy, steroid therapy		
Iron overload		
No risk factors	36	20

Data are presented as %.

In the Darwin study, 51 out of 252 cases (20%) had no identified risk factor and there was only one fatality in this group [61], confirming that severe disease and fatalities are uncommon in those without risk factors. Risk factors are less commonly present in children than in adults [92–95].

The current author has previously suggested that the predisposition to melioidosis in diabetics, cases of alcohol excess and those with chronic renal disease may reflect their impaired polymorphonuclear leukocyte (PMNL) functions, such as mobilisation, delivery, adherence and ingestion [61]. There has been recent support for the role of PMNL in initial host containment of infections with other fast-replicating intracellular bacteria such as *Listeria monocytogenes*, *Salmonella typhimurium* and *Mycobacterium fortuitum* [96]. Melioidosis has also been described in chronic granulomatous disease [97]. The possible primary role of PMNL function in containing *B. pseudomallei* led to empirical use of granulocyte colony-stimulating factor (G-CSF) in patients with strictly defined septic shock in the intensive care unit at Royal Darwin Hospital. Uncontrolled data show a significant improvement in survival with G-CSF (see below) [61, 98].

The clinical diversity of melioidosis

The earliest descriptions of melioidosis documented the fulminant end of the clinical spectrum, with abscesses throughout both lungs and in many organs [1, 2]. At the other end of the spectrum are asymptomatic infections and localised skin ulcers or abscesses without systemic illness. Inoculating dose of *B. pseudomallei*, mode of transmission, virulence of the infecting bacterial strain, and host predisposition and underlying risk factors will all influence the clinical presentation and outcome.

HOWE *et al.* [64] classified melioidosis as acute, subacute and chronic, while the Darwin study separated acute and chronic disease [59]. In the Darwin study, 46% of cases were bacteraemic and overall mortality was 19% [61], compared with 60% bacteraemia and 44% mortality in Thailand [60], and 52% bacteraemia and 46% mortality in Singapore [27].

Pulmonary melioidosis

Pneumonia is the commonest clinical presentation of melioidosis in all studies with around half of all melioidosis patients presenting with pneumonia [4, 14, 16, 17, 21, 27, 46, 61, 64, 99]. The proportion with pneumonia is much higher in those with chronic disease and those presenting after reactivation from a latent focus. In the Darwin prospective study, pneumonia was the primary diagnosis in 127 out of 252 cases (50%) and an additional 21 cases (8%) developed pneumonia after another primary diagnosis [61].

Acute melioidosis pneumonia has a spectrum from fulminant

septic shock (mortality 84% in the Darwin study) to mild undifferentiated pneumonia, which can be acute or subacute in nature, both with little mortality. Septicaemic patients present as acutely unwell with high fevers and prostration and often little initial cough or pleuritic pain [17, 63]. There may also be multiple abscesses in abdominal organs. On chest radiography they often have diffuse nodular infiltrates throughout both lungs, which coalesce, cavitate and progress rapidly, consistent with the caseous necrosis and multiple metastatic abscess formation seen at autopsy [63, 100, 101]. However, some septicaemic pneumonia patients have a more predominant cough with productive sputum, dyspnoea and chest radiography showing discrete but progressive consolidation in one or more lobes (fig. 1) [100–102]. Acute pneumonia with upper lobe consolidation in endemic regions warrants consideration of melioidosis (fig. 2). While such upper lobe disease has predominated in some reports [101,



Fig. 1. – Acute fulminant septicemic melioidosis.



Fig. 2. – Acute melioidosis pneumonia.

102], lower lobe infiltrates were more common overall in non-septicaemic acute and subacute melioidosis from one study [100]. Pleural effusions have generally been uncommon in acute melioidosis [102], but effusions and empyema can still occur, especially with lower lobe disease [100, 101].

Patients with chronic pulmonary melioidosis have fevers, weight loss and a productive cough, sometimes with haemoptysis [102, 103]. Pleuritic chest pain occurs in half [103]. Disease is often slowly progressive over months (fig. 3). It can also be remitting and relapsing over many years, but acute deterioration with septicaemia may also occur. Classically upper lobe changes with infiltrates and/or cavitation are seen on chest radiography, being present in 37 out of 39 (95%) chronic cases in one study [103]. Initial chest radiography showed cavitory disease with or without infiltrates in 27 out of 39 (69%) and infiltrates in 12 out of 39 (31%) [103]. Pleural effusions were present in two out of 39, hilar adenopathy in only one out of 39 and only three out of 39 had bilateral disease. The cavities are usually single and thin walled and rarely contain an air-fluid level [102–104]. Computed tomography (CT) scan may show small cavities not evident on chest radiography [104]. There are numerous reports of chronic melioidosis being initially misdiagnosed as tuberculosis.

Re-activation of melioidosis from a latent focus can present as fulminant or less severe melioidosis, with chest radiography changes as in the range of acute disease described above [63]. Chest radiography before re-activation may be normal or may show focal changes from within which reactivation occurs [59].

Melioidosis and cystic fibrosis

An important recent observation is that *B. pseudomallei* can both colonise lungs and cause disease in patients with CF. There have been at least three cases of melioidosis in CF patients returning to Europe from a melioidosis-endemic region [40, 57, 58]. The similarity to infection with *B. cepacia*



Fig. 3. – Chronic pulmonary melioidosis.

in CF is of concern, given the association of *B. cepacia* with more rapid deterioration in lung function [13, 105]. Most recently, in a series of four patients with CF infected in northern Australia, there was suggested transmission of *B. pseudomallei* between two affected siblings [106]. Person-to-person transmission of *B. pseudomallei* is very unusual and this, together with the prolonged carriage time of *B. pseudomallei* seen in some of the CF cases, parallels the specific predilection seen for *B. cepacia*. Patients with CF travelling to melioidosis-endemic locations should be warned of the risk of melioidosis and this should be considered if they become sick after returning.

A summary of other presentations of melioidosis is given in table 2.

Laboratory diagnosis of melioidosis

Definitive diagnosis of melioidosis requires a positive culture of *B. pseudomallei*. Melioidosis must be considered in febrile patients in or returning from endemic regions to enable appropriate samples and laboratory awareness. *B. pseudomallei* readily grows in commercially available blood culture media but it is not unusual for laboratories in nonendemic locations to misidentify the bacteria. Culture from nonsterile sites can be problematic and the likelihood of successful culture is increased if sputum, throat swabs, ulcer/skin lesion swabs and rectal swabs are placed into Ashdown's medium, a gentamicin-containing liquid transport broth that results in selective growth of *B. pseudomallei* [107]. Identification of *B. pseudomallei* can be made by combining the commercial API 20NE or 20E biochemical kit with a simple screening system involving Gram stain, the oxidase reaction, typical growth characteristics and resistance to certain antibiotics [108, 109]. Some commercial identification systems are poor at identifying *B. pseudomallei* [110]. Any confirmed culture of *B. pseudomallei* should be considered to represent active infection, although the rare possibility of sputum colonisation in people with chronic lung disease such as CF has recently been noted [59, 106].

There are a variety of antigen- and DNA-detection techniques used in endemic regions but these are not yet commercially available [109, 111, 112]. IHA and other serological assays are available. In endemic areas their usefulness is limited by high rates of background antibody positivity [60, 112]. In acute septicemic melioidosis, IHA is often initially negative, but repeat testing may show seroconversion. A positive IHA in a tourist returning from a melioidosis-endemic region is useful in supporting the possibility of melioidosis, but definitive diagnosis still requires a positive culture.

Table 2. – Summary of other presentations of melioidosis

Skin ulcers/abscess(es)
Bacteremia no focus evident
Spleen, liver, kidney, adrenal abscess(es) +/- bacteremia
Prostatic abscess(es)
Septic arthritis
Parotid abscess
Encephalomyelitis
Lymph gland mass(es)
Mycotic aneurysm
Soft tissue abscess
Other genitourinary (e.g. orchitis)
Osteomyelitis
Muscle abscess(es) (e.g. psoas)
Brain abscess(es)
Mediastinal mass
Pericardial collection

Antibiotic therapy for melioidosis

B. pseudomallei is characteristically resistant to penicillin, ampicillin, first and second generation cephalosporins, gentamicin, tobramycin, and streptomycin [4, 113, 114]. Before 1989, “conventional therapy” for melioidosis consisted of a combination of chloramphenicol, trimethoprim/ sulfamethoxazole (cotrimoxazole), doxycycline and sometimes kanamycin, given for 6 weeks–6 months [4, 115].

Studies subsequently showed *B. pseudomallei* to be susceptible to various newer β -lactam antibiotics, especially ceftazidime, imipenem, piperacillin, amoxicillin/clavulanate, ceftriaxone and cefotaxime [116–122]. Unlike with “conventional antibiotics”, most studies suggested these newer β -lactam antibiotics to be bactericidal for *B. pseudomallei* [118, 120].

Initial intensive therapy for melioidosis

The most important therapeutic study for melioidosis was an open-label randomised trial in Thailand comparing ceftazidime (120 mg·kg⁻¹·day⁻¹) with “conventional therapy”, which showed that ceftazidime was associated with a 50% lower overall mortality in severe melioidosis [115]. Ceftazidime then became the drug of choice for initial intensive therapy for melioidosis. Another study from Thailand showed similar results when ceftazidime was used in combination with cotrimoxazole for initial intensive therapy [123]. Whether cotrimoxazole added to ceftazidime is superior to ceftazidime alone is currently being studied in Thailand.

The carbapenems, imipenem and meropenem, have the lowest minimum inhibitory concentrations against *B. pseudomallei* [118–122, 124]. Most recently, high-dose imipenem has been shown in another comparative trial from Thailand to be at least as effective as ceftazidime for severe melioidosis, with no differences in mortality between the groups and fewer treatment failures in those given imipenem [99].

In patients with septic shock, preliminary data suggest addition of G-CSF in addition to state-of-the-art intensive care management may increase survival [61, 98].

The duration of initial intensive therapy should be ≥ 10 days [114], with the Darwin study guidelines stating at least 14 days, with a longer time period required if critically ill, or for extensive pulmonary disease, deep seated collections or organ abscesses, osteomyelitis, and septic arthritis and neurological melioidosis [61, 125].

Ceftazidime infusions (6g over 24 h, adult dose) through a peripherally inserted central catheter (PICC line) using an elastomeric infusion device (Baxter, Sydney, Australia) in home therapy have enabled early discharge from hospital [61]. The absence of any post-antibiotic effect with ceftazidime allows such a continuous infusion, a theoretical advantage over intermittent dosing [126].

Subsequent eradication therapy for melioidosis

Following initial intensive therapy, using ceftazidime, imipenem or meropenem, possibly in combination with cotrimoxazole, subsequent eradication therapy is considered necessary for preventing re-crudescence or later relapse of melioidosis.

Both duration of eradication therapy and the best antibiotics to use remain unclear. Molecular typing of isolates from recurrent melioidosis has confirmed that by far the majority are true relapses from failed eradication, rather than new infection [59, 73, 74, 127, 128].

Table 3. – Summary of therapy for melioidosis

Initial intensive therapy, minimum of 14 days
Ceftazidime, 50 mg·kg ⁻¹ up to 2g, every 6 h
or Meropenem, 25 mg·kg ⁻¹ up to 1 g, every 8 h
or Imipenem, 25 mg·kg ⁻¹ up to 1 g, every 6 h
+/- Trimethoprim/sulfamethoxazole, 8/40mg·kg ⁻¹ up to 320/1600 mg, every 12 h
Eradication therapy, minimum of 3 months
Trimethoprim/sulfamethoxazole, 8/40 mg·kg ⁻¹ up to 320/1600 mg, every 12 h
+/- Doxycycline, 2 mg·kg ⁻¹ up to 100 mg, 12 h

The most important recent trial of eradication therapy was a comparison of doxycycline alone (the most commonly used eradication regimen in the Darwin study until 1998) versus “conventional” chloramphenicol (first 4 weeks only), cotrimoxazole and doxycycline in combination [129]. Relapses were significantly commoner in the doxycycline alone group, resulting in a recommendation that doxycycline should not be used alone as a first-line eradication therapy [129]. Similar failures of doxycycline alone as eradication therapy were noted in the Darwin study [59], with some *Burkholderia pseudomallei* relapse isolates showing acquired doxycycline resistance [130]. Since changing to eradication therapy with cotrimoxazole alone in the Darwin study, relapses have been almost exclusively in noncompliant patients [61], consistent with the hypothesis that it is cotrimoxazole which is the critical component in the “conventional” combination therapy. Current trials in Thailand will hopefully ascertain whether it is still beneficial to have combination therapy for the eradication phase of melioidosis treatment or whether cotrimoxazole alone is adequate.

See table 3 for Summary of therapy for melioidosis.

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