



An informative intragenic microsatellite marker suggests the IL-1 receptor as a genetic modifier in cystic fibrosis

To the Editor:

Recent studies in mice with cystic fibrosis (CF)-like lung disease identified interleukin (IL)-1 receptor (IL-1R) signalling as an important pathway triggering neutrophilic airway inflammation that constitutes a key risk factor in the onset and progression of lung disease in patients with CF [1–4]. These studies demonstrated that CF-like airway mucus obstruction causes epithelial hypoxia and necrosis, which in turn leads to the release of IL-1 α from dying cells and activation of IL-1R signalling triggering neutrophilic inflammation and structural lung damage *in vivo* [1, 5]. Further, necrotic epithelial cells were detected in mucus-obstructed airways in lung sections from patients with CF [1]. Hypoxic cell death is a well-established trigger of sterile neutrophilic inflammation in many other organs and previous studies have identified IL-1R signalling as a key pathway required for triggering this inflammatory response to dying cells [6]. In addition, the gene encoding the IL-1R ligand IL-1 β that is induced by bacterial infection has been identified as a genetic modifier of CF by independent North American and European CF modifier studies [7, 8]. Collectively, these studies suggest that IL-1R signalling may play an important role in the pathogenesis of neutrophilic inflammation that is invariably detected in the airways of patients with CF, in the absence and presence of bacterial infection [2, 3]. However, the role of IL-1R and its association with disease severity in patients with CF remains unknown.

The aim of this study was therefore to assess the potential role of the *IL1R* gene, encoding a central molecule in the IL-1R signalling pathway, as a modifier of disease severity in patients with CF. To achieve this goal, we genotyped the informative microsatellite marker IL1RSat located within intron 1 of *IL1R* in several independent CF patient cohorts and a population control derived from the PopGen Biobank (figure 1). We amplified the (AAT) n repeat motif IL1RSat characterised by high polymorphism information content, which is a prerequisite for a marker to capture functional genetic variation at linked sites (figure 1a). We considered IL1RSat to be a neutral genetic marker and used it as a surrogate marker for other variants in its vicinity that are co-inherited together with IL1RSat because of linkage disequilibrium. We genotyped three independent CF patient cohorts at IL1RSat: CF patients of the European Twin and Sibling Study (EUCFTSib) who were born between 1957 and 1990 (mean year of birth 1979) and recruited in 1996 whereby only pairs of two siblings old enough to perform lung function measurements were accepted [8], and patients from the CF centres in Munich (<1970M) and Hannover (<1970H) who were born before 1970 and survived at least until 1990. The EUCFTSib population contains 39 patient pairs that were selected from a total of 318 pairs using the following strategy: anthropometry and lung function data were used to rank all individuals according to their disease severity [8]. These selected pairs represent both mild and severe CF disease, whereby the individuals' phenotypes are described by clinical parameters for growth and lung function and all selected patients from EUCFTSib rank below the 25th centile (severe) or above the 75th centile (mild) for both parameters [8]. Pairs were included into the analysis of CF-modifying genes if 1) they were F508del homozygous in order to minimise the effect of the *CFTR* genotype on phenotype variability, thus emphasising the influence of non-*CFTR* genes; and 2) if the pair's phenotype was either extremely concordant (concordant mildly affected, 13 pairs; concordant severely affected, 12 pairs) or discordant (14 pairs) [8]. As a higher impact of inherited factors on phenotype manifestation can be expected when studying extreme phenotypes [11, 12], this strategy leads to an enrichment of functional variants in CF-modifying genes in EUCFTSib.



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IL1R modifies disease severity in CF patients, emphasizing the significance of IL-1 signalling for CF pathogenesis <http://ow.ly/pIN730gL10G>

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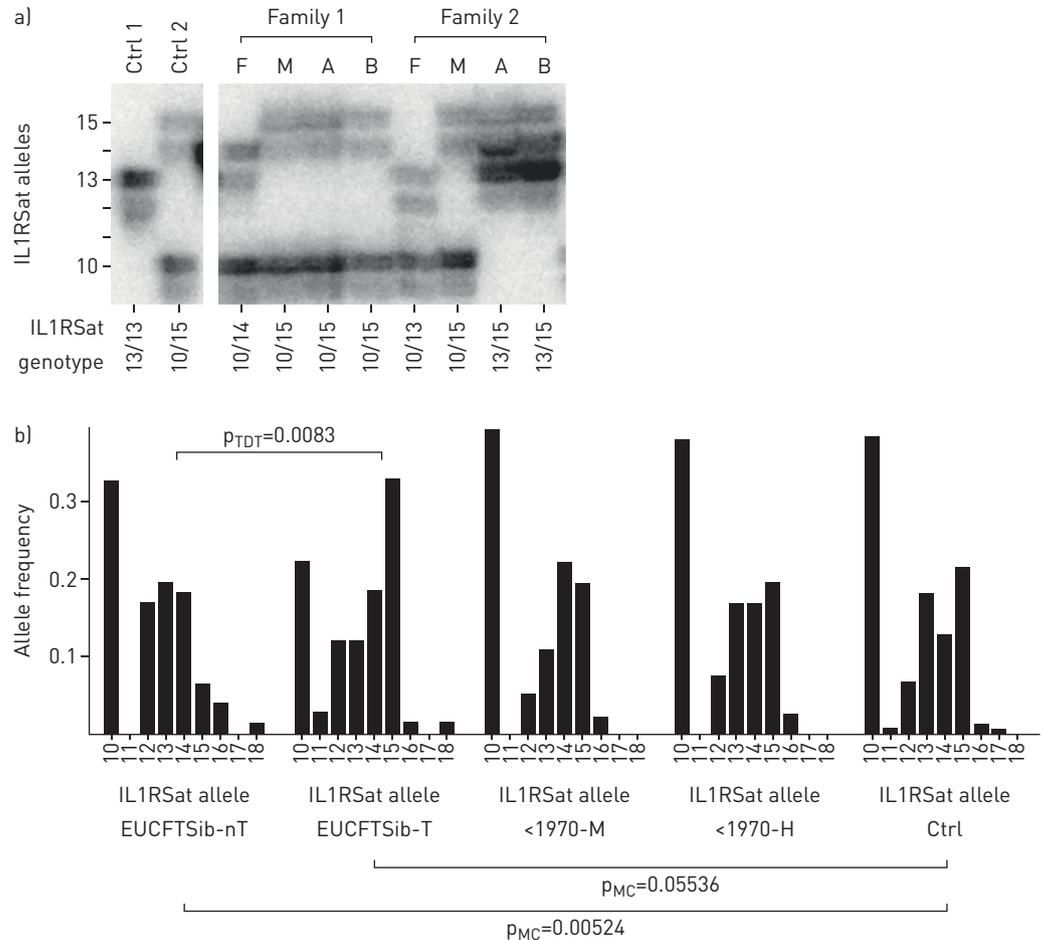


FIGURE 1 An informative microsatellite marker within intron 1 of the interleukin 1 receptor (IL1R) gene is associated with survival in patients with cystic fibrosis. This microsatellite IL1RSat was selected for genotyping from a set of eight microsatellite motifs, all located within intron 1 of IL1R that were tested on five unrelated control samples. IL1RSat is located within intron 1 of IL1R, starting at nucleotide 102725407 of the chromosome 2 reference sequence GRCh37.p13, with primers (5'-tcaactctgactcccagctca-3' and 5'-gcggtacctaaaccaggtc-3') surrounding an (AAT) n repeat motif. We observed signals that vary in size by three nucleotides, describing a total of nine IL1RSat alleles, indicating that the number of AAT repeat motifs differs between individuals in a population and is suitable to reflect at least partially the genetic background of the IL1R gene. a) IL1RSat genotyping primary data. Two control samples were used on all analyses to identify the allele by size. Assignment of the IL1RSat genotype to observed signals is shown for two families from the European CF Twin and Sibling (EUCFTSib) study. For example, in family 2, alleles transmitted from parents to offspring are 13 (paternal) and 15 (maternal) while non-transmitted alleles are 10 (paternal) and 10 (maternal). b) IL1RSat allele distributions among cystic fibrosis (CF) families and patients. The interleukin 1 receptor (IL1R) gene was interrogated as a genetic modifier of CF by genotyping of the informative microsatellite marker IL1RSat within intron 1 of IL1R in F508del-CFTR homozygous patients from EUCFTSib (39 families with 78 patients), from the CF centre Munich (<1970-M, 18 unrelated patients) and from the CF centre in Hannover (<1970-H, 39 unrelated patients). Patients of these independent cohorts had survived at least until school age (affected patient pairs of EUCFTSib; mean age at day of recruitment 17 years, range 6–39 years) or adulthood (unrelated patients born prior to 1970 who have survived until recruitment for DNA analysis in 1990 or later; <1970-M and <1970-H; mean age in 1990, 24 years, range 21–32 years) and a population control from Schleswig-Holstein in Germany (Ctrl, 94 individuals). Distributions of IL1RSat allele frequencies are shown for transmitted (EUCFTSib-T) and non-transmitted (EUCFTSib-nT) alleles in families of the European CF Twin and Sibling Study and for independent patient cohorts followed at the CF centres in Munich (<1970-M) and Hannover (<1970-H). Genetic data of CF patients and their parents in the EUCFTSib were analysed with the software package FAMHAP, which corrects for sibpair dependency in association and linkage studies (p_{TDT}) [9]. Allele distributions between cohorts of unrelated patients and controls were directly compared using Monte Carlo simulation with CLUMP (p_{MC}) [10].

Patients from the other two CF cohorts were F508del-CFTR homozygous but not selected for any disease manifestation to cover the entire spectrum of disease severity among the survivors. For unrelated patients, epidemiological data on CF birth cohorts provide an estimate on how mortality rates influence the proportion of patients that could be recruited in our study. An analysis of survival of CF patients in the

UK during the time period 1977 to 1985 reported a 50% survival at 19 years of age [13]. These UK survivors showed a birth year range similar to our two German cohorts of unrelated patients, indicating that we would have included only ~50% of the CF population if we had recruited those survivors in 1977–1985 and that even fewer would have been enrolled in 1990 or later. There is no corresponding epidemiologic data available that describe the survival among CF sibling pairs, but the chance to recruit two life siblings simultaneously from a birth cohort that exhibits significant mortality [13, 14] is even lower (and thus the survivor effect higher) than the probability to recruit unrelated patients from the same birth cohort. We therefore expect that the inclusion criterion “two surviving siblings” contributed to a further enrichment of benign alleles in the EUCFTSib population compared with individual patients.

We first used the transmission disequilibrium test (TDT), originally developed by SPIELMAN *et al.* [15] in their discovery of the diabetes-causing locus, to compare the distribution of IL1RSat allele frequencies between the inherited genetic information transmitted to the CF patients (transmitted alleles) *versus* the information that was retained in the parental generation (non-transmitted alleles) from CF families of EUCFTSib. We found a transmission disequilibrium with allele 15 at IL1RSat over-represented in the recruited CF siblings ($p_{\text{TDT}}=0.0083$), annotating the IL1RSat allele 15 as a benign variant associated with survival of CF patient until enrolment into the study (mean age 17 years, range 6–39 years). The allele distribution in the control population (Ctrl) differed from the IL1RSat allele distribution on the transmitted alleles (*i.e.* surviving CF patients) ($p_{\text{MC}}=0.055$), as well as the non-transmitted alleles ($p_{\text{MC}}=0.00524$) of the parents from EUCFTSib (figure 1). In independent cohorts of CF patients that were born before 1970 and survived until at least 1990, besides the ubiquitous allele 10 that accounted for a third of the chromosomes in all populations, IL1RSat alleles 14 (<1970M) and 15 (<1970H) were the most frequently observed alleles (figure 1), albeit this trend towards a similar pattern of enrichment of allele 15 at IL1RSat did not reach statistical significance ($p_{\text{best}}=0.08$) with the number of patients that were available for this study.

The TDT builds on the comparison of transmitted and non-transmitted alleles within nuclear families as an association study with internal controls enabling the identification of disease-causing genes [15]. For the monogenic disease CF that is caused by the *CFTR* gene, we previously showed that the TDT can also be used to detect the influence of additional genetic modifiers that have an impact on patient survival prior to enrolment in cross-sectional studies [8]. The distortion of allele frequencies because of a cross-sectional recruitment strategy in the CF patient population, for which an improvement in disease management and patient survival could be accomplished within the last decades, has been noticed before through an enrichment of mild pancreatic sufficient-associated *CFTR* mutations that account for 18% of *CFTR* alleles for patients born in 1966 or earlier who were recruited for *CFTR* mutation analysis in 1990 or later [16]. Moreover, a survivor effect has been detected through an enrichment of *TGFBI*-Leu10 in a F508del-*CFTR* homozygous CF patient population ($p=0.066$) [16] and by TDT among EUCFTSib at D12S889 in the *TNF α* receptor gene *TNFR1* ($p=0.0067$), in the gene encoding the γ -subunit of the amiloride-sensitive sodium channel *SCNN1G* ($p=0.0034$), and, by default, at the *CFTR* locus ($p=10^{-6}$) as an indication of the CF disease-causing gene itself [8].

In summary, we have assessed whether a highly polymorphic microsatellite in intron 1 of the *IL1R* gene shows enrichment of a specific allele in patients with CF who have survived to take part in the study. Our finding that the informative microsatellite marker within intron 1 of *IL1R* detects a survival advantage for patients with CF, revealed through a transmission disequilibrium in patient cohorts (figure 1) with a survivor-dependent recruitment bias [13, 14], supports involvement of IL-1R in CF pathogenesis. This concept is in agreement with an important role of the IL-1R signalling pathway in the pathogenesis of neutrophilic inflammation, as well as findings from previous studies that identified *IL1B* encoding the IL-1R ligand IL-1 β as a modifier gene in CF [7, 8]. Earlier, LEVY *et al.* [7] described an association signal with lung disease at *IL1R* among female F508del-*CFTR* homozygous patients from the University of North Carolina and Case Western Reserve (UNC/CWRU) Cohort. Taken together, these studies draw attention to the role of the IL-1R signalling pathway, either activated by IL-1 α released from hypoxic cells in mucus-obstructed airways or by IL-1 β induced by bacterial infection [1, 5, 6, 17], in the pathogenesis of chronic neutrophilic inflammation causing progressive lung damage in patients with CF [1–4]. These findings from genetic and functional studies also indicate that inhibition of IL-1R signalling may be a promising anti-inflammatory strategy in CF and potentially other lung diseases associated with chronic airway mucus obstruction and neutrophilic inflammation.

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References

- 1 Fritzsche B, Zhou-Suckow Z, Trojanek JB, *et al*. Hypoxic epithelial necrosis triggers neutrophilic inflammation via IL-1 receptor signaling in cystic fibrosis lung disease. *Am J Respir Crit Care Med* 2015; 191: 902–913.
- 2 Sly PD, Gangell CL, Chen L, *et al*. Risk factors for bronchiectasis in children with cystic fibrosis. *N Engl J Med* 2013; 368: 1963–1970.
- 3 Sagel SD, Wagner BD, Anthony MM, *et al*. Sputum biomarkers of inflammation and lung function decline in children with cystic fibrosis. *Am J Respir Crit Care Med* 2012; 186: 857–865.
- 4 Montgomery ST, Mall MA, Kicic A, *et al*. Hypoxia and sterile inflammation in cystic fibrosis airways: mechanisms and potential therapies. *Eur Respir J* 2017; 49: 1600903.
- 5 Mall MA, Harkema JR, Trojanek JB, *et al*. Development of chronic bronchitis and emphysema in β -epithelial Na⁺ channel-overexpressing mice. *Am J Respir Crit Care Med* 2008; 177: 730–742.
- 6 Chen CJ, Kono H, Golenbock D, *et al*. Identification of a key pathway required for the sterile inflammatory response triggered by dying cells. *Nat Med* 2007; 13: 851–856.
- 7 Levy H, Murphy A, Zou F, *et al*. IL1B polymorphisms modulate cystic fibrosis lung disease. *Pediatr Pulmonol* 2009; 44: 580–593.
- 8 Stanke F, Becker T, Kumar V, *et al*. Genes that determine immunology and inflammation modify the basic defect of impaired ion conductance in cystic fibrosis epithelia. *J Med Genet* 2011; 48: 24–31.
- 9 Herold C, Becker T. Genetic association analysis with FAMHAP: a major program update. *Bioinformatics* 2009; 25: 134–136.
- 10 Sham PC, Curtis D. Monte Carlo tests for associations between disease and alleles at highly polymorphic loci. *Ann Hum Genet* 1995; 59: 97–105.
- 11 Risch N, Zhang H. Extreme discordant sib pairs for mapping quantitative trait loci in humans. *Science* 1995; 268: 1584–1589.
- 12 Dolan CV, Boomsma DI. Optimal selection of sib pairs from random samples for linkage analysis of a QTL using the EDAC test. *Behav Genet* 1998; 28: 197–206.
- 13 British Paediatric Association Working Party on Cystic Fibrosis. Cystic fibrosis in the United Kingdom 1977–85: an improving picture. *BMJ* 1988; 297: 1599–1602.
- 14 Elborn JS, Shale DJ, Britton JR. Cystic fibrosis: current survival and population estimates to the year 2000. *Thorax* 1991; 46: 881–885.
- 15 Spielman RS, McGinnis RE, Ewens WJ. Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM). *Am J Hum Genet* 1993; 52: 506–516.
- 16 Becker T, Jansen S, Tamm S, *et al*. Transmission ratio distortion and maternal effects confound the analysis of modulators of cystic fibrosis disease severity on 19q13. *Eur J Hum Genet* 2007; 15: 774–778.
- 17 Dinarello CA, Simon A, van der Meer JW. Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. *Nat Rev Drug Discov* 2012; 11: 633–652.

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