

**SERIES 'OCCUPATIONAL ASTHMA'**  
*Edited by C. Mapp*

## Aetiological agents in occupational asthma

M. Chan-Yeung\*, J-L. Malo\*\*

*Aetiological agents in occupational asthma. M. Chan-Yeung, J-L. Malo. ©ERS Journals Ltd 1994.*

**ABSTRACT:** Occupational asthma has become the most prevalent occupational lung disease in developed countries. At present, about 200 agents have been implicated in causing occupational asthma in the workplace. These agents can be divided into two categories by their mechanism of action: immunological and nonimmunological.

Immunological causes can be further divided into those that induce asthma through an immunoglobulin E (IgE)-dependent mechanism, and those that induce asthma through a non-IgE-dependent mechanism. In the latter category, specific IgE antibodies are found only in a small percentage of the patients with proven disease, even though the clinical picture is compatible with an allergic reaction. The immunological mechanism(s) responsible for these agents has yet to be identified.

The best known example of nonimmunological asthma is Reactive Airways Dysfunction Syndrome (RADS) or irritant-induced asthma.

In this review, examples of types of agents causing occupational asthma are discussed and a compendium table of aetiological agents is given.

*Eur Respir J., 1994, 7, 346-371.*

\*Dept of Medicine, Vancouver General Hospital, Vancouver, Canada. \*\*Dept of Chest Medicine, Hôpital du Sacré-Coeur, Montreal, Canada.

Correspondence: J-L. Malo, Dept of Chest Medicine, Hôpital du Sacré-Coeur, 5400 West Gouin, Montreal, H4J ICS, Canada.

Keywords: Asthma  
occupational asthma

Received: July 16 1993

JL Malo and M Chan-Yeung are members of the Canadian Network of Excellence in Respiratory Health.

The prevalence of occupational asthma has increased over the past two decades [1, 2]. This is due partly to a better recognition of the condition, and partly to an increase in the number of new chemicals capable of causing occupational asthma in the workplace. At present, nearly 200 agents have been implicated in causing occupational asthma in the workplace [3].

### Definition of occupational asthma

In this article, occupational asthma is defined as recommended by BERNSTEIN *et al.* [4]. It is a disease characterized by variable airflow limitation and/or nonspecific bronchial hyperresponsiveness (NSBH) due to causes and conditions which are attributable to a particular occupational environment and not to stimuli encountered outside the workplace. Occupational asthma may encompass both immunological and nonimmunological causes 1) immunological occupational asthma occurs upon exposure to an agent after a latent period of immune sensitization; 2) nonimmunological asthma may or may not occur after a latency period of exposure to an agent(s) which does not induce immune sensitization as determined by currently available technology

Under certain exposure conditions, immunological and nonimmunological asthma may be concurrent.

### Aetiological agents in occupational asthma

The aetiological causes of occupational asthma can be classified into immunological and nonimmunological. Immunological causes can be further divided into those

that induce asthma through an immunoglobulin E (IgE)-dependent mechanism, and those that induce asthma through a non-IgE-dependent mechanism. In the latter category, specific IgE antibodies are found in only a small percentage of the patients with proven disease, even though the clinical picture is compatible with an allergic reaction. The immunological mechanism has yet to be identified. As it is not possible to cover all aetiological causes in this article because of limited space, only the most commonly encountered and most studied causes will be discussed.

### Agents causing asthma through immunological mechanisms

Occupational asthma due to immunological causes has the following characteristics that distinguish it from occupational asthma due to nonimmunological causes: 1) there is a latent period between the onset of exposure and the onset of respiratory symptoms; and 2) in sensitized subjects, re-exposure to a small amount of the causative agent leads to the occurrence of asthma.

### IgE-dependent causes

It has been demonstrated for many years that agents inducing asthma through an IgE-dependent mechanism are high molecular weight proteins or polysaccharides. Recently, several low molecular weight compounds have been shown to produce specific IgE antibodies, by

combining with a protein to form a hapten-protein conjugate [5]. There is good evidence to suggest that specific IgE antibodies to the hapten-protein conjugate are responsible for the patients' symptoms. Occupational asthma due to IgE-dependent aetiological agents mostly affects atopic subjects. In sensitized subjects with symptoms, specific inhalation challenge tests with extracts of these agents induced isolated immediate asthmatic reaction or a biphasic asthmatic reaction, but rarely an isolated late asthmatic reaction. Smoking has been shown by some to be an important determinant of some types of occupational asthma but not others [6]. Table 1 in the Appendix shows some of the IgE-dependent causes of occupational asthma, according to industries, jobs or work processes where the exposure can be found.

### High Molecular weight compounds (table)

#### *Animal products*

*Laboratory animal allergens.* Exposure to laboratory animals occurs in the pharmaceutical industry, in university and research units, and in animal breeding facilities. Small animals are often used in these places. The most common cause of asthma is rats, followed by mice, and rabbits. The allergens are usually found in the excreta and the secretions of these animals [7]. In rats and mice, the major source of allergen is in the alpha<sub>2</sub>-globulin and/or the prealbumin portion of the protein in urine [7]. The allergenic protein appears in the urine of male rats after puberty and disappears with senility. In guinea-pigs, allergens have been identified in the urine, serum, saliva and pelt. Again the allergenic activity is highest in the prealbumin region. The nature and the sources of rabbit allergens have been less extensively studied.

The prevalence of asthma due to laboratory animal allergy is not known. In the United Kingdom, the Surveillance of Work-Related and Occupational Respiratory Disease (SWORD) project group [1] reported an estimated incidence of 204 cases of occupational asthma per million animal handlers per year. About one third of the animal handlers have allergic symptoms, such as allergic rhinitis and contact urticaria; whilst 10% have symptoms of asthma. The results of a longitudinal study of workers in the pharmaceutical industry were reported by BOTHAM *et al.* [8]. The prevalence of specific IgE antibodies to rat urine proteins was 28% during the first year of employment, 37% in the subsequent two years, but fell to around 10% in the fifth to seventh year [8]. The improvement was attributed to improved work practice and the use of personal protection rather than to selection.

Clinically, allergic symptoms develop within the first 2–3 yrs of exposure. A recent study of patients with different types of occupational asthma showed that the latency period between the onset of exposure and the onset of symptoms is longer for patients with asthma due to high molecular weight compounds than for those with asthma due to low molecular weight compounds [9].

Asthma usually occurs several months after the development of other symptoms. Atopic animal handlers are more prone to develop asthma [10].

The diagnosis of occupational asthma in laboratory animal handlers is based on the presence of a compatible history, with objective evidence of work relationship and evidence of sensitization. Objective evidence of work relationship can be obtained by peak expiratory flow rate (PEFR) monitoring for a period at work and a period off-work, or by a specific inhalation challenge test with appropriate extracts if available. Evidence of sensitization can also be obtained by a positive skin test or presence of specific IgE antibodies with the appropriate extracts.

One of the important technological advances in occupational asthma is the ability to measure the amount of airborne allergens [11]. This enabled investigators to study factors that determine the levels of allergens in animal laboratories and methods of reducing these allergens [12–14]. The latter includes the reduction of stock density, the use of female rats rather than male rats, the use of corn cob rather than wood shaving as bedding, and covering of the cages with filter caps [13].

*Other animal-derived allergens.* Other animal-derived proteins have been incriminated as causing occupational asthma. Asthma caused by exposure to pig-derived allergens in a meat factory has been described [15]. Handling of bovine serum albumin powder caused occupational asthma in a laboratory technician [16]. Exposure to egg proteins has been reported to cause occupational asthma [17]. In a survey of 188 workers employed in a factory that produced liquid and powdered egg products, 7% were found to have physician-diagnosed asthma and evidence of sensitization to one or more egg proteins [18].

Different types of insects can give rise to occupational asthma. Storage mites have been incriminated as allergens for occupational asthma among farmers [19, 20]. Other types of mites such as spider mites and poultry mites have been found to be the cause of occupational asthma in greenhouse workers and poultry workers, respectively, [21, 22]. Laboratory workers breeding insects such as grain weevil [23], locust [24, 25] and fruit fly [26], are at risk of developing occupational asthma.

#### *Plant proteins of polysaccharides*

*Flour.* In some countries, baker's asthma is the most prevalent type of occupational asthma [27]. The majority of patients with baker's asthma are allergic to wheat and rye flour. There is considerable cross-reactivity between antigens in wheat and rye flour and those in other cereals, such as barley and triticale, but less to oats, corn and rice [28]. Crossed immunoelectrophoresis studies of wheat extract identified 40 different allergens [29]. However, the major allergens are found in globulin, gliadin and glutenin fractions [30].

Several other non-cereal antigens have been identified as capable of causing asthma in bakers. Sensitization to

Aspergillus and Alternaria has been implicated in two bakers with asthma [31]. Other food additives such as alpha-amylase, hemicellulase and papain, used as enzymes in bakeries, can also cause sensitization [32, 33].

It has been estimated that about 10–20% of bakers suffer from baker's asthma [27]. A cross-sectional study of 318 bakers showed that 13% had respiratory work-related symptoms [34]. The degree of nonspecific bronchial hyperresponsiveness (NSBH), lung function level, and skin test reactivity to antigens tested correlated with the current level of flour dust exposure and the estimated total exposure.

The onset of asthma in bakers is usually preceded by nasal symptoms, such as sneezing, rhinorrhoea and itchy eyes. The latent period between the onset of exposure and the onset of respiratory symptoms varies between a few months and a few years [35]. Almost all patients are atopic subjects with skin reactivity to common allergens. The diagnosis of baker's asthma is usually based on a compatible clinical history, presence of airflow limitation and/or presence of NSBH, or the demonstration of positive skin test to the appropriate flour extract. Positive bronchial reactions to flour extract occurred in symptomatic workers in association with high specific IgE level and a higher degree of NSBH [36].

Workers exposed to grain dust such as grain elevator terminal workers and farmers do not, as a rule, become sensitized to cereal flours but to other components in the grain dust. For example, occupational asthma in grain farmers is usually related to sensitization to storage mites such as *Glycyphagus destructor* [37].

*Pharmaceutical products.* Some pharmaceutical products are proteins, such as ipecacuanha, derived from the roots of *Cephaelis ipecacuanha*, or polysaccharides, such as psyllium.

Occupational asthma due to psyllium has been studied more extensively than other proteinaceous pharmaceutical products [38]. The clinical picture of psyllium-induced asthma is well-documented in several case reports [39–44]. In addition to asthma, many patients also complained of ocular and nasal symptoms [45]. Specific IgE antibodies to psyllium have been found in these patients. The prevalence of sensitization among pharmaceutical workers was between 28% [46] and 44% [47]; whilst the prevalence of occupational asthma confirmed by specific challenge test was 4% [46]. Among nurses working in long-term care facilities, the prevalence of positive skin test was lower (5%), but the prevalence of confirmed occupational asthma by specific challenge test was also 4% [38].

*Gums.* Gums are derived from plants and are high polymer carbohydrates that produce mucilages when they react with water. Common examples include acacia or arabic gum, used in the food and pharmaceutical industries; tragacanth, used mainly in printing; karaya, which grows in India, and can be used instead of tragacanth. Guar gum, another natural gum, was reported to be responsible for asthma in the carpet-manufacturing industry [48]. Occupational asthma due to acacia gum was

described as early as 1933 [49]. Later, cases of asthma among printers exposed to acacia gums were documented [50]. Five percent of workers in a carpet-manufacturing factory was found to have positive skin tests to guar gum, while 2% had occupational asthma diagnosed by specific challenge tests [48].

### Enzymes

*Detergent enzymes.* Proteolytic enzymes from *Bacillus subtilis* were introduced in detergent products in the 1960s. Within a few years of their introduction, several investigators reported the association of exposure to detergent enzymes with asthma, nasal and ocular symptoms among workers involved in the manufacture of detergents [51, 52]. These workers showed positive skin test reactivity to *B. subtilis* alcalase and developed an immediate or late asthmatic reaction on inhalation challenge [53]. A fall in diffusing capacity was found in some workers following inhalation challenge test, suggesting involvement of lung parenchyma as well as airways [52]. An epidemiological study of detergent manufacturing workers demonstrated that 21% had skin sensitivity to alcalase; of these, 66% were atopic subjects [54]. Reports of sensitization in consumers followed later. Four percent of 461 individuals attending an allergy clinic with no known occupational exposure to detergent enzymes had positive skin test to alcalase [53]. Positive nasal or bronchial reactions to challenge with the enzyme were documented in some of the consumers [53].

Since these reports, measures were taken to reduce exposure not only for workers but also consumers. Ventilation was improved in factories, and changes were made in manufacturing processes, such as enclosing the purification of enzymes. These enzymes were also glued to phosphate granules to prevent them from getting dispersed, or were encapsulated. Despite the substantial reduction in levels, a study showed that sensitization still occurred in 25% of workers [55]. However, in a study of the general population, no sensitization to alcalase was found, indicating that the use of enzymes in granules and encapsulation had been successful, at least in eradicating the disease among consumers [56].

*Other enzymes.* Both plant-derived enzymes, such as papain from papaya and bromelin from pineapple, and animal-derived enzymes, such as hog trypsin, are frequently used in the food industries. Some of them have been found to cause occupational asthma [57].

### Fish and seafood protein

Processing of several types of seafood can cause occupational asthma. Oyster handlers were found to be sensitized to the "hoya" or "sea-squirt" contaminating oysters, and developed symptoms of rhinitis and asthma [58]. Oysters themselves were not the source of allergen. Processing of prawns gave rise to asthma in 18 out of 50 workers [59]. Inhalation challenge test with the

prawn extract on two volunteers induced isolated late asthmatic reaction in one and dual asthmatic reaction in the other [59].

Occupational asthma associated with snow crab processing has been studied in detail [60]. During the process of boiling, a vapour is released from the snow crab. Inhalation of the vapour containing proteins from snow crabs resulted in sensitization of workers. In a study of two snow crab processing plants along the St. Lawrence River with 313 employees, 33 were confirmed to have occupational asthma by specific challenge tests [60].

### Low molecular weight compounds (table 1)

Some low molecular weight compounds have been shown to cause occupational asthma associated with the production of specific IgE antibodies against the hapten-protein conjugates. The most well known compounds in this group include acid anhydrides, metals such as platinum and some pharmaceutical products.

#### *Acid anhydrides*

There are several acid anhydrides: phthalic anhydride, trimellitic anhydride, hexahydrophthalic anhydride, himic anhydride and tetrachlorophthalic anhydride. These anhydrides are used in alkyd and epoxy resins. Alkyd resins form the base for paints, varnishes and plastics. Epoxy resins are widely-used in adhesives, casting, coating, and sealants. Trimellitic anhydrides are mainly used as plasticizers for polyvinyl chloride, especially when temperature stability is required. Acid anhydrides can give rise to hypersensitivity reactions in exposed population. They act as haptens and combine with proteins to form conjugates with antibodies recognizing the haptens [61]. They can also combine with self-proteins to generate new, carrier-dependent antigenic determinants with antibody combining sites being directed against a conformational change in the self-proteins [61].

*Phthalic anhydride.* Since 1939, phthalic anhydride has been known to cause asthma and rhinitis [62]. Specific IgE antibodies to phthalic anhydride conjugated to human serum albumin have been demonstrated [63].

*Trimellitic anhydride.* Trimellitic anhydride induced not only specific immunoglobulin E, A, M and G (IgE, IgA, IgM and IgG) antibodies against the hapten but also to the new antigenic determinants that arise when trimellitic anhydride couples with proteins [61]. Trimellitic anhydride has been shown to cause a spectrum of lung diseases: asthma and rhinitis, late respiratory systemic syndrome and pulmonary disease-anaemia syndrome [64]. Whilst IgE antibodies are probably responsible for asthma, connections have been made between other antibodies and late respiratory systemic syndrome and pulmonary disease-anaemia syndrome [65]. A study was conducted on 474 employees exposed to trimellitic anhy-

dride. Evidence of trimellitic anhydride immunological lung disease was observed in 6.8% [66]. A direct relationship was found between the levels of total and IgE antibodies and the degree of exposure.

*Tetrachlorophthalate anhydride.* In a survey of 276 workers exposed to tetrachlorophthalate anhydride (TCPA), specific IgE antibodies to the TCPA-conjugate were found in 12% of atopic subjects and 6% of nonatopic subjects. There was a strong interaction between smoking and atopy; smokers who were atopic subjects had the highest prevalence of positive specific IgE antibodies [67]. A follow-up study of workers with TCPA-induced asthma showed a progressive reduction in levels of specific IgE antibodies after removal from exposure. The half-life of these antibodies was one year [68]. Various animals: rats, Rhesus monkeys, dogs, rabbits and guinea-pigs have been used, without success, to induce a model of IgE-mediated airway disease to anhydrides [61]. Although other types of hypersensitivity reactions were induced, airway hyperresponsiveness was not found.

#### *Metals*

In addition to asthma, inhalation of metallic compounds can give rise to chemical pneumonitis, bronchitis and metal fume fever. Chronic exposure has been associated with chronic bronchitis, emphysema and, in the case of hard metals, pneumoconiosis.

Exposure to metals may occur not only in metal mining and metallurgical industries, but in a number of other trades, such as welding and soldering. Chromates are found in paints, chemical and plastic industries, while cobalt is found in diamond polishing discs [69]. Metals that can induce asthma can be divided into three categories: 1) transition metals, such as vanadium, chromium, nickel and zinc; 2) precious metals, such as platinum and palladium; and 3) hard metals, such as tungsten carbide and cobalt.

*Nickel.* Exposure to nickel occurs during mining, milling, smelting, and refining processes, as well as in electroplating industries and in the production of nickel catalysts. The occurrence of nickel-induced asthma among exposed workers is rare compared to contact dermatitis. In the literature, there have been several case reports of nickel-induced asthma associated with exposure to nickel sulphate; some confirmed by inhalation challenge tests [70, 71]. Specific IgE antibodies to nickel-human serum albumin conjugate have been reported in some cases [70]. Recently, lymphocyte transformation was demonstrated in patients with nickel-induced asthma, suggesting that cell-mediated hypersensitivity may play a part in nickel-induced asthma [72].

*Chromium.* Chromium metal is nonallergenic. Chromium salts are well-known to give rise to contact dermatitis. Exposure to chromium salts occurs in electroplating, tanning, pigment and cement production. The development of asthma is frequently preceded by a history of

contact dermatitis. As in nickel-induced asthma, cell-mediated hypersensitivity may be important.

**Platinum.** Platinum salts, particularly halides, are more potent in inducing asthma than other metallic salts. Exposure to platinum occurs in mining, in primary and secondary refining processes, and in chemical industries where platinum is used as a catalyst. During the refining process, hydrochloric acid and chlorine are added. It is the halide salts that give platinum its allergenicity. Platinum-induced asthma has been recognized for a long time [73, 74]. Positive skin tests with the appropriate platinum salts (unconjugated) were found in sensitized workers [75]. Specific IgE antibodies were detected by radio allergosorbent test (RAST) procedures [76]. A Type I allergic reaction is likely to be responsible for the pathogenesis of platinum salt-induced asthma.

In a recent cross-sectional study of a secondary refinery, 14% of the 107 current employees and 28% of the 29 workers laid-off for medical reasons had positive skin test reactivity to platinum salts, suggesting that the current employees were "healthier". In this study, the prevalence of atopy was similar among those with positive and negative skin tests to platinum salts, and those of the general population, suggesting that atopy is not an important determinant. However, smoking was found to be strongly associated with sensitization to platinum salts [77].

**Zinc.** Exposure to zinc occurs in the steel manufacturing industry, where it is used as an alloy. It is also used as a component of brass, and in the insecticide industry. The most important exposure to zinc occurs in welding of galvanized iron. Exposure to fumes of zinc oxide is associated with metal fume fever, which is a flu-like illness, sometimes associated with cough and shortness of breath. Several cases of occupational asthma have been described related to exposure to fumes of zinc oxide [78, 79].

#### Other

There are other low molecular weight compounds that can act as haptens and cause asthma by inducing specific IgE antibodies. It is not possible to discuss all of them here.

#### Non-IgE-dependent causes

The majority of low molecular weight compounds induce asthma by mechanisms as yet unidentified, as specific IgE antibodies to the offending agents have not been found, or found in only a small proportion of patients with the disease. The clinical picture of these patients

is compatible with that of an allergic disease. The accompanying table shows some of these causes. Specific challenge tests with these agents commonly induce late asthmatic reactions, either as an isolated asthmatic reaction or as the late component of biphasic reactions, and in some cases reactions that are atypical [80]. The majority of affected subjects are nonatopic and nonsmokers [81]. Thus, these non-IgE-dependent agents are different from other types of low molecular weight compounds that induce specific IgE antibody production. Asthma due to diisocyanates and asthma due to the Western red cedar are examples of non-IgE-dependent causes that have been studied in detail and will be discussed here.

#### Diisocyanates

Occupational asthma due to diisocyanates has been reviewed in detail by VANDENPLAS *et al.* [82]. Diisocyanates are being used extensively in a number of important industries all over the world. They are a group of small molecular weight cyclic or linear organic chemical compounds, synthesized by the reaction of amines or their hydrochlorides with phosgene. These chemicals all have an  $-N=C=O$  group attached to a radical, and react exothermically with compounds containing active hydrogen atoms, such as polyglycols, to form a polymeric mass, polyurethane (fig. 1). Monomers of isocyanates can exist in aromatic, aliphatic or cycloaliphatic forms (fig. 2). Prepolymers of isocyanates have also been shown to give rise to occupational asthma [82]. Toluene diisocyanate (TDI) was first developed during the second World War and is the most commonly used diisocyanate. Since that time, other isocyanates have been developed, such as methylene diphenyl diisocyanate (MDI), hexamethylene diisocyanate (HDI), naphthylene diisocyanate (NDI), and isophorone diisocyanate (IPDI). They are used in the production of rigid and flexible foams, as protective coatings for electric wiring, for painting automobiles, as adhesives and binders in foundries, as liners for mine and grain elevator chutes, soles of shoes and spandex fibres. Thus, they have a wide application in both industrial and domestic use.

**Toluene diisocyanate.** TDI-induced asthma has been the most common cause of occupational asthma in many parts of the world for many years. However, in recent years, it has been replaced by HDI and MDI [82]. The prevalence of TDI-induced asthma varies from 5 to 10%, depending on the type of industry, the type of diisocyanate, and the level of exposure [82]. Many cases of TDI-induced asthma occurred after a high level of exposure, *e.g.*, a spill [83]. In those situations, a Reactive Airways

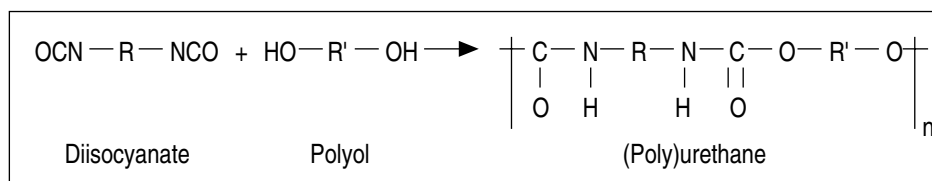


Fig. 1.  $-N=C=O$  group attached to a radical, reacts with compounds containing active hydrogen atoms, such as polyglycols, to form a polymeric mass, polyurethane.

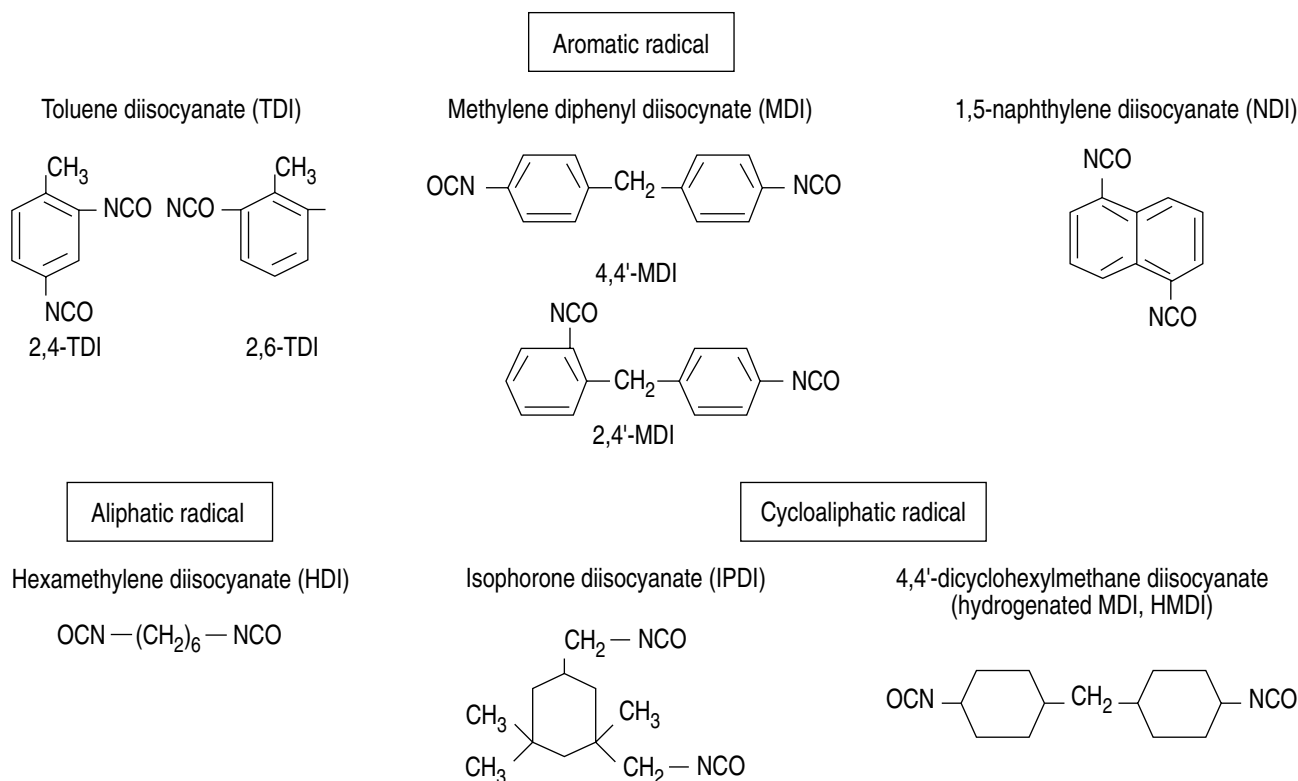


Fig. 2. — Structure of different monomers of isocyanates.

Dysfunction Syndrome (RADS)-type of occupational asthma could not be excluded. Recently, there has been a report that certain genetic factors related to human leucocyte antigen (HLA) Class II may convey protection, whilst others convey susceptibility [84]. Obviously, a great deal of work needs to be done in this area.

The dose and length of exposure to TDI which will induce asthma are not known. Recommendations from the National Institute for Occupational Safety and Health (NIOSH) are that humans should not be exposed to concentrations above 20 ppb [85]. Several animal models have been developed to study TDI hypersensitivity reaction. In guinea pigs, specific IgG<sub>1</sub> and not IgE antibodies were found; in another study, specific IgE antibodies were detected [86, 87]. Irrespective of whether antibodies were present or not, the animals developed airflow obstruction and NSBH. The uptake and distribution of TDI have also been studied in guinea-pigs [88, 89]. Most of the TDI was absorbed in the upper airways; the uptake, as reflected by the concentration of <sup>14</sup>C in the blood, correlated with the concentration in inhaled air. In the airways, labelled TDI was found in the epithelium and subepithelium from the nose to terminal bronchioles but not in the alveoli [88, 89]. In the blood, the labelled TDI was found in the plasma, conjugated to a 70,000 kD a molecular weight protein. In the airways, TDI was also found conjugated with a 70,000 kD a molecular weight protein, which was shown to be laminin. These studies may be helpful for the identification of the protein conjugated to isocyanates in humans.

Clinically, once a subject has been sensitized to TDI, very low levels of TDI (as low as 1 ppb) may trigger an attack of asthma. Specific challenge tests with TDI-induced isolated immediate, isolated late, biphasic or a

continuous reaction in sensitive subjects. Recurrent late asthmatic reaction has been reported after one single challenge to TDI [90]. The development of late asthmatic reaction has been shown to be accompanied by the development of NSBH in about 60% of subjects [80]. Once asthma has developed, the majority had persistence of symptoms even after they were removed from exposure [91–94]. Favourable prognostic factors include early diagnosis and early removal from exposure. The mechanism of TDI-induced asthma is not known. Specific IgE antibodies to a protein conjugate of TDI or a monoisocyanate were present in only a small proportion (10–20%) of patients who were shown to have specific asthmatic reactions on inhalation challenge test. Skin prick testing with hapten-protein conjugates has been negative [95]. There have been reports indicating that the immune response could be directed against neoantigens formed by the reaction of TDI with body proteins [96], as in other highly reactive low molecular weight compounds [97]. Specific lymphocyte blastogenic activity and lymphokine leucocyte inhibitory factor responses to TDI have been detected in sensitive workers, and suggest that cell-mediated hypersensitivity reaction may play a part [96]. TDI has several other effects that might induce NSBH or potentiate sensitizing properties. It has been shown to have beta-adrenergic blockade properties [98], to stimulate the release of neuropeptides and to inhibit neutral endopeptidases in *in vivo* and *in vitro* studies [99].

Irrespective of the mechanism of induction of asthma, airway inflammation was found in patients with TDI-induced asthma both by bronchoalveolar lavage (BAL) and bronchial biopsy studies [99]. The pathology of the lungs of a patient with isocyanate-induced asthma who died during an attack showed similar findings as those

who died from status asthmaticus [100]. The airways were plugged with mucus. There was sloughing of the epithelium and marked cellular infiltration of neutrophils and eosinophils. The basement membrane was thickened and the smooth muscle hypertrophied. The pathology of the airways of patients with TDI asthma was studied before and after exposure ceased. The thickening of the basement membrane observed when the patients were still exposed and symptomatic became less severe, suggesting this change was partly reversible [99].

#### *Methylene diphenyl diisocyanate (MDI)*

Methylene diphenyl diisocyanate is used in the production of rigid foam, and as a binder in almost all mould and core processes in iron and steel foundries. At room temperature, MDI is a solid. It has to be heated above 60°C before vapours are given off. The health effects of MDI exposure are less well-studied than TDI. Both occupational asthma and hypersensitivity pneumonitis and a combination of both have been described [82]. The prevalence of these diseases among exposed workers is not known. In one study of 78 workers in an iron and steel foundry utilizing MDI as a binder, the prevalence of cough, wheeze and chest tightness associated with NSBH was 18% [101]. A specific challenge test with MDI on 11 of the workers showed that six had a specific asthmatic reaction, two had a dual reaction, and four had an isolated late reaction. Four out of the six workers had recurrent nocturnal asthmatic reaction after one challenge test. Both specific IgE and specific IgG antibodies to MDI-HSA conjugate were detected in workers exposed to MDI. The relationship between the clinical disease and the presence of antibodies is not clear in MDI-induced asthma. At this stage, the presence of antibodies should indicate the presence of sensitization and not necessarily disease. In MDI Induced hypersensitivity pneumonitis, however, specific IgE or IgG antibodies are usually present [102].

*Hexamethylene diisocyanate (HDI)*. Hexamethylene diisocyanate is more volatile than TDI but it is used extensively in automobile and aircraft spray paints to produce finishing coatings. Cases of occupational asthma due to HDI exposure have been reported in the literature [82]. A prospective study was carried out on 150 workers exposed to HDI and its trimer during an 18 month period [103]. Specific IgE and IgG antibodies to a protein conjugate were detected in 21% of the workers. The presence of antibodies was not associated with the presence of clinical disease. However, the duration of follow-up was short, and it was not possible to draw any firm conclusions.

#### *Western red cedar*

Occupational asthma due to Western red cedar dust exposure is the most common type of occupational asthma in the Pacific Northwest. In British Columbia, Canada, it accounts for 70% of all the compensation claims for

occupational asthma (Chan-Yeung M, unpublished observation). Western red cedar is used extensively for both indoor and outdoor construction, because of the durability. It is different from other wood in its high content of chemicals in the wood. Plicatic acid (PA) is one of the chemicals present uniquely in this wood and it has been found to be responsible for red cedar asthma [104]. Its structural formula is shown in figure 3. It has a molecular weight of 440 Da.

Red cedar asthma affects 4–14% of the exposed population, depending on the type of industry and the level of dust exposure [105]. There is a good correlation between the dust level and the prevalence of work-related asthma in exposed workers [106, 107]. The higher the dust concentration, the higher the prevalence of work-related asthma. As in TDI-induced asthma, the patients are mostly nonatopic subjects and nonsmokers. Recently, a follow up study of workers in a cedar sawmill showed that NSBH only developed in parallel with the development of red cedar asthma, indicating that NSBH is not a predisposing host factor [108].

The clinical feature of red cedar asthma is similar to TDI-induced asthma. There is a latent period between the onset of exposure and the onset of symptoms. At the onset of the disease, the patients usually complain of cough, wheeze, and shortness of breath at the end of the workshift. Later, with continual exposure, the patients complain of symptoms immediately they are exposed to the wood dust. In the first stages of illness, there is a remission of symptoms during weekends and holidays. Later on, there is no remission of symptoms. Inhalation challenge test with an extract of Western red cedar or plicatic acid induce the same types of asthmatic reactions as patients with TDI. Systemic reaction or alveolar reaction has not been observed.

Follow-up studies of patients with red cedar asthma showed that about 60% of patients failed to recover several years after they were removed from exposure [105]. Those who recovered, tended to be diagnosed early and removed from exposure early. They had normal lung function and a lower degree of NSBH at the time of diagnosis compared to those who failed to recover. For those who had to remain in the same job, persistent asthma was the rule. The use of respirators and the reduction of exposure by a change to a less dusty job did not improve the prognosis [109].

The pathogenic mechanism of asthma induced by PA is not clear. Skin tests with extracts of red cedar and PA have been negative in patients with proven disease [104]. Plicatic acid can be conjugated to human serum albumin (HSA) to form an allergen. Using this conjugate as antigen, specific IgE antibodies can be found by radioallergosorbent tests (RASTs) method in about 20% of patients proven by inhalation challenge [110]. Bronchoalveolar lavage studies of patients before and during immediate asthmatic reaction showed a release of inflammatory mediators, including histamine, prostaglandin D<sub>2</sub>, leukotriene E<sub>4</sub> and thromboxane B<sub>2</sub> into the BAL fluid [111]. During the late reaction, an increase in eosinophils and albumin was found, together with sloughing of the epithelium [112]. Bronchial biopsy performed during

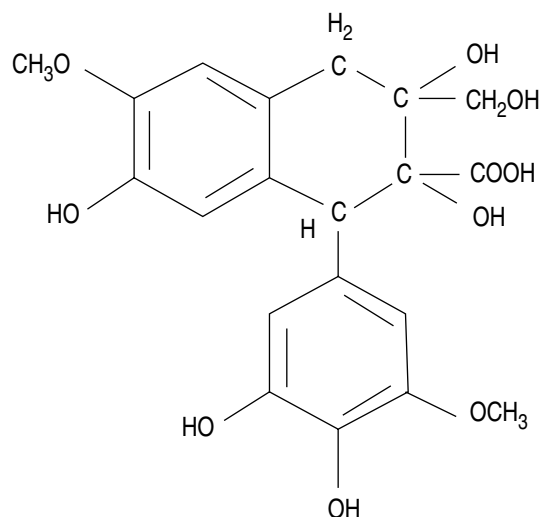


Fig. 3. — Structure of plicatic acid.

the late reaction showed similar findings to patients with allergen-induced asthma [112]. These findings suggest that there might be low levels of specific IgE antibodies to PA-HSA conjugate, not detectable by the RAST method.

To test the above possibility, serum from patients with red cedar asthma was used to passively sensitize human lung fragments, and the fragments were then challenged with PA. No release of histamine was found, even though, under the same conditions, the lung fragments released histamine when passively sensitized and then challenged with grass pollen extract [113]. Basophils from patients with red cedar asthma were found to release histamine on direct challenge with PA. There was no inhibition of release of histamine when the basophils were previously desensitized by incubation with anti-IgE, while this desensitizing protocol reduced the responsiveness of basophils from atopic subjects to a challenge with grass pollen or with anti-IgE. These findings confirmed that specific IgE antibodies cannot be responsible for the release of histamine in patients with red cedar asthma. Further studies need to be carried out to clarify the pathogenic mechanism in red cedar asthma, as this may help us to understand the pathogenic mechanism of nonatopic or intrinsic asthma in general.

#### Amines

Various secondary, tertiary and quaternary amines, either aliphatic, heterocyclic or aromatic, have been reported to cause occupational asthma [114]. These chemicals are found in primary manufacturing and in secondary industries, such as the rubber, cosmetics, shellac, fur and hair dye industries, aluminium soldering, acrylate paint, and photography development [115].

Amines have been shown to cause occupational asthma in workers exposed to epoxy resins, which were produced by amines acting as cross-linking agents binding the resin molecules together [114]. Ethylenediamine

caused occupational asthma in the beauty culture industry and among radiographers and photographers who develop films [115]. Another amine, piperazine, is used as an anthelmintic drug [116]. A survey of 131 workers exposed to piperazine and ethylenediamine revealed that 8% were asthmatic [117]. Specific IgE antibodies to piperazine-HSA conjugate were detected in five out of 72 exposed workers [117]. The mechanism by which amines cause asthma remains unclear.

#### Colophony and fluxes

Colophony is widely-used as a flux in electronic industries to prevent corrosion. It is obtained from pine trees containing abietic acid. The mechanism of occupational asthma due to colophony is not known at present. A specific challenge test induced isolated immediate, isolated late, or dual asthmatic reaction in sensitized subjects. In a survey of 446 electronic workers, a prevalence of work-related symptoms was found in 22% [118]. Atopy was a weak predisposing factor and smoking was not a determinant of work-related symptoms. There was a relationship between the degree of exposure and the prevalence of work-related symptoms. A follow-up study of 39 workers with asthma due to colophony exposure showed that symptoms persisted in half of them up to 4 yrs after exposure ceased [119].

Other soldering fluxes that can cause occupational asthma include zinc chloride and ammonium chloride [120], as well as one containing polyether alcohol-polypropylene glycol [121].

#### Pharmaceutical products

Whilst a number of pharmaceutical products have been shown to cause occupational asthma *via* an IgE-dependent mechanism, such as psyllium, ipecacuanha, pancreatic and glandular extracts, there are others that can cause occupational asthma *via* as yet unidentified mechanisms. These include several antibiotics, such as penicillins, amoxicillin, ampicillin, penicillamine, cephalosporins, phenylglycine acid chloride, and spiramycin [122]. Many of the cases were proven by specific challenge tests. Several nonantibiotic drugs can also cause occupational asthma, such as tylosin tartrate [123], alpha-methyldopa [124], amprolium hydrochloride [125], and morphine [126].

#### Miscellaneous

Exposure to many other chemical compounds in industries, such as chloramine T, a sterilizing agent used in the food and beverage industry [127], persulphate salts used among hairdressers [128], diazonium salts in photo-copying process [129], reactive dyes in textile industry [130] and azobisformamide in plastics industry can give rise to occupational asthma [131]. It will not be possible to discuss each one of them in detail. Readers should refer



to table 1 in the Appendix and the text book "Asthma in the Workplace" [3, 4].

### Agents causing occupational asthma through nonimmunological mechanisms

Nonimmunological occupational asthma may or may not occur after a latency period of exposure to an agent(s) which does not induce immune sensitization as determined by currently available technology. The main distinction between this type of occupational asthma and immunologically-dependent ones is that in the majority of cases re-exposure of the affected subjects to small amounts of the offending agent does not reproduce the symptoms. Several types of occupational asthma belong to this category, and are described briefly below.

#### *Reactive Airways Dysfunction Syndrome (RADS)*

This syndrome was originally coined by BROOKS and co-workers [132] in 1985, to describe 10 subjects who developed symptoms of asthma and NSBH after one single exposure to high levels of an irritant vapour, fume or smoke. None of the subjects had a previous history of respiratory symptoms. Respiratory symptoms and NSBH persisted for about 3 yrs after the initial event. The incriminated exposure was either short-lived, often lasting just a few minutes or, on occasions, was as long as 12 h. There was a time interval between exposure and development of symptoms, varying from several minutes to several hours. The exposure was usually due to an accident or a situation where there was very poor ventilation and high exposure. Bronchial biopsies were performed on two cases. Whilst there was evidence of epithelial injury, with bronchial wall inflammation, the cells were lymphocytes and plasma cells; eosinophils were not found. In addition, there was no evidence of mucous gland hyperplasia, basement membrane thickening or smooth muscle hyperplasia. However, more recent biopsy data on subjects with this syndrome revealed typical histopathological features of asthma [133].

The incriminating agents were all irritants, including floor sealant, heated acid, fumigating fog, metal coating remover, acetic acid [134], smoke inhalation, TDI, and ammonia fumes [133].

Since re-exposure of these subjects to low levels of the incriminating agent did not reproduce symptoms, the diagnosis is a clinical one based on the history. The number of case reports is limited. The outcome of these subjects has not been as well documented as other types of occupational asthma. The syndrome needs to be studied in more detail to define it better.

Although BROOKS and co-workers [132] first coined this syndrome as RADS, there have been reports of persistent symptoms and a single, high concentration environmental or occupational irritant exposure. Five cases of accidental exposure to high levels of sulphur dioxide have been reported, with three survivors developing severe and another showing a mild degree of airflow obstruction [135]. A man aged 50 yrs developed obstructive

airways disease after exposure to high concentration of ammonia vapours [136]. Several other studies of the sequelae of chlorine gas exposure showed similar findings [137, 138].

There are no data on the prevalence of this syndrome. The host risk factors are not known. A recent review of 59 consecutive cases of occupational asthma in the Toronto area in Canada showed that 17% were caused by exposure to irritants, suggesting that this syndrome is not that uncommon [139]. Longitudinal data are not available to indicate clearly that these subjects had no evidence of NSBH before the incident.

The pathogenesis of RADS is not clear. It has been postulated that the high level of exposure induces massive airway injury, followed by activation of non-adrenergic, noncholinergic (NANC) pathways *via* axon reflexes and the onset of neurogenic inflammation [133]. Nonspecific macrophage activation and mast cell degranulation may also occur, releasing proinflammatory mediators, with recruitment of inflammatory cells to the site of injury [133]. Epithelial injury has been shown to reduce the availability of an epithelial-derived relaxing factor and decrease neutral endopeptidase, leading to activation of the NANC nerves. As with other aspects of this syndrome, more research is necessary.

#### *Potroom asthma*

Potroom asthma has been reported among workers on potlines in aluminum smelting plants for many years [140]. Potroom workers are exposed to a number of contaminants, including particulate and gaseous fluoride, hydrofluoric acid, sulphur dioxide and hydrocarbons. The agent responsible for potroom asthma has not yet been identified. It has been postulated that the air contaminants in the potroom, if present in high concentrations, may induce asthma, as in RADS. The latent period between onset of exposure and the onset of symptoms varied from one week to 10 yrs [140]. Potroom asthma has been reported more frequently in smelters in Australia and Norway than in North America.

#### *Machining fluid*

Workers exposed to machining fluid can develop work-related asthma [141]. Machining fluids are contaminated with trace metals, additives such as antioxidants, detergents, anti-foam agents, and bactericidal agents. The latter are necessary to prevent the growth of bacteria and fungi. In addition to well-defined cases of asthma, cross-shift change in forced expiratory volume in one second (FEV<sub>1</sub>) of less than 10% was reported in 24% of the heavily exposed and in 10% of the least exposed machinists [141]. The pathogenesis is not known, although the machining fluid is likely to act as an irritant.

#### *Formaldehyde*

Formaldehyde is an irritant at high concentrations, but a sensitizer in lower concentrations. Formaldehyde is

widely used in hospital environments, furniture manufacture and as insulation in buildings in the 1980s. In a prevalence study of 230 workers exposed to formaldehyde at work, 12 had significant bronchoconstriction at levels of 1–2 ppm during specific challenge tests [142]. Attempts to detect specific antibodies in the sera of workers exposed to formaldehyde have so far been unsuccessful [143].

### Summary

This review gives a brief summary of the more commonly encountered aetiological causes of occupational asthma. Interested readers should refer to the textbook "Asthma in the Workplace" edited by BERNSTEIN *et al.* [4]. With the introduction of more chemicals into the workplace, the list is likely to increase. It should be emphasized that the absence of the suspected agent in any listing of aetiological causes of occupational asthma should not exclude the diagnosis. An international effort to prepare a data base similar to the Minitel System in France would be most helpful to clinicians and investigators in this field [144].

### References

- Meredith SK, Taylor VM, McDonald JC. Occupational respiratory disease in the United Kingdom, 1989: a report to the British Thoracic Society and the Society of Occupational Medicine by the SWORD project group. *Br J Ind Med* 1991; 48: 292–298.
- Lagier F, Cartier A, Malo JL. Medico-legal statistics on occupational asthma in Quebec between 1986 and 1988. *Rev Mal Respir* 1990; 7: 337–341.
- Chan-Yeung M, Malo JL. Compendium I. Table of the major inducers of occupational asthma. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 595–623.
- Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993.
- Agius RM, Nee J, McGovern B, Robertson A. Structure activity hypothesis in occupational asthma caused by low molecular weight substances. *Ann Occup Hyg* 1991; 35: 129–137.
- Venables KM, Dally MB, Nunn AJ, *et al.* Smoking and occupational allergy in workers in a platinum refinery. *Br Med J* 1989; 299: 939–942.
- Newman-Taylor AJ, Gordon S. Laboratory animal and insect allergy. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 399–414.
- Botham PA, Davies GE, Teasdale EL. Allergy to laboratory animals: a prospective study of its incidence and of the influence of atopy on its development. *Br J Ind Med* 1987; 44: 627–632.
- Chan-Yeung M, Malo JL. Natural history of occupational asthma. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 299–322.
- Slovak AJM, Hill RN. Does atopy have any predictive value for laboratory animal allergy? A comparison of different concepts of atopy. *Br J Ind Med* 1987; 44: 129–132.
- Reed CE, Swanson MC, Li JTC. Environmental monitoring of protein aeroallergens. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 249–275.
- Edwards RG, Beeson MF, Dewdney JM. Laboratory animal allergy: the measurement of airborne urinary allergens and the effects of different environmental conditions. *Lab Animal* 1983; 17: 235–239.
- Gordon S, Tee RD, Lawson D, Wallace J, Newman-Taylor AJ. Reduction of airborne allergenic urine proteins from laboratory rats. *Br J Ind Med* 1992; 49: 416–422.
- Sakaguchi M, Inouye S, Kamimura H, Kimura M, Yamazaki S. Evaluation of countermeasures for reduction of mouse airborne allergens. *Lab Animal Sci* 1990; 40: 613–615.
- Brennan NJ. Pig Butcher's asthma: case report and review of the literature. *Ir Med J* 1985; 78: 321–322.
- Joliat TL, Weber RW. Occupational asthma and rhinoconjunctivitis from inhalation of crystalline bovine serum albumin powder. *Ann Allergy* 1991; 66: 301–304.
- Bernstein DI, Smith AB, Moller DR, *et al.* Clinical and immunologic studies among egg-processing workers with occupational asthma. *J Allergy Clin Immunol* 1987; 80: 791–797.
- Smith AB, Bernstein DI, London MA, *et al.* Evaluation of occupational asthma from airborne egg protein exposure in multiple settings. *Chest* 1990; 98: 398–404.
- Warren CPW, Holford-Strevens V, Sinha RN. Sensitization in a grain handler to the storage mite *Lepidoglyphus destructor* (Schrack) *Ann Allergy* 1983; 50: 30–33.
- Cuthbert OD, Brostoff J, Wraith DG, Brighton WD. "Barn allergy": asthma and rhinitis due to storage mites. *Clin Allergy* 1979; 9: 229–236.
- Reunala T, Bjorksten F, Forstrom L, Kanerva L. IgE-mediated occupational allergy to a spider mite. *Clin Allergy* 1983; 13: 383–388.
- Lutsky I, Teichtahl H, Bar-Sela S. Occupational asthma due to poultry mites. *J Allergy Clin Immunol* 1984; 73: 56–60.
- Frankland AW, Lunn JA. Asthma caused by the grain weevil. *Br J Ind Med* 1965; 22: 157–159.
- Frankland AW. Locust sensitivity. *Ann Allergy* 1953; 11: 445–453.
- Burge PS, Edge G, O'Brien IM, Harries MG, Hawkins R, Pepys J. Occupational asthma in a research centre breeding locusts. *Clin Allergy* 1980; 10: 355–363.
- Spiekma FTM, Vooren PH, Kramps JA, Dijkman JH. Respiratory allergy to laboratory fruit flies (*Drosophila melanogaster*). *J Allergy Clin Immunol* 1986; 77: 108–113.
- Thiel H. Baker's asthma. Epidemiological and clinical findings: needs for prospective studies. In: JW Kerr, MA Ganderton, eds. *Proceedings of invited symposia. Eleventh International Congress of Allergology and Clinical Immunology*. 1983; London: pp. 429–433.
- Block G, Tse KS, Kijek K, Chan H, Chan-Yeung M. Baker's asthma. Studies of the cross-antigenicity between different cereal grains. *Clin Allergy* 1984; 14: 177–185.
- Blands J, Diamant B, Kallos P, Kallos-Deffner L, Lowenstein H. Flour allergy in bakers. I. Identification of allergenic fractions in flour and comparison of diagnostic methods. *Int Arch Allergy Appl Immunol* 1976; 52: 392–402.
- Walsh BJ, Wrigley CW, Musk AW, Baldo BA. A comparison of the binding of IgE in the sera of patients

- with baker's asthma to soluble and insoluble wheat-grain proteins. *J Allergy Clin Immunol* 1985; 76: 23–28.
31. Klaustermeyer WB, Bardana EJ, Hale FC. Pulmonary hypersensitivity to *Alternaria* and *Aspergillus* in baker's asthma. *Clin Allergy* 1977; 7: 227–233.
  32. Baur X, Fruhmann G, Haug B, Rasche B, Reiher W, Weiss W. Role of *Aspergillus* amylase in baker's asthma. *Lancet* 1986; i: 43.
  33. Baur X, Sauer W, Weiss W. Baking additives as new allergens in baker's asthma. *Respiration* 1988; 54: 70–72.
  34. Musk AW, Venables KM, Crook B, *et al.* Respiratory symptoms, lung function, and sensitisation to flour in a British bakery. *Br J Ind Med* 1989; 46: 636–642.
  35. Malo JL, Ghezzi H, D'Aquino C, L'Archevêque J, Cartier A, Chan-Yeung M. Natural history of occupational asthma: relevance of type of agent and other factors in the rate of development of symptoms in affected subjects. *J Allergy Clin Immunol* 1992; 90: 937–944.
  36. Block G, Tse KS, Kijek K, Chan H, Chan-Yeung M. Baker's asthma. *Clin Allergy* 1983; 13: 359–370.
  37. Davies RJ, Green M, Schoefield NM. Recurrent nocturnal asthma after exposure to grain dust. *Am Rev Respir Dis* 1976; 114: 1011–1019.
  38. Malo JL, Cartier A, L'Archevêque J, *et al.* Prevalence of occupational asthma and immunologic sensitization to psyllium among health personnel in chronic care hospitals. *Am Rev Respir Dis* 1990; 142: 1359–1366.
  39. Busse WW, Schoenwetter WF. Asthma from psyllium in laxative manufacture. *Ann Intern Med* 1975; 83: 361–362.
  40. Cartier A, Malo JL, Dolovich J. Occupational asthma in nurses handling psyllium. *Clin Allergy* 1987; 17: 1–6.
  41. Gauss WF, Alarie JP, Karol MH. Workplace allergenicity of a psyllium-containing bulk laxative. *Allergy* 1985; 40: 73–76.
  42. Scott D. Psyllium-induced asthma. *Postgrad Med* 1987; 82: 160–167.
  43. Terho EO, Torkko M. Occupational asthma from psyllium laxatives. *Duodecim* 1980; 96: 1213–1216.
  44. Bernton HS. The allergenicity of psyllium seed. *Med Ann DC* 1970; 39: 313–317.
  45. Schwartz HJ, Arnold JL, Strohl KP. Occupational allergic rhinitis reaction to psyllium. *J Occup Med* 1989; 31: 624–626.
  46. Bardy JD, Malo JL, Séguin P, *et al.* Occupational asthma and IgE sensitization in a pharmaceutical company processing psyllium. *Am Rev Respir Dis* 1987; 135: 1033–1038.
  47. Goransson K, Michaelson NG. Ispagula powder. An allergen in the work environment. *Scand J Work Environ Health* 1979; 5: 257–261.
  48. Malo JL, Cartier A, L'Archevêque J, *et al.* Prevalence of occupational asthma and immunological sensitization to guar gum among employees at a carpet-manufacturing plant. *J Allergy Clin Immunol* 1990; 86: 562–569.
  49. Spielman AD, Baldwin HS. Atopy to acacia (gum arabic). *J Am Med Assoc* 1933; 101: 444–445.
  50. Bohner CB, Sheldon JM, Trenis JW. Sensitivity to gum acacia, with a report of ten cases of asthma in printers. *J Allergy* 1941; 12: 290–294.
  51. Flindt MLH. Pulmonary disease due to inhalation of derivatives of *Bacillus subtilis* containing proteolytic enzyme. *Lancet* 1969; i: 1177–1181.
  52. Pepys J, Longbottom JL, Hargreave FE, Faux J. Allergic reactions of the lungs to enzymes of *Bacillus subtilis*. *Lancet* 1969; 1: 1811–1814.
  53. Newhouse ML, Tagg B, Pocock SJ, McEwan AC. An epidemiological study of workers producing enzyme washing powders. *Lancet* 1970; i: 689–693.
  54. Bernstein IL. Enzyme allergy in populations exposed to long-term, low-level concentrations of household laundry products. *J Allergy Clin Immunol* 1972; 49: 219–237.
  55. Perdu D, Lavaud F, Cossart C, *et al.* Enzymes des lessives: le risque de sensibilisation professionnelle a-t-il disparu? *Rev Mal Respir* 1992; 9: 443–448.
  56. Pepys J, Mitchell J, Hawkins R, Malo JL, Ashforth GK, Wilson ER. A longitudinal study of possible allergy to enzyme detergents. *Clin Allergy* 1985; 15: 101–116.
  57. Bernstein DI, Malo JL. High molecular weight agents. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 373–398.
  58. Jyo T, Kohmoto K, Katsutani T, Otsuka T, Oka SD, Mitsui S. Hoya (sea-squirt) asthma. In: *Occupational Asthma*. London, Von Nostrand Reinhold, 1980; pp. 209–228.
  59. Gaddie J, Legge JS, Friend JAR, Reid TMS. Pulmonary hypersensitivity in prawn workers. *Lancet* 1980; ii: 1350–1353.
  60. Cartier A, Malo J-L, Forest F, *et al.* Occupational asthma in snow crab-processing workers. *J Allergy Clin Immunol* 1984; 74: 261–269.
  61. Zeiss CR, Patterson R. Acid anhydrides. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 439–457.
  62. Kern RA. Asthma and allergic rhinitis due to sensitization to phthalic anhydride. Report of a case. *J Allergy* 1939; 10: 164–165.
  63. Maccia CA, Bernstein IL, Emmett EA, Brooks SM. *In vitro* demonstration of specific IgE in phthalic anhydride hypersensitivity. *Am Rev Respir Dis* 1976; 113: 701–704.
  64. Zeiss CR, Wolkonsky P, Chacon R, *et al.* Syndromes in workers exposed to trimellitic anhydride. *Ann Intern Med* 1983; 98: 8–12.
  65. Zeiss CR, Patterson R, Pruzansky JJ, Miller MM, Rosenberg M, Levitz D. Trimellitic anhydride-induced airway syndromes: clinical and immunologic studies. *J Allergy Clin Immunol* 1977; 60: 96–103.
  66. Zeiss CR, Mitchell J, Van Peenen PFD, *et al.* Evaluation of an entire chemical plant related to trimellitic anhydride (TMA) exposure. *J Allergy Clin Immunol* 1990; 85: 190.
  67. Venables KM, Topping MD, Howe W, Luczynska CM, Hawkins R, Newman Taylor AJ. Interaction of smoking and atopy in producing specific IgE antibody against a hapten protein conjugate. *Br Med J* 1985; 290: 201–204.
  68. Venables KM, Topping MD, Nunn AJ, Howe W, Newman-Taylor AJ. Immunologic and functional consequences of chemical (tetrachlorophthalic anhydride)-induced asthma after four years of avoidance of exposure. *J Allergy Clin Immunol* 1987; 80: 212–218.
  69. Bernstein IL, Brooks SM. Metals. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 459–479.
  70. Malo JL, Cartier A, Doepner M, Nieboer E, Evans S, Dolovich J. Occupational asthma caused by nickel sulfate. *J Allergy Clin Immunol* 1982; 69: 55–59.
  71. Block GT, Yeung M. Asthma induced by nickel. *J Am Med Assoc* 1982; 247: 1600–1602.
  72. Kusaka Y, Nakano Y, Shirakawa T, Fujimura N, Kato

- M, Heki S. Lymphocyte transformation test with nickel in hard metal asthma: another sensitizing component of hard metal. *Ind Health* 1991; 29: 153-160.
73. Karasek ST, Karasek M. The use of platinum paper. Report of Illinois State Commission of Occupational Disease, Springfield, Illinois. 1911; 97.
  74. Hunter D, Milton R, Perry KMA. Asthma caused by the complex salts of platinum. *Br J Ind Med* 1945; 2: 92-98.
  75. Pepys J, Pickering CAC, Hughes EG. Asthma due to inhaled chemical agents: complex salts of platinum. *Clin Allergy* 1972; 2: 391-396.
  76. Biagini RE, Bernstein IL, Gallagher JS, Moorman WJ, Brooks S, Gann PH. The diversity of reaginic immune responses to platinum and palladium metallic salts. *J Allergy Clin Immunol* 1985; 76: 794-802.
  77. Hughes EG. Medical surveillance of platinum refinery workers. *J Soc Occup Med* 1980; 30: 27-30.
  78. Malo JL, Cartier A. Occupational asthma due to fumes of galvanized metal. *Chest* 1987; 92: 375-377.
  79. Kawane H, Soejima R, Umeki S, Niki Y. Metal fume and asthma. *Chest* 1988; 93: 1116-1117.
  80. Perrin B, Cartier A, Ghezze H, et al. Reassessment of the temporal patterns of bronchial obstruction after exposure to occupational sensitizing agents. *J Allergy Clin Immunol* 1991; 87: 630-639.
  81. Chan-Yeung M. Occupational asthma. *Chest* 1990; 98: 148S-161S.
  82. Vandenplas O, Malo JL, Saetta M, Mapp CE, Fabbri L. Occupational asthma and extrinsic alveolitis due to isocyanates: current status and perspectives. *Br J Ind Med* 1993; 50: 213-228.
  83. Butcher BT, Jones RN, O'Neil CE, et al. Longitudinal study of workers employed in the manufacture of toluene diisocyanate. *Am Rev Respir Dis* 1977; 116: 411-421.
  84. Bignon J, Aron A, Li YJ, et al. HLA DQB1 and DPB1 alleles are associated with isocyanate-sensitive occupational asthma. *Am Rev Respir Dis* 1992; A321.
  85. NIOSH recommended standard for occupational exposure to diisocyanates. US Department of Health, Education and Welfare, Public Health Service, Center for Disease Control, National Institute for Occupational Safety and Health. No. 78-125, 1979.
  86. Karol MH. Concentration-dependent immunologic response to toluene diisocyanate (TDI) following inhalation exposure. *Toxicol Appl Pharmacol* 1983; 68: 229-241.
  87. Cibulas W Jr, Murlas CG, Miller ML, et al. Toluene diisocyanate-induced airway hyperreactivity and pathology in the guinea-pig. *J Allergy Clin Immunol* 1986; 77: 828-834.
  88. Kennedy AL, Stock MF, Alarie Y, Brown WE. Uptake and distribution of <sup>14</sup>C during and following inhalation exposure to radioactive toluene diisocyanate. *Toxicol Appl Pharmacol* 1989; 100: 280-292.
  89. Kennedy AL, Brown WE. Modification of airway proteins and induction of secondary responses by inhalation exposure to isocyanates. *Am Rev Respir Dis* 1989; 139: 387A
  90. Banks DE, Rando RJ. Recurrent asthma induced by toluene diisocyanate. *Thorax* 1988; 43: 660-662.
  91. Mapp CE, Corona PC, de Marzo N, Fabbri L. Persistent asthma due to isocyanates. A follow-up study of subjects with occupational asthma due to toluene diisocyanate. *Am Rev Respir Dis* 1988; 137: 1326-1329.
  92. Paggiaro PL, Loi AM, Rossi O, et al. Follow-up study of patients with respiratory disease due to toluene diisocyanate (TDI). *Clin Allergy* 1984; 14: 463-469.
  93. Allard C, Cartier A, Ghezze H, Malo J-L. Occupational asthma due to various agents. Absence of clinical and functional improvement at an interval of four or more years after cessation of exposure. *Chest* 1989; 96: 1046-1049.
  94. Lozewicz S, Assoufi BK, Hawkins R, Newman-Taylor AJ. Outcome of asthma induced by isocyanates. *Br J Dis Chest* 1987; 81: 14-27.
  95. Patterson R, Hargreave FE, Grammer LC, Harris KE, Dolovich J. Toluene diisocyanate respiratory reactions I. Reassessment of the problem. *Int Arch Allergy Appl Immunol* 1987; 84: 93-100.
  96. Bernstein IL. Isocyanate-induced pulmonary diseases: a current perspective. *J Allergy Clin Immunol* 1982; 70: 24-31.
  97. Patterson R, Zeiss CR, Pruzansky JJ. Immunology and immunopathology of trimellitic anhydride pulmonary reactions. *J Allergy Clin Immunol* 1982; 70: 19-23.
  98. Davies RJ, Butcher BT, O'Neil CE, Salvaggio JE. The *in vitro* effect of toluene diisocyanate on lymphocyte cyclic adenosine monophosphate production by isoproterenol, prostaglandin, and histamine. A possible mode of action. *J Allergy Clin Immunol* 1977; 60: 223-229.
  99. Fabbri LM, Ciaccia A, Maestrelli P, Saetta M, Mapp CE. Pathophysiology of occupational asthma. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 61-92.
  100. Fabbri LM, Danieli D, Crescioli S, et al. Fatal asthma in a subject sensitized to toluene diisocyanate. *Am Rev Respir Dis* 1988; 137: 1494-1498.
  101. Zammit-Tabona M, Sherkin M, Kijek K, Chan H, Chan-Yeung M. Asthma caused by diphenylmethane diisocyanate in foundry workers. Clinical, bronchial provocation, and immunologic studies. *Am Rev Respir Dis* 1983; 128: 226-230.
  102. Vandenplas O, Malo JL, Dugas M, et al. Hypersensitivity pneumonitis-like reaction among workers exposed to diphenylmethane diisocyanate (MDI). *Am Rev Respir Dis* 1993; 147: 338-346.
  103. Grammer LC, Eggum P, Silverstein M, Shaughnessy MA, Liotta JL, Patterson R. Prospective immunologic and clinical study of a population exposed to hexamethylene diisocyanate. *J Allergy Clin Immunol* 1988; 82: 627-633.
  104. Chan-Yeung M, Barton GM, MacLean L, Grzybowski S. Occupational asthma and rhinitis due to western red cedar (*Thuja plicata*). *Am Rev Respir Dis* 1973; 108: 1094-1102.
  105. Chan-Yeung M. Western red cedar and other wood dusts. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *New York, Marcel Dekker Inc., 1993; pp. 503-531.*
  106. Vedal S, Chan-Yeung M, Enarson D, et al. Symptoms and pulmonary function in western red cedar workers related to duration of employment and dust exposure. *Arch Environ Health* 1986; 41: 179-183.
  107. Brooks SM, Edwards JJ, Apol A, Edwards FH. An epidemiologic study of workers exposed to western red cedar and other wood dust. *Chest* 1981; 80: 30-32.
  108. Chan-Yeung M, Desjardins A. Nonspecific bronchial hyperresponsiveness and dust exposure in patients with asthma due to western red cedar. Serial observations before and after the development of symptoms. *Am Rev Respir Dis* 1992; 146: 1606-1609.
  109. Côté J, Kennedy S, Chan-Yeung M. Outcome of patients with cedar asthma with continuous exposure. *Am Rev Respir Dis* 1990; 141: 373-376.

110. Tse KS, Chan H, Chan-Yeung M. Specific IgE antibodies in workers with occupational asthma due to western red cedar. *Clin Allergy* 1982; 12: 249–258.
111. Chan-Yeung M, Chan H, Salari H, Lam S. Histamine, leukotrienes and prostaglandins release in bronchial fluid during plicatic acid-induced bronchoconstriction. *J Allergy Clin Immunol* 1989; 84: 762–768.
112. Lam S, LeRiche J, Phillips D, Chan-Yeung M. Cellular and protein changes in bronchial lavage fluid after late asthmatic reaction in patients with red cedar asthma. *J Allergy Clin Immunol* 1987; 80: 44–50.
113. Frew AJ, Chan H, Dryden P, Salari H, Lam S, Chan-Yeung M. Immunological studies of the mechanisms of occupational asthma due to western red cedar. *J Allergy Clin Immunol* 1993; 92 (3): 466–478.
114. Hagmar L, Nielsen J, Skerfving S. Clinical features and epidemiology of occupational obstructive respiratory disease caused by small molecular weight organic chemicals. Epidemiology of allergic diseases. *Monogr Allergy* 1987; 21: 42–58.
115. Gelfand HH. Respiratory allergy due to chemical compounds encountered in the rubber, lacquer, shellac, and beauty culture industries. *J Allergy* 1963; 34: 374–381.
116. McCullagh SF. Allergenicity of piperazine: a study in environmental aetiology. *Br J Ind Med* 1968; 25: 319–325.
117. Hagmar L, Welinder H. Prevalence of specific IgE antibodies against piperazine in employees of a chemical plant. *Int Arch Allergy Appl Immunol* 1986; 81: 12–16.
118. Herks WH, Burge PS, Rehahn M, Green M. Work-related respiratory disease in employees leaving an electronics factory. *Thorax* 1979; 34: 19–22.
119. Burge PS. Occupational asthma in electronics workers caused by colophony fumes: follow-up of affected workers. *Thorax* 1982; 37: 348–353.
120. Weir DC, Robertson AS, Jones S, Burge PS. Occupational asthma due to soft corrosive soldering fluxes containing zinc chloride and ammonium chloride. *Thorax* 1989; 44: 220–223.
121. Stevens JJ. Asthma due to soldering flux: a polyether alcohol-polypropylene glycol mixture. *Ann Allergy* 1976; 36: 419–422.
122. Lee HS, Wang YT, Yeo CT, Tan KT, Ratnam KV. Occupational asthma due to tylosin tartrate. *Br J Ind Med* 1989; 46: 498–499.
123. Harries MG, Newman Taylor A, Wooden J, MacAuslan A. Bronchial asthma due to alpha-methyl dopa. *Br Med J* 1979; 1 (6176): 1461.
124. Greene SA, Freedman S. Asthma due to inhaled chemical agents: amprolium hydrochloride. *Clin Allergy* 1976; 6: 105–108.
125. Biagini RE, Klineciewicz SL, Henningsen GM, et al. Antibodies to morphine in workers exposed to opiates at a narcotics manufacturing facility and evidence for similar antibodies in heroin abusers. *Life Sci* 1990; 47: 897–908.
126. Bourne MS, Flindt MLH, Walker JM. Asthma due to industrial use of chloramine. *Br Med J* 1979; 2: 10–12.
127. Blainey AD, Ollier S, Cundell D, Smith RE, Davies RJ. Occupational asthma in a hairdressing salon. *Thorax* 1986; 41: 42–50.
128. Armeli G. Bronchial asthma from diazonium salts. *Med Lav* 1968; 59: 463–466.
129. Alanko K, Keskinen H, Byorksten F, Ojanen S. – Immediate-type hypersensitivity to reactive dyes. *Clin Allergy* 1978; 8: 25–31.
130. Malo JL, Pineau L, Cartier A. Occupational asthma due to azobisformamide. *Clin Allergy* 1985; 15: 261–264.
131. Malo JL, Bernstein IL. Other chemical substances causing occupational asthma. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 481–502.
132. Brooks SM, Weiss MA, Bernstein IL. Reactive airways dysfunction syndrome (RADS). Persistent asthma syndrome after high level irritant exposures. *Chest* 1985; 88: 376–384.
133. Brooks SM, Bernstein IL. Reactive airways dysfunction syndrome or irritant-induced asthma. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 533–549.
134. Kern DG. Outbreak of the reactive airways dysfunction syndrome after a spill of glacial acetic acid. *Am Rev Respir Dis* 1991; 144: 1058–1064.
135. Charan NB, Myers CG, Lakshminarayan S, Spencer TM. Pulmonary injuries associated with acute sulphur dioxide inhalation. *Am Rev Respir Dis* 1979; 119: 555–560.
136. Flury KE, Ames DE, Rodarte JR, Rodgers R. Airway obstruction due to inhalation of ammonia. *Mayo Clin Proc* 1983; 58: 389–393.
137. Salisbury DA, Enarson DA, Chan-Yeung M, Kennedy SM. First-aid reports of acute chlorine gassing among pulp mill workers as predictors of lung health consequences. *Am J Ind Med* 1991; 20: 71–81.
138. Kennedy SM, Enarson DA, Janssen RG, Chan-Yeung M. Lung health consequences of reported accidental chlorine gas exposures among pulp mill workers. *Am Rev Respir Dis* 1991; 143: 74–79.
139. Tarlo SM, Broder I. Irritant-induced occupational asthma. *Chest* 1989; 96: 297–300.
140. Abramson MJ, Wlodarczyk JH, Saunders NA, Hensley MJ. Does aluminium smelting cause lung disease. *Am Rev Respir Dis* 1989; 139: 1042–1057.
141. Kennedy SM, Greaves IA, Kriebel D, Eisen EA, Smith TJ, Woskie SR. Acute pulmonary responses among automobile workers exposed to aerosols of machining fluids. *Am J Ind Med* 1989; 15: 627–641.
142. Nordman H, Keskinen H, Tuppurainen M. Formaldehyde asthma: rare or overlooked? *J Allergy Clin Immunol* 1985; 75: 91–99.
143. Grammer LC, Harris KE, Shaughnessy MA, et al. Clinical and immunologic evaluation of 37 workers exposed to gaseous formaldehyde. *J Allergy Clin Immunol* 1990; 86: 177–181.
144. Perrin B, Dhivert H, Godard P, Bousquet J, Michel FB. The telematic information service (Minitel) on occupational asthma in France. In: Bernstein IL, Chan-Yeung M, Malo JL, Bernstein DI, eds. *Asthma in the Workplace*. New York, Marcel Dekker Inc., 1993; pp. 635–638.

## Appendix

Table 1. – Selection of key references in occupational asthma (Reference list at end of table)

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
<b>High molecular weight agents</b>									
<b>Animal-derived antigens</b>									
Laboratory animal	Laboratory workers	[1]	296	13	17% +	34% of 255 +	ND	ND	
Cow dander	Agricultural workers	[2]	5	NA	100% +	100% +	Neg precipitin	100% +	
Chicken	Poultry workers	[3]	49	NA	100%	ND	Immunoblotting	ND	
Pig	Butcher	[4, 5]	14	NA	79% +	79% +	ND	1/1+	
Frog	Frog catcher	[6]	1	NA	ND	+	ND	ND	+PEFR monitoring
Lactoserum	Dairy industry	[7]	1	NA	+	+	Neg precipitin + basophil	ND	
Casein (cow's milk)	Tanner	[8]	1	NA	+	+	Degranulation ND	+	
Bat guano	Various	[9]	7	NA	+	+	RAST inhibition	ND	
[10]									
<b>Insects</b>									
Grain mite	Farmers	[11]	290	12	21% +	19% of 219 +	ND	ND	
	Grain-store workers	[12]	133	33	25 +	23% of 128 +	ND	1/1 +	21% of 116 with + PC <sub>20</sub> Reduced FEV <sub>1</sub>
Locust	Laboratory workers	[13]	118	26	32% of 113 +	Done	Specific IgG	ND	
		[14]	15	60	77% +	53%	RAST inhibition	ND	
Screw worm fly	Flight crews	[15]	182	25	91% of 11 +	ND	ND	ND	
Cricket	Laboratory workers	[16]	2	NA	+	+	Passive transfer	+	
Bee moth	Fish bait breeder	[17]	1	NA	+	ND	Passive transfer	ND	
							Histamine release	+	
Moth, butterfly	Entomologists	[18]	2	NA	+	ND	ND	ND	
Mexican bean weevil	Seed house	[19]	2	NA	+	ND	Passive transfer	ND	
Fruit fly	Laboratory workers	[20]	22	32	27% +	27% +	RAST inhibition	21% of 14 +	
Honeybee	Honey processors	[21]	1	NA	+	+	ND	+	
<i>L. caesar</i> larvae	Anglers	[22]	5	NA	75% of 4 +	80% of 5 +	RAST inhibition	1/1 +	
Lesser mealworm	Grain and poultry workers	[23]	3	NA	Neg	100% of 3 +	RAST inhibition	ND	
Fowl mite	Poultry workers	[24]	13	NA	77% +	60%	ND	1/1 +	
Barn mite	Farmers	[25]	38	NA	100% +	~100%	ND	ND	
Mites and parasites	Flour handlers	[26]	12	NA	ND	+	ND	ND	
Acarian	Apple growers	[27]	4	NA	+	ND	Neg precipitins	ND	
Daphnia	Fish food-store	[28]	2	NA	+	+	ND	2/2 +	
Weeping fig	Plant keepers	[29]	84	7	21% +	21%	ND	100% of 6 +	PC <sub>20</sub>
Sheep blowfly	Technicians	[30]	53	24	ND	67% of 15 +	ND	ND	
Silkworm	Silk workers	[31]	53	34%	ND	ND	ND	ND	
<b>Plants</b>									
Grain dust	Grain elevators	[32]	610	~40	9% +	ND	Neg precipitins	ND	Spirometry pre-post shift
		[33, 34]	502	47	~50% of 51 exposed +	ND	ND	ND	FEV <sub>1</sub> , volumes
		[35]	22	NA	0% +	ND	Neg precipitins	27% +	50% PC <sub>20</sub> +
Wheat, rye and soya flour	Bakers, millers	[36]	279	35	9% + (cereals)	ND	ND	ND	FEV <sub>1</sub> , PC <sub>20</sub>
		[37]	7	100	100%+	100%+	100% neg	57% +	
		[38]	9	100	ND	100% +	Western blotting, etc.	ND	
<i>Lathyrus sativus</i>	Flour handler	[39]	1	NA	+	ND	+ precipitins	+	
<i>Vicia sativa</i>	Farmer	[40]	1	NA	+	+	+ precipitins, passive transfer	+	
Buckwheat	Bakers	[41]	3	NA	100% +	ND	ND	ND	
Gluten	Bakers	[42]	1	NA	+	+	RAST inhibition	+	
Coffee bean	Food processor	[43]	372	34	24% +	12% +	ND	ND	Function tests
		[44]	45	9	9–40% +	ND	ND	ND	Spirometry
		[45]	22	NA	82% +	50% +	ND	67% of 12 +	PC <sub>20</sub> + in 14
Castor bean	Oil industry	[46]	14	NA	100% +	100% +	ND	ND	
Tea	Tea processors	[47]	3	NA	100% neg	ND	ND	100% of 3+	
Herbal tea	Herbal tea processors	[48]	1	NA	ND	Neg	ND	+	

The number of subjects tested is not specified if it included all subjects; otherwise it is mentioned. †: Prevalence of symptoms compatible with work-related asthma unless otherwise stated. \*: based on challenge data; \*\*: presence of bronchial hyperresponsiveness; †: subjects with symptoms. All proportions including 3 or more as the denominator are expressed as %. BAL: bronchoalveolar lavage; 3-DMAPA: 3-(dimethylamino)-propylamine; EPO: epoxy; FEV<sub>1</sub>: forced expiratory volume in one second; HDI: hexamethylene diisocyanate; HSA: human serum albumin; IgG: immunoglobulin G; indust: industry; manuf: manufacturer; MDI: methylene diphenyldiisocyanate; NA: not applicable; ND: not done; neg: negative. PCA: passive cutaneous anaphylaxis; PC<sub>20</sub>: provocative concentration producing a 20% fall in FEV<sub>1</sub>; PEF: peak expiratory flow rate; PPI: polymethylene polyphenylisocyanate; RAST: radioallergosorbent test; TDI: toluene diisocyanate;

Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
Tobacco leaf	Tobacco manufacturers	[49] [50]	1 16	NA 69	+ ND	+ ND	ND ND	+ ND	PEFR monitoring
Hops	Brewery chemist	[51]	1	NA	+	ND	ND	ND	
Baby's breath	Florist	[52]	1	NA	+	+	Histamine release	+	
Freesia and paprika	Horticulture	[53]	2	NA	+	+	Histamine release	ND	
Mushroom	Mushroom soup processors	[54]	8	NA	+	ND	ND	50% of 8 +	
Cocoon seed	Decorator	[55]	1	NA	+	ND	ND	ND	
Chicory	Chicory grower	[56]	1	NA	Neg	ND	ND	ND	FEV <sub>1</sub> , PC <sub>20</sub>
Rose hips	Pharmaceutical	[57]	9	NA	67% +	67% +	ND	50% of 4 +	
Sunflower	Laboratory workers	[58]	1	NA	+	+	RAST inhibition	+	
Garlic dust	Food packaging	[59]	1	NA	+	+	ND	+	
		[60]	1	NA	+	+	RAST inhibition	+	
Lycopodium	Powder	[61]	30	7	ND	ND	ND	2/2 +	
Sericin	Hairdresser	[62]	2	NA	1/1+	ND	ND	ND	
Nacre dust	Nacre buttons	[63]	1	NA	+	ND	Neg precipitin	+	
Pectin	Christmas candy maker	[64]	1	NA	+	-	Specific IgG <sub>4</sub>	+	
Henna (conchiolin?)	Hairdressers	[65]	2	NA	+	+	ND	1/2 +	
Neurospora	Plywood factory worker	[66]	1	NA	+	+	ND	+	
<b>Biologic enzymes</b>									
<i>B. subtilis</i>	Detergent industry	[67]	1642	3.2 (over 7 yrs)	4.5-75% +	26% of 248 +	ND	ND	Lung function
		[68]	38	NA	66% +	ND	Passive transfer 100% of 5 + precipitin (nonspecific)	90% +	Lung function
		[69]	667 14	NA 29	ND +	5% +	ND ND	ND 75% of 4 +	
Trypsin	Plastic, pharmaceutical	[69]	14	NA 29	ND +	5% +	ND ND	ND 75% of 4 +	
Papain	Pharmaceutical	[70]	29	45	34% +	34% +	ND	89% of 9 +	
Pepsin	Pharmaceutical	[71]	1	NA	+	+	ND	+	
Pancreatin	Pharmaceutical	[72]	14	NA	93% +	100% of 3 +	ND	100% of 8 +	Lung function
Flaviastase	Pharmaceutical	[73]	3	NA	+	+	+ precipitin	ND	
Bromelin	Pharmaceutical	[74]	76	11	25% +	ND	ND	ND	
		[75]	2	NA	+	ND	ND	2/2+	
Egg lysosyme	Pharmaceutical	[76]	1	NA	+	+	ND	+	+ PEFR monitoring
Fungal amylase	Bakers	[77]	118	NA	100% of 10 +	2% exposed + 34% occup. asthma +	ND	ND	
		[78]	1	NA	+	+	ND	+	
Fungal amyloglu- cosidase and hemicellulase	Bakers	[79]	140	NA	ND	5-24%	ND	ND	
Esperase	Detergent industry	[80]	667	NA	ND	5%	ND	ND	
<b>Vegetable gums</b>									
Acacia	Printers	[81]	63	19% of 31 (selection)	ND	ND	ND	ND	
		[82]	10	NA	+	ND	passive transfer (3+)	ND	
Tragacanth	Gum importer	[83]	1	NA	+	ND	ND	ND	
Karaya	Hairdressers	[84]	9	4	+	ND	Passive transfer	ND	
Guar	Carpet manufacturing	[85]	162	2	5% +	8% +	ND	67% of 3 +	PC <sub>20</sub>
<b>Other</b>									
Crab	Snow-crab processors	[86]	303	16	22% +	ND	ND	72% of 46 +	PEFR + PC <sub>20</sub> monitoring
Prawn	Prawn processors	[87]	50	36	26% +	16% +	ND	2/2 +	
Hoya	Oyster farm	[88]	1413	29	82% of 511 with asthma +	89% of ~180 with asthma +	ND	ND	
Cuttle-fish	Deep-sea fishermen	[89]	66	Incidence of 1%/yr	ND	ND	ND	ND	
Trout (?)	Trout-processors	[90]	5	NA	ND	100% neg	100% +	ND	
Shrimp meal	Technician	[91]	1	NA	+	+	ND	+	
Larva of silkworm	Sericulture	[92]	5519	0.2	100% of 9 (?) +	1/1 (?) +	P-K reaction	100% of 9 +	
Egg protein	Egg producers	[93]	188	7	34% +	29% +	ND	ND	PEFR, 7% +
Fish-feed (Echinodorus larva)	Aquarium keeper	[94]	1	NA	+	+	ND	+	
Red soft coral	Fishermen	[95]	74	9	2/2 +	ND	ND	ND	

For abbreviations see beginning of table. References are placed at the end of the table.

Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
<b>Low molecular weight agents</b>									
<b>Diisocyanates</b>									
Toluene diisocyanate	Polyurethane	[96]	112	12.5	3% +	0% +	0% + PCA	45% of 11 +	
	Plastics, varnish	[97]	26		ND	19% +	ND	100% +	
		[98]	195	28.0	ND	5% +	ND	70% of 17 +	
		[99]	91			ND	0% +	ND	
Diphenylmethane diisocyanate	Foundry	[100]	162 <sup>†</sup>		ND	ND	Specific IgG ND	57% +	
		[101]	11		ND	27% +	36% +	55% +	
		[102]	76	13.2	ND	3% +	7% +	ND	
		[103]	26	27.0	4% +	4% +	Specific IgG 15% + specific IgG	ND	
1,5 naphthylene diisocyanate	Manuf. rubber	[104]	3		ND	ND	ND	100% +	
Isophorone diisocyanate	Spray painter	[105]	1		ND	ND	ND	+	
Prepolymers of TDI	Floor varnishers	[106]	2	NA	ND	0% +	Specific IgG-	+	
Prepolymers of HDI	Spray painters	[107]	9	45%	ND	33% +	56% +	+	
<b>Combination of diisocyanates</b>									
TDI, MDI, HDI, PPI	Paint shop	[108]	51	11.8*	ND	ND	ND	60% of 10 + to PPI	
TDI, MDI, HDI	Various indust.	[109]	24		ND	ND	ND	70% + to TDI 33% to MDI 33% of 9 + to HDI	
		[110]	247 <sup>†</sup>		60% of 53 + 14%	ND	ND		
		[111]	62		ND	15% +	47% + specific IgG	6% to TDI 16% to MDI 24% to HDI	
TDI, MDI		[112]	28		ND TDI-HSA 83% of 6 + MDI-HSA	27% of 22 +	ND	100% +	
<b>Anhydrides</b>									
Phthalic anhydride	Plastics	[113]	1		+	+	ND	+	
	Tool setter, resin plant agent	[114]	3		ND	ND	ND	100% +	
	Production of resins	[115]	118	28	18% of 11 +	ND	ND	ND	
		[116]	60	14	ND	7% +	7% +	ND	
Trimellitic	Epoxy resins, plastics	[117]	4		100% +	75% +	specific IgG 100% + Specific IgG	100% of 1 +	
Tetrachlorophthalic anhydride	Epoxy resins, plastics	[118]	5		ND	ND	ND	100% +	
		[119]	7		100% +	100% +	ND	100% +	
Pyromellitic dianhydride	Epoxy adhesive	[120]	7		ND	ND	ND	30% +	
Methyl tetrahydrophthalic anhydride	Curing agent	[121]	1		+	+	specific IgG ND	ND	Improvement with removal PEFR monitoring +
Hexahydrophthalic anhydride	Chemical worker	[122]	1	NA	ND	ND Evidence	ND Test	+	
Himic anhydride	Manufacture of flame retardent	[123]	20	35	ND	40% of 7 +	RAST inhibition	ND	
<b>Aliphatic amines</b>									
<b>Ethyleneamines</b>									
Ethylene diamine	Shellac handlers	[124]	7		100% +	ND	ND	100% +	
	Photography	[125]	1		ND	ND	ND	+	
Hexamethylene tetramine	Lacquer handlers	[124]	7		100% +	ND	ND	100% +	
Triethylene tetramine	Aircraft fitter	[114]	1		ND	ND	ND	+	
Mixture of trimethyl-hexanediamine and isophorondiamine	Floor covering material salesman	[126]	1	NA	-	ND	ND	+	+ BAL
<b>Ethanolamines</b>									
Monoethanolamine	Beauty culture	[124]	10	100% +	ND	ND	100% +		
Aminoethyl-ethanolamine	Soldering	[127]	3		ND	ND	ND	100% +	
	Cable jointer	[128]	2		ND	ND	ND	+	
Dimethylethanolamine	Spray paint	[129]	1		-	ND	ND	+	

For abbreviations see beginning of table. References are placed at the end of the table.



Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
<b>Other</b>									
(3-DMAPA)	Ski manufacture	[130]	34	11.7	ND	ND	ND	ND	Cross-shift changes in FEV <sub>1</sub>
<b>Heterocyclic amines</b>									
Piperazine hydrochloride	Chemist	[131]	2		50%	ND	ND	100%	
	Pharmaceutical	[132]	131	11.4	ND	ND	ND	100% of 1 +	
	Chemical plant	[133]	2		50% +	100% +	ND	ND	
N-methylmorpholine		[134]	48	16.6**	ND	ND	ND	ND	
<b>Aromatic amines</b>									
Paraphenylene diamine	Fur dyeing	[135]	80	37.0	66% +	ND	ND	74% +	
<b>Mixture of amines</b>									
EPO 60	Mould maker	[136]	1	NA	ND	ND	ND	+	
<b>Fluxes</b>									
Colophony	Electronic workers	[137]	34		ND	ND	ND	100% +	
	Manufacture solder flux	[138]	68 low 14 med 6 high	4 21 21	ND ND ND	ND ND ND	ND ND ND	ND ND ND	
Zinc chloride and ammonium chloride flux	Metal jointing	[139]	2		ND	ND	ND	+	Changes in PC <sub>20</sub>
95% alkylaral polyether alcohol +5% polypro- pylene glycol	Electronic assembler	[140]	1		ND	ND	ND	+	
<b>Wood dust or bark</b>									
Western red cedar ( <i>Thuja plicata</i> )	Carpentry	[141]	35		ND	ND	ND	ND	Improvement on removal
	Furniture making	[142]	1320	3.4	1.9% +	ND	ND	ND	
	Cabinet-making, carpentry	[143]	22		100% -	ND	100% -	82% +	
	Sawmill	[144]	185		100% -	ND	ND	100% +	
		[145]	652	4.1	100% -	ND	ND	ND	Questionnaire
California redwood ( <i>Sequoia sempervirens</i> )	Woodcarvers	[146]	2		-	ND	-	+	
	Carpenter	[147]	1		ND	ND	precipitin ND	+	
Cedar of Lebanon ( <i>Cedra libani</i> )		[148]	6		17% +	ND	100% - precipitin	ND	
Cocabolla ( <i>Dalbergia retusa</i> )		[149]	3		100% -	ND	ND	ND	Improvement on removal
Iroko ( <i>Chlorophora excelsa</i> )	Carpenter	[150]	1		+	ND	+	+	
		[151]	1		ND	ND	precipitin ND	+	
Oak ( <i>Quercus robur</i> )		[152]	1		-	ND	+	+	
		[152]	1		-	ND	precipitin +	+	
Mahogany ( <i>Shorea</i> Sp.)		[152]	1		-	ND	precipitin +	+	
Abiruana ( <i>Pouteria</i> )		[153]	2		+	ND	precipitin -	+	
African maple ( <i>Triplochiton scleroxylon</i> )		[154]	2		+	+	precipitin Passive transfer	+	
Tanganyika aningre		[155]	3		100% +	100% -	100% - precipitin	100% +	
Central American walnut ( <i>Juglans olanchana</i> )		[156]	1		-	-	- precipitin	+	
Kejaat ( <i>Pterocarpus angolensis</i> )		[157]	1		+	ND	ND	ND	
African zebra-wood ( <i>Microberlinia</i> )		[158]	1		+	+	ND	+	
Ramin ( <i>Gonystylus bancanus</i> )	Woodworker	[159]	2		+	+	ND	+	
Quillaja bark	Factory to produce Saponin	[160]	1		ND	+	ND	+	
Fernambouc ( <i>Caesalpinia echinata</i> )	Bow-making	[161]	36	33.3	100% -	ND	ND	100% of 1 +	
Ashwood ( <i>Fraxinus americana</i> )	Sawmill	[162]	1		-	-	ND	+	

For abbreviations see beginning of table. References are placed at the end of the table.

Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
Pau Marfim ( <i>Balfourodendron riedelianum</i> )	Wood-worker	[163]	1	NA	+	+	ND	+	ND
Capreua ( <i>Myyrocarpus fastigiatus Fr. All.</i> )	Parquet floor layer	[164]	1	NA	ND	ND	ND	+	ND
Eastern white cedar ( <i>Thuja occidentalis</i> )	Sawmill	[165]	1		ND	-	ND	+	PEFR recording
Ebony wood ( <i>Diospyros crassiflora</i> )		[166]	1		-	ND	ND	+	
Kotibe wood ( <i>Nesorgordonia papveifera</i> )		[167]	1		+	ND	Passive transfer	+	
Cinnamon ( <i>Cinnamomum zeylanicum</i> )		[168]	40	22.5	ND	ND	ND	100% of 1+	
?	Sawmills of Eastern Canada and USA	[169]	11	ND	ND	ND	ND	+	PEFR monitoring
<b>Metals</b>									
Platinum	Platinum	[170]	16		62% +	ND	ND	62% +	
	refinery	[171]	136	29	17% +	21% +	ND	ND	
Nickel	Metal plating	[172]	1		+	ND	-	+	
		[173]	1		+	ND	precipitin -	+	
		[174]	1		+	+	precipitin ND	+	
Cobalt	Hard metal grinders	[175]	4		25% +	ND	ND	50% +	
	Diamond polisher	[176]	3		ND	ND	ND	100% +	
Zinc fumes	Solderers	[177]	2		ND	ND	ND	+	
	Locksmith	[178]	1		ND	ND	ND	+	
Tungsten carbide	Grinder	[179]	1		ND	ND	ND	ND	Recovery on removal
Chromium	Printer	[180]	1		+	ND	ND	ND	
	Plater	[181]	1		+	ND	ND	ND	
Chromium and nickel	Welder	[182]	5		ND	ND	ND	100% of 2 +	
	Tanning	[183]	1		-	+	ND	+	
Cobalt and nickel		[184]	8		75% + cobalt 62% + nickel	62% + cobalt 50% + nickel	ND	100% + to both cobalt and nickel	
<b>Drugs</b>									
Penicillins and ampicillin	Pharmaceutical	[185]	4		100% -	ND	ND	75% +	
Penicillamine	Pharmaceutical	[186]	1		ND	-	ND	+	PEFR recording
Cephalosporins	Pharmaceutical	[187]	2		+	ND	ND	+	
	Pharmaceutical	[188]	91	7.7	7.1% +	ND	ND	ND	Improvement off-work
Phenylglycine acid chloride	Pharmaceutical	[189]	24	29	37% +	37% +	Passive transfer	100% of 2 +	
Psyllium	Laxative manuf.	[190]	3		100% +	ND	ND	60% +	
	Pharmaceutical	[191]	130	4*	19% of 120 + 26% of 118 +	ND	27% of 18 +		
	Nurses	[192]	5		80% +	100% +	ND	100% +	
	Health personnel	[193]	193	4*	3% +	12% of 162 +	ND	26% of 15 +	
Methyl dopa	Pharmaceutical	[194]	1		-	ND	ND	+	
Spiramycin	Pharmaceutical	[195]	1		+	ND	ND	+	
	Pharmaceutical	[196]	51	7.8*	100% -	ND	ND	25% of 12 +	
	Pharmaceutical	[197]	2		ND	-	ND	+	
Salbutamol intermediate	Pharmaceutical	[198]	1		-	ND	ND	+	
Amprolium	Poultry feed mixer	[199]	1		ND	ND	ND	+	
Tetracycline	Pharmaceutical	[200]	1		ND	ND	ND	+	
	Pharmaceutical	[201]	1		+	ND	ND	+	
Isonicotinic acid hydrazide	Hospital pharmacy	[202]	1		+	+	ND	+	
Hydralazine	Pharmaceutical	[203]	1		-	-	-	+	
							Specific IgG		
Tylosin tartrate	Pharmaceutical	[204]	1		ND	ND	ND	-	
Ipecacuanha	Pharmaceutical	[205]	42	47.6	52% of 19 +	66% of 18 +	ND	ND	
Cimetidine	Pharmaceutical	[206]	4		ND	ND	ND	25% +	
Opiate compounds	Pharmaceutical	[207]	39	26%	+	ND	ND	ND	PEFR pre-post shift FEV <sub>1</sub>

For abbreviations beginning end of table. References are placed at the end of the table.

Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
<b>Chemicals</b>									
Chloramine T	Chemical manuf.	[208]	6		100% +	ND	66% + passive transfer	ND	
	Brewery	[209]	7		100% +	ND		ND	ND
Polyvinyl chloride (fumes)	Janitor-cleaning	[210]	5		100% of 4 +	ND	ND	100% of 3 + 27% of 11+	
	Meat wrapper	[211]	96	69.0	ND	ND	ND		
	Meat wrapper	[212]	3		ND	ND	ND		History only
Ethylcyanoacrylate ester	Manuf. bottle caps	[213]	1		ND	ND	ND	+	PEFR recording
	Building airplane models	[214]	1		ND	ND	ND	+	
Organic phosphate insecticides	Chemical pack- aging plant	[215]	1		ND	ND	ND	ND	History only
Levafix brilliant yellow E36	Pre dye solution	[216]	1		+	ND	ND	+	
Drimaren brilliant									
Yellow K-3GL	Textile industry	[216]	1		+	ND	ND	+	
Cibachrome brilliant scarlet 32	Textile industry	[216]	1		+	ND	ND	+	
Drimaren brilliant blue K-BL	Textile industry	[216]	1		+	ND	Evidence ND	Test +	
Reactive dyes	Reactive dyes manufacture	[217]	309	25.2	7% + orange	17% + orange	ND	65% of 20 +	
				8% + black	17% + black				
Lanasol yellow 4G	Dyer	[218]	1	NA	+	ND	ND	+	
Persulphate salts & henna	Hairdressing	[219]	2		+	ND	ND	+	
	Hairdressing	[220]	2		+	ND	ND	+	
	Hairdressing	[221]	23	17.4	4% +	ND	ND	100% of 4 +	
	Hairdressing	[222]	1		-	ND	ND	+	
	Hairdressing	[223]	1		ND	ND	ND	+	
Azodicarbonamide (azobisformamide)	Plastics, rubber	[224]	151	18.5	ND	ND	ND	ND	Removal with improvement
	Plastic	[225]	2		ND	ND	ND	+	
Diazonium salt	Plastics	[226]	4		ND	ND	ND	100% of 2 +	
	Manuf. of photocopy paper	[227]	1		ND	ND	ND	+	
Hexachlorophene (sterilizing agent)	Manuf. of fluorine polymer precursor	[228]	45	55.5	ND	20% +	ND	100% of 2	
	Hospital staff	[229]	1		ND	ND	ND	+	
Formaldehyde	Hospital staff	[230]	28	29*	ND	ND	ND	50% of 4 +	
	Different indust.	[231]	15		ND	ND	ND	60% +	
		[232]	230	5.2	ND	ND	ND	5% +	
Urea formaldehyde	Resin	[233]	2		-	ND	ND	+	
Freon	Manuf. of foam	[234]	1		ND	ND	ND	+	
	Refrigeration	[235]	1		ND	ND	ND	+	
Furfuryl alcohol (furan based resin)	Foundry mold making	[236]	1		ND	ND	ND	+	
	Wool dye house	[237]	6		ND	83% +	100% +	ND	
Styrene	Plastics factory	[238]	2		-	ND	ND	+	
Glutaraldehyde	Hospital endoscopy unit	[239]	9	88.8	ND	ND	ND	ND	Questionnaire
Methyl methacrylate and cyanocrylates	Adhesive	[240]	7		ND	ND	ND	86% +	PEFR 14% +
	Nurse	[241]	1		ND	ND	ND	+	
Iso-nonyl oxybenzene sulphonate	Lab. technician	[242]	1		ND	ND	ND	+	
Chlorhexidine	Nurse	[243]	2		ND	ND	ND	+	
Tetrazene	Detonator manuf.	[244]	1		ND	ND	ND	+	+ PEFR recording
Ethylene dioxide	Nurse	[245]	1	NA	ND	+	ND	+	Changes in PC <sub>20</sub>
Polyethylene	Paper packer	[246]	1	NA	ND	ND	ND	+	+ PEFR recording
Tetrachloro- isophthalonitrile (fungicide)	Farmer	[247]	1	NA	ND	-	+ patch test	+	FEV <sub>1</sub> recording at work
Tributyl tin oxide (fungicide)	Venipuncture technician	[248]	1	NA	-	ND	ND	+	
Tall oil (pine resin)	Rubber tyre manufacturer	[249]	1	NA	-	ND	-patch test	+	+ PEFR recording

**Synthetic materials** See beginning of table. References are placed at the end of the table.

Table 1. continued

Agents	Occupation	[Ref]	Subjects n	Prevalence %	Skin test	Specific IgE	Other immunologic	Broncho- provocation test	Other evidence
Plexiglass		[250]	1		ND	ND	ND	+	
Latex	Glove manuf.	[251]	81	6	11% +	ND	ND	ND	PEFR recording, Pre-post shift change in FEV <sub>1</sub>
<b>Unidentified</b>									
(?)	Resp. therapist	[252]	194	18.7	ND	ND	ND	ND	Questionnaire
(?)	Mineral analysis laboratory	[253]	21	23.8*	ND	ND	ND	ND	Questionnaire PC <sub>20</sub>
(?) Oil mists	Toolsetter	[254]	1		ND	ND	ND	+	PEFR recording
(?) Fluorine	Potroom	[255]	52		ND	ND	ND	ND	History
(?) Aluminum	Potroom	[256]	227	7.0	ND	ND	ND	ND	Questionnaire
	Potroom	[257]	35		ND	ND	ND	ND	History
	Potroom	[258]	57		ND	ND	ND	ND	History

For abbreviations beginning end of table. References are placed at the end of the table.

### Table References

- Venables KM, Tee RD, Hawkins ER, *et al.* Laboratory animal allergy in a pharmaceutical company. *Br J Ind Med* 1988; 45: 660–666.
- Newman-Taylor AJ, Longbottom JL, Pepys J. Respiratory allergy to urine proteins of rats and mice. *Lancet* 1977; 847–849.
- Mäntyjärvi J, Ylönen R, Taivainen A, Virtanen T. IgG and IgE antibody responses to cow dander and urine in farmers with cow-induced asthma. *Clin Exp Allergy* 1992; 22: 83–90.
- Bar-Sela S, Teichtahl H, Lutsky I. Occupational asthma in poultry workers. *J Allergy Clin Immunol* 1984; 73: 271–275.
- Lutsky I, Teichtahl H, Bar-Sela S. Occupational asthma due to poultry mites. *J Allergy Clin Immunol* 1984; 73: 56–60.
- Brennan NJ. Pig Butcher's asthma: case report and review of the literature. *Irish Med J* 1985; 78: 321–322.
- Armentia A, Martin-Santos J, Subiza J, *et al.* Occupational asthma due to frogs. *Ann Allergy* 1988; 60: 209–210.
- Moneret-Vautrin DA, Pupil P, Courtine D, Grilliat JP. Asthme professionnel aux protéines du lactosérum. *Rev Fr Allergol* 1984; 24: 93–95.
- Olaguibel JM, Hernandez D, Morales P, Peris A, Basomba A. Occupational asthma caused by inhalation of casein. *Allergy* 1990; 45: 306–308.
- El-Ansary EH, Gordon DJ, Tee RD, Newman-Taylor AJ. Respiratory allergy to inhaled bat guano. *Lancet* 1987; i: 316–318.
- Cuthbert OD, Jeffrey IG, McNeill HB, Wood J, Topping MD. Barn allergy among Scottish farmers. *Clin Allergy* 1984; 14: 197–206.
- Blainey AD, Topping MD, Ollier S, Davies RJ. Allergic respiratory disease in grain workers: the role of storage mites. *J Allergy Clin Immunol* 1989; 84: 296–303.
- Burge PS, Edge G, O'Brien IM, Harries MG, Hawkins R, Pepys J. Occupational asthma in a research centre breeding locusts. *Clin Allergy* 1980; 10: 355–363.
- Tee RD, Gordon DJ, Hawkins ER, *et al.* Occupational allergy to locusts: an investigation of the sources of the allergen. *J Allergy Clin Immunol* 1988; 81: 517–525.
- Gibbons HL, Dille JR, Cowley RG. Inhalant allergy to the screw worm fly. *Arch Environ Health* 1965; 10: 424–430.
- Bagenstose AH, Mathews KP, Homburger HA, Saaveard-Delgado AP. – Inhalant allergy due to crickets. *J Allergy Clin Immunol* 1980; 65: 71–74.
- Stevenson DD, Mathews KP. Occupational asthma following inhalation of moth particles. *J Allergy* 1967; 39: 274–283.
- Randolph H. Allergic reaction to dust of insect origin. *J Am Med Assoc* 1934; 103: 560–562.
- Wittich FW. Allergic rhinitis and asthma due to sensitization to the Mexican bean weevil (*Zabrotes subfasciatus boh.*). *J Allergy* 1940; 12: 42–45.
- Spieksma FTM, Vooren PH, Kramps JA, Dijkman JH. Respiratory allergy to laboratory fruit flies (*Drosophila melanogaster*). *J Allergy Clin Immunol* 1986; 77: 108–113.
- Ostrom NK, Swanson MC, Agarwal MK, Yunginger JW. Occupational allergy to honeybee-body dust in a honey-processing plant. *J Allergy Clin Immunol* 1986; 77: 736–740.
- Siracusa A, Verga A, Bacoccoli R, Fabbri A, Felicioni D. L'asma da bigattini (Larve della mosca camaria): studio clinico e immunologico. *Med Lav* 1989; 80: 489–497.
- Schroeckenstein DC, Meier-Davis S, Graziano FM, Falomo A, Bush RK. Occupational sensitivity to *Alphitobius diaperinus* (Panzer) (lesser mealworm). *J Allergy Clin Immunol* 1988; 82:1081–1088.
- Lutsky I, Bar-Sela S. Northern fowl mite (*Ornithonyssus sylvianum*) in occupational asthma of poultry workers. *Lancet* 1982; 2: 874–875.
- Cuthbert OD, Brostoff J, Wraith DG, Brighton WD. "Barn allergy": asthma and rhinitis due to storage mites. *Clin Allergy* 1979; 9: 229–236.
- Granel-Tena C, Cistero-Bahima A, Olive-Perez A. Allergens in asthma and baker's rhinitis. *Alergia* 1985; 32: 69–73.
- Michel FB, Guin JJ, Seignalet C, *et al.* Allergie à *Panonychus ulmi* (Koch). *Rev Fr Allergol* 1977; 17: 93–97.
- Meister W. Professional asthma owing to *Daphnia* allergy. *Allerg Immunol (Leipz)* 1978; 24: 191–193.
- Axelsson IGK, Johansson SGO, Zetterstrom O. Occupational allergy to weeping fig in plant keepers. *Allergy* 1987; 42: 161–167.
- Kaufman GL, Gandevia BH, Bellas TE, Tovey ER, Baldo BA. Occupational allergy in an entomological research centre. I. Clinical aspects of reactions to the sheep blowfly *Lucilia cuprina*. *Br J Ind Med* 1989; 46: 473–478.

31. Uragoda CG, Wijekoon PMB. Asthma in silk workers. *J Soc Occup Med* 1991; 41: 140-142.
32. Chan-Yeung M, Schulzer M, MacLean L, Dorken E, Grzybowski S. Epidemiologic health survey of grain elevator workers in British Columbia. *Am Rev Respir Dis* 1980; 121: 329-338.
33. Williams N, Skoulas A, Merriman JE. Exposure to grain dust. I. A survey of the effects. *J Occup Med* 1964; 6: 319-329.
34. Skoulas A, Williams N, Merriman JE. Exposure to grain dust. II. A clinical study of the effects. *J Occup Med* 1964; 6: 359-372.
35. Chan-Yeung M, Wong R, MacLean L. Respiratory abnormalities among grain elevator workers. *Chest* 1979; 75: 461-467.
36. Musk AW, Venables KM, Crook B, et al. Respiratory symptoms, lung function, and sensitisation to flour in a British bakery. *Br J Ind Med* 1989; 46: 636-642.
37. Block G, Tse KS, Kijek K, Chan H, Chan-Yeung M. Baker's asthma. *Clin Allergy* 1983; 13: 359-370.
38. Sutton R, Skerritt JH, Baldo BA, Wrigley CW. The diversity of allergens involved in baker's asthma. *Clin Allergy* 1984; 14: 93-107.
39. Valdivieso R, Quirce S, Sainz T. Bronchial asthma caused by *Lathyrus sativus* flour. *Allergy* 1988; 43: 536-539.
40. Picon SJ, Carmona JGB, Sotillos MDMG. Occupational asthma caused by vetch (*Vicia sativa*). *J Allergy Clin Immunol* 1991; 88: 135-136.
41. Ordman D. Buckwheat allergy. *S Afr Med J* 1947; 21: 737-739.
42. Lachance P, Cartier A, Dolovich J, Malo J-L. Occupational asthma from reactivity to an alkaline hydrolysis derivative of gluten. *J Allergy Clin Immunol* 1988; 81: 385-390.
43. Jones RN, Hughes JM, Lehrer SB, et al. Lung function consequences of exposure and hypersensitivity in workers who process green coffee beans. *Am Rev Respir Dis* 1982; 125: 199-202.
44. Zuskin E, Valic F, Kanceljak B. Immunological and respiratory changes in coffee workers. *Thorax* 1981; 36: 9-13.
45. Osterman K, Johansson SGO, Zetterstrom O. Diagnostic tests in allergy to green coffee. *Allergy* 1985; 40: 336-343.
46. Panzani R, Johansson SGO. Results of skin test and RAST in allergy to a clinically potent allergen (castor bean). *Clin Allergy* 1986; 16: 259-266.
47. Cartier A, Malo JL. Occupational asthma due to tea dust. *Thorax* 1990; 45: 203-206.
48. Blanc PD, Trainor WD, Lim DT. Herbal tea asthma. *Br J Ind Med* 1986; 43: 137-138.
49. Gleich GJ, Welsh PW, Yunginger JW, Hyatt RE, Catlett JB. Allergy to tobacco: an occupational hazard. *N Engl J Med* 1980; 302: 617-619.
50. Lander F, Gravesen S. Respiratory disorders among tobacco workers. *Br J Ind Med* 1988; 45: 500-502.
51. Newmark FM. Hops allergy and terpene sensitivity: an occupational disease. *Ann Allergy* 1978; 41: 311-312.
52. Twiggs JT, Yunginger JW, Agarwal MK, Reed CE. Occupational asthma in a florist caused by the dried plant, baby's breath. *J Allergy Clin Immunol* 1982; 69: 474-477.
53. van Toorenenbergen AW, Dieges PH. Occupational allergy in horticulture: demonstration of immediate-type allergic reactivity to freesia and paprika plants. *Int Arch Allergy Appl Immunol* 1984; 75: 44-47.
54. Symington IS, Kerr JW, McLean DA. Type I allergy in mushroom soup processors. *Clin Allergy* 1981; 11: 43-47.
55. Rubin JM, Duke MB. Unusual cause of bronchial asthma. Cocoon seed used for decorative purposes. *NY State J Med* 1974; 74 (3): 538-539.
56. Nemery B, Demedts M. Occupational asthma in a chicory grower. *Lancet* 1989; i: 672-673.
57. Kweselov A, Rowe M, Sears-Ewald D, Ownby D. Rose hips: a new occupational allergen. *J Allergy Clin Immunol* 1990; 85: 704-708.
58. Bousquet OJ, Dhivert H, Clauzel AM, Hewitt B, Michel FB. Occupational allergy to sunflower pollen. *J Allergy Clin Immunol* 1985; 75: 70-75.
59. Falleroni AE, Zeiss CR, Levitz D. Occupational asthma secondary to inhalation of garlic dust. *J Allergy Clin Immunol* 1981; 68: 156-160.
60. Lybarger JA, Gallagher JS, Pulver DW, Litwin A, Brooks S, Bernstein IL. Occupational asthma induced by inhalation and ingestion of garlic. *J Allergy Clin Immunol* 1982; 69: 448-454.
61. Catilina P, Chamoux A, Gabrillargues D, Catilina MJ, Royfe MH, Wahl D. Contribution à l'étude des asthmes d'origine professionnelle: l'asthme à la poudre de lycopode. *Arch Mal Prof* 1988; 49: 143-148.
62. Charpin J, Blanc M. Une cause nouvelle d'allergie professionnelle chez les coiffeuses: l'allergie à la séricine. *Marseille Médical* 1967; 104: 169-170.
63. Zedda S. A case of bronchial asthma from inhalation of nacre dust. *Med Lav* 1967; 58: 459-464.
64. Kraut A, Peng Z, Becker AB, Warren CPW. Christmas candy maker's asthma. IgG<sub>4</sub>-mediated pectin allergy. *Chest* 1992; 102: 1605-1607.
65. Starr JC, Yunginger J, Brahser GW. Immediate type I asthmatic response to henna following occupational exposure in hairdressers. *Ann Allergy* 1982; 48: 98-99.
66. Côté J, Chan H, Brochu G, Chan-Yeung M. Occupational asthma caused by exposure to neurospora in a plywood factory worker. *Br J Ind Med* 1991; 48: 279-282.
67. Juniper CP, How MJ, Goodwin BFJ. *Bacillus subtilis* enzymes: a 7-year clinical, epidemiological and immunological study of an industrial allergen. *J Soc Occup Med* 1977; 27: 3-12.
68. Franz T, McMurray KD, Brooks S, Bernstein IL. Clinical, immunologic and physiologic observations in factory workers exposed to *B. subtilis* enzyme dust. *J Allergy* 1971; 47: 170-179.
69. Colten HR, Polakoff PL, Weinstein SF, Strieder DJ. Immediate hypersensitivity to hog trypsin resulting from industrial exposure. *N Engl J Med* 1975; 292: 1050-1053.
70. Baur X, König G, Benze K, Fruhmann G. Clinical symptoms and results of skin test, RAST and bronchial provocation test in thirty three papain workers: evidence for strong immunogenic potency and clinically relevant "proteolytic effects of airborne papain". *Clin Allergy* 1982; 12: 9-17.
71. Cartier A, Malo J-L, Pineau L, Dolovich J. Occupational asthma due to pepsin. *J Allergy Clin Immunol* 1984; 73: 574-577.
72. Wiessmann KJ, Baur X. Occupational lung disease following long-term inhalation of pancreatic extracts. *Eur J Respir Dis* 1985; 66: 13-20.
73. Pauwels R, Devos M, Callens L, Van der Straeten M. - Respiratory hazards from proteolytic enzymes. *Lancet* 1978; i: 669.
74. Cortona G, Beretta F, Traina G, Nava C. Preliminary investigation in a pharmaceutical industry: bromelin-induced pathology. *Med Lav* 1980; 1: 70-75.
75. Galleguillos F, Rodriguez JC. Asthma caused by bromelin inhalation. *Clin Allergy* 1978; 8: 21-24.

76. Bernstein JA, Kraut A, Bernstein DI, Warrington R, Bolin T, Warren CPW. Occupational asthma induced by inhaled egg lysozyme. *Chest* 1993; 103: 532-535.
77. Baur X, Fruhmans G, Haug B, Rasche B, Reiher W, Weiss W. Role of *Aspergillus* amylase in baker's asthma. *Lancet* 1986; i: 43.
78. Bimbaum J, Latil F, Vervloet D, Senft M, Charpin J. Rôle de l'alpha-amylase dans l'asthme du boulanger. *Rev Mal Respir* 1988; 5: 519-521.
79. Baur X, Weiss W, Sauer W, et al. Baking components as a contributory cause of baker's asthma. *Dtsch Med Wschr* 1988; 113: 1275-1278.
80. Zachariae H, Høegh-Thomsen J, Witmeur O, Wide L. Detergent enzymes and occupational safety. Observations on sensitization during Esperase® production. *Allergy* 1981; 36: 513-516.
81. Fowler PBS. Printer's asthma. *Lancet* 1952; ii: 755-757.
82. Bohner CB, Sheldon JM, Trenis JW. Sensitivity to gum acacia, with a report of ten cases of asthma in printers. *J Allergy* 1941; 12: 290-294.
83. Gelfand HH. The allergenic properties of vegetable gums: a case of asthma due to tragacanth. *J Allergy* 1943; 14: 203-219.
84. Feinberg SM, Schoenkerman BB. Karaya and related gums as causes of atopy. *Wisconsin Med J* 1940; 39: 734.
85. Malo JL, Cartier A, L'Archevêque J, et al. Prevalence of occupational asthma and immunological sensitization to guar gum among employees at a carpet-manufacturing plant. *J Allergy Clin Immunol* 1990; 86: 562-569.
86. Cartier A, Malo JL, Forest F, et al. Occupational asthma in snow-crab processing workers. *J Allergy Clin Immunol* 1984; 74: 261-269.
87. Gaddie J, Legge JS, Friend JAR, Reid TMS. Pulmonary hypersensitivity in prawn workers. *Lancet* 1980; ii: 1350-1353.
88. Jyo T, Kohmoto K, Katsutani T, Otsuka T, Oka SD, Mitsui S. Hoya (sea-squirt) asthma. *Occupational Asthma*. London, Von Nostrand Reinhold, 1980; pp. 209-228.
89. Tomaszunas S, Weclawik Z, Lewinski M. Allergic reactions to cuttlefish in deep-sea fishermen. *Lancet* 1988; i: 1116-1117.
90. Sherson D, Hansen I, Sigsgaard T. Occupationally-related respiratory symptoms in trout-processing workers. *Allergy* 1989; 44: 336-341.
91. Carino M, Elia G, Molinini R, Nuzzaco A, Ambrosi L. Shrimp meal asthma in the aquaculture industry. *Med Lav* 1985; 76: 471-475.
92. Kobayashi S. Different aspects of occupational asthma in Japan. *Occupational asthma*. In: Frazier CA, ed. New York, Van Nostrand Reinhold Co. 1980; pp. 229-244.
93. Smith AB, Bernstein DI, London MA, et al. Evaluation of occupational asthma from airborne egg protein exposure in multiple settings. *Chest* 1990; 98: 398-404.
94. Resta O, Foschino-Barbaro MP, Carnimeo N, Napoli PL, Di Pavese I, Schino P. Occupational asthma from fish-feed. *Med Lav* 1982; 3: 234-236.
95. Onizuka R, Inoue K, Kamiya H. Red soft coral-induced allergic symptoms observed in spiny lobster fishermen. *Aerugi* 1990; 39: 339-347.
96. Butcher BT, Salvaggio JE, Weill H, Ziskind MM. Toluene diisocyanate (TDI) pulmonary disease: immunologic and inhalation challenge studies. *J Allergy Clin Immunol* 1976; 58: 89-100.
97. Butcher BT, O'Neil CE, Reed MA, Salvaggio JE. Radioallergosorbent testing of toluene diisocyanate-reactive individuals using p-tolyl isocyanate antigen. *J Allergy Clin Immunol* 1980; 66: 213-216.
98. Baur X, Fruhmans G. Specific IgE antibodies in patients with isocyanate asthma. *Chest* 1981; 80: 73S-76S.
99. Paggiaro PL, Filieri M, Loi AM, et al. Absence of IgG antibodies to TDI-HSA in a radioimmunological study. *Clin Allergy* 1983; 13: 75-79.
100. Mapp CE, Boschetto P, Dal Vecchio L, Maestrelli P, Fabbri LM. Occupational asthma due to isocyanates. *Eur Respir J* 1988; 1: 273-279.
101. Zammit-Tabona M, Sherkin M, Kijek K, Chan H, Chan-Yeung M. Asthma caused by diphenylmethane diisocyanate in foundry workers. Clinical, bronchial provocation, and immunologic studies. *Am Rev Respir Dis* 1983; 128: 226-230.
102. Tse KS, Johnson A, Chan H, Chan-Yeung M. A study of serum antibody activity in workers with occupational exposure to diphenylmethane diisocyanate. *Allergy* 1985; 40: 314-320.
103. Liss GM, Bernstein DI, Moller DR, Gallagher JS, Stephenson RL, Bernstein IL. Pulmonary and immunologic evaluation of foundry workers exposed to methylene diphenyldiisocyanate (MDI). *J Allergy Clin Immunol* 1988; 82: 55-61.
104. Harris MG, Burge PS, Samson M, Taylor AJ, Pepys J. Isocyanate asthma: respiratory symptoms due to 1,5 naphthylene diisocyanate. *Thorax* 1979; 34: 762-766.
105. Clarke CW, Aldons PM. Isophorone diisocyanate-induced respiratory disease (IPDI). *Aust NZ J Med* 1981; 11: 290-292.
106. Vandenplas O, Cartier A, Lesage J, Perrault G, Grammer LC, Malo JL. Occupational asthma caused by a prepolymer but not the monomer of toluene diisocyanate (TDI). *J Allergy Clin Immunol* 1992; 89: 1183-1188.
107. Vandenplas O, Cartier A, Lesage J, et al. Prepolymers of hexamethylene diisocyanate (HDI) as a cause of occupational asthma. *J Allergy Clin Immunol* 1993; 91: 850-861.
108. Séguin P, Allard A, Cartier A, Malo JL. Prevalence of occupational asthma in spray painters exposed to several types of isocyanates, including polymethylene polyphenylisocyanates. *J Occup Med* 1987; 29: 340-344.
109. O'Brien IM, Harries MG, Burge PS, Pepys J. Toluene diisocyanate-induced asthma. I. Reactions to TDI, MDI, HDI and histamine. *Clin Allergy* 1979; 9: 1-6.
110. Baur X, Dewair M, Fruhmans G. Detection of immunologically sensitized isocyanate workers by RAST and intracutaneous skin tests. *J Allergy Clin Immunol* 1984; 73: 610-618.
111. Cartier A, Grammer L, Malo JL, et al. Specific serum antibodies against isocyanates: association with occupational asthma. *J Allergy Clin Immunol* 1989; 84: 507-514.
112. Pezzini A, Riviera A, Paggiaro P, et al. Specific IgE antibodies in twenty eight workers with diisocyanate-induced bronchial asthma. *Clin Allergy* 1984; 14: 453-461.
113. Maccia CA, Bernstein IL, Emmett EA, Brooks SM. *In vitro* demonstration of specific IgE in phthalic anhydride hypersensitivity. *Am Rev Respir Dis* 1976; 113: 701-704.
114. Fawcett IW, Newman-Taylor AJ, Pepys J. Asthma due to inhaled chemical agents: epoxy resin systems containing phthalic acid anhydride, trimellitic acid anhydride and triethylene tetramine. *Clin Allergy* 1977; 7: 1-14.
115. Wernfors M, Nielsen J, Schütz A, Skerfving S. Phthalic anhydride-induced occupational asthma. *Int Arch Allergy Appl Immunol* 1986; 79: 77-82.
116. Nielsen J, Welinder H, Schütz A, Skerfving S. Specific

- serum antibodies against phthalic anhydride in occupationally exposed subjects. *J Allergy Clin Immunol* 1988; 82: 126-133.
117. Zeiss CR, Patterson R, Pruzansky JJ, Miller MM, Rosenberg M, Levitz D. Trimellitic anhydride-induced airway syndromes: clinical and immunologic studies. *J Allergy Clin Immunol* 1977; 60: 96-103.
  118. Schlueter DP, Banaszak EF, Fink JN, Barboriak J. Occupational asthma due to tetrachlorophthalic anhydride. *J Occup Med* 1978; 20: 183-187.
  119. Howe W, Venables KM, Topping MD, *et al.* Tetrachlorophthalic anhydride asthma: evidence for specific IgE antibody. *J Allergy Clin Immunol* 1983; 71: 5-11.
  120. Meadway J. Asthma and atopy in workers with an epoxy adhesive. *Br J Dis Chest* 1980; 74: 149-154.
  121. Nielsen J, Welinder H, Skerfving S. Allergic airway disease caused by methyl tetrahydrophthalic anhydride in epoxy resin. *Scand J Work Environ Health* 1989; 15: 154-155.
  122. Chee CBE, Lee HS, Cheong TH, Wang YT. Occupational asthma due to hexahydrophthalic anhydride: a case report. *Br J Ind* 1991; 48: 643-645.
  123. Rosenman KD, Bernstein DI, O'Leary K, Gallagher JS, D'Souza L, Bernstein IL. Occupational asthma caused by hemic anhydride. *Scand J Work Environ Health* 1987; 13: 150-154.
  124. Gelfand HH. Respiratory allergy due to chemical compounds encountered in the rubber, lacquer, shellac, and beauty culture industries. *J Allergy* 1963; 34: 374-381.
  125. Lam S, Chan-Yeung M. Ethylenediamine-induced asthma. *Am Rev Respir Dis* 1980; 121: 151-155.
  126. Aleva RM, Aalbers R, Koëter GH, de Monchy JGR. Occupational asthma caused by a hardener containing an aliphatic and cycloaliphatic diamine. *Am Rev Respir Dis* 1992; 145: 1217-1218.
  127. Pepys J, Pickering CAC. Asthma due to inhaled chemical fumes: amino-ethyl ethanolamine in aluminium soldering flux. *Clin Allergy* 1972; 2: 197-204.
  128. Sterling GM. Asthma due to aluminium soldering flux. *Thorax* 1967; 22: 533-537.
  129. Vallières M, Cockcroft DW, Taylor DM, Dolovich J, Hargreave FE. Dimethyl ethanolamine-induced asthma. *Am Rev Respir Dis* 1977; 115: 867-871.
  130. Sargent EV, Mitchell CA, Brubaker RE. Respiratory effects of occupational exposure to an epoxy resin system. *Arch Environ Health* 1976; 31: 236-240.
  131. Pepys J, Pickering CAC, Loudon HWG. Asthma due to inhaled chemical agents: piperazine dihydrochloride. *Clin Allergy* 1972; 2: 189-196.
  132. Hagmar L, Bellander T, Bergöo B, Simonsson BG. Piperazine-induced occupational asthma. *J Occup Med* 1982; 24: 193-197.
  133. Welinder H, Hagmar L, Gustavsson C. IgE antibodies against piperazine and N-methyl-piperazine in two asthmatic subjects. *Int Arch Allergy Appl Immunol* 1986; 79: 259-262.
  134. Belin L, Wass U, Audunsson G, Mathiasson L. Amines: possible causative agents in the development of bronchial hyperreactivity in workers manufacturing polyurethanes from isocyanates. *Br J Ind Med* 1983; 40: 251-257.
  135. Silberman DE, Sorrell AH. Allergy in fur workers with special reference to paraphenylenediamine. *J Allergy* 1959; 30: 11-18.
  136. Lamboum EM, Hayes JP, McAllister WA, Newman-Taylor AJ. Occupational asthma due to EPO 60. *Br J Ind Med* 1992; 49: 294-295.
  137. Burge PS, Harries MG, O'Brien I, Pepys J. Bronchial provocation studies in workers exposed to the fumes of electronic soldering fluxes. *Clin Allergy* 1980; 10: 137-149.
  138. Burge PS, Edge G, Hawkins R, White V, Taylor AN. Occupational asthma in a factory making flux-cored solder containing colophony. *Thorax* 1981; 36: 828-834.
  139. Weir DC, Robertson AS, Jones S, Burge PS. Occupational asthma due to soft corrosive soldering fluxes containing zinc chloride and ammonium chloride. *Thorax* 1989; 44: 220-223.
  140. Stevens JJ. Asthma due to soldering flux: a polyether alcohol-polypropylene glycol mixture. *Ann Allergy* 1976; 36: 419-422.
  141. Milne J, Gandevia B. Occupational asthma and rhinitis due to western (Canadian) red cedar. *Med J Aust* 1969; 2: 741-744.
  142. Ishizaki T, Sluda T, Miyamoto T, Matsumara Y, Mizuno K, Tomaru M. Occupational asthma from western red cedar dust (*Thuja plicata*) in furniture factory workers. *J Occup Med* 1973; 15: 580-585.
  143. Chan-Yeung M, Barton GM, Maclean L, Grzybowski S. Occupational asthma and rhinitis due to western red cedar (*Thuja plicata*). *Am Rev Respir Dis* 1973; 108: 1094-1102.
  144. Chan-Yeung M, Lam S, Koener S. Clinical features and natural history of occupational asthma due to western red cedar (*Thuja plicata*). *Am J Med* 1982; 72: 411-415.
  145. Chan-Yeung M, Vedal S, Kus J, Maclean L, Enarson D, Tse KS. Symptoms, pulmonary function, and bronchial hyperreactivity in western red cedar workers compared with those in office workers. *Am Rev Respir Dis* 1984; 130: 1038-1041.
  146. Chan-Yeung M, Abboud R. Occupational asthma due to California redwood (*Sequoia sempervirens*) dusts. *Am Rev Respir Dis* 1976; 114: 1027-1031.
  147. doPico GA. Asthma due to dust from redwood (*Sequoia sempervirens*). *Chest* 1978; 73: 424-425.
  148. Greenberg M. Respiratory symptoms following brief exposure to cedar of Lebanon (*Cedra libani*) dust. *Clin Allergy* 1972; 2: 219-224.
  149. Eaton KK. Respiratory allergy to exotic wood dust. *Clin Allergy* 1973; 3: 307-310.
  150. Pickering CAC, Batten JC, Pepys J. Asthma due to inhaled wood dusts - western red cedar and iroko. *Clin Allergy* 1972; 2: 213-218.
  151. Azofra J, Olaguibel JM. Occupational asthma caused by iroko wood. *Allergy* 1989; 44: 156-158.
  152. Sosman AJ, Schlueter DP, Fink JN, Barboriak JJ. Hypersensitivity to wood dust. *N Engl J Med* 1969; 281: 977-980.
  153. Booth BH, Lefoldt RH, Moffitt EM. Hypersensitivity to wood dust. *J Allergy Clin Immunol* 1976; 57: 352-357.
  154. Hinojosa M, Moneo I, Dominguez J, Delgado E, Losada E, Alcover R. Asthma caused by African maple (*Triplochiton scleroxylon*) wood dust. *J Allergy Clin Immunol* 1984; 74: 782-786.
  155. Paggiaro PL, Cantalupi R, Filieri M, *et al.* Bronchial asthma due to inhaled wood dust: *Tanganyika aningre*. *Clin Allergy* 1981; 11: 605-610.
  156. Bush RK, Clayton D. Asthma due to central american walnut (*Juglans olanchana*) dust. *Clin Allergy* 1983; 13: 389-394.
  157. Ordman D. Wood dust as an inhalant allergen. Bronchial asthma caused by kejaat wood (*Pterocarpus angolensis*). *S Afr Med* 1949; 23: 973-975.
  158. Bush RK, Yunginger JW, Reed CE. Asthma due to

- African zebrawood (*Microberlinia*) dust. *Am Rev Respir Dis* 1978; 117: 601-603.
159. Hinojosa M, Losada E, Moneo I, Dominguez J, Carrillo T, Sanchez-Cano M. Occupational asthma caused by African maple (Obeche) and Ramin: evidence of cross-reactivity between these two woods. *Clin Allergy* 1986; 16: 145-153.
  160. Raghuprasad PK, Brooks SM, Litwin A, Edwards JJ, Bernstein IL, Gallagher J. Quillaja bark (soapbark)-induced asthma. *J Allergy Clin Immunol* 1980; 65: 285-287.
  161. Hausen BM, Herrmann B. Bow-makers disease: an occupational disease in the manufacture of wooden bows for string instruments. *Dtsch Med Wochenschr* 1990; 115: 169-173.
  162. Malo JL, Cartier A. Occupational asthma caused by exposure to ash wood dust (*Fraxinus americana*). *Eur Respir J* 1989; 2: 385-387.
  163. Basomba A, Burches E, Almodovar A, Rojas D, Hernandez F de. Occupational rhinitis and asthma caused by inhalation of *Balfourodendron riedelianum* (Pau Marfim) wood dust. *Allergy* 1991; 46: 316-318.
  164. Innocenti A, Romeo R, Mariano A. Asthma and systemic toxic reaction due to cabreuva (*Myrocarpus fastigiatus* Fr. All.) wood dust. *Med Lav* 1991; 82: 446-450.
  165. Cartier A, Chan H, Malo JL, Pineau L, Tse KS, Chan-Yeung M. Occupational asthma caused by Eastern white cedar (*Thuja occidentalis*) with demonstration that plicatic acid is present in this wood dust and is the causal agent. *J Allergy Clin Immunol* 1986; 77: 639-645.
  166. Maestrelli P, Marcer G, Dal Vecchio L. Occupational asthma due to ebony wood (*Diospyros crassiflora*) dust. *Ann Allergy* 1987; 59: 347-349.
  167. Reques FG, Fernandez RP. Asthme professionnel à un bois exotique. *Nesorgordonia papaverifera* (danta ou kotibe). *Rev Mal Respir* 1988; 5: 71-73.
  168. Uragoda CG. Asthma and other symptoms in cinnamon workers. *Br J Ind Med* 1984; 41: 224-227.
  169. Malo JL, Cartier A, Boulet LP. Occupational asthma in sawmills of Eastern Canada and United States. *J Allergy Clin Immunol* 1986; 78: 392-398.
  170. Pepys J, Pickering CAC, Hughes EG. Asthma due to inhaled chemical agents: complex salts of platinum. *Clin Allergy* 1972; 2: 391-396.
  171. Brooks SM, Baker DB, Gann PH, et al. Cold air challenge and platinum skin reactivity in platinum refinery workers. *Chest* 1990; 97: 1401-1407.
  172. McConnell LH, Fink JN, Schlueter DP, Schmidt MG. Asthma caused by nickel sensitivity. *Ann Intern Med* 1973; 78: 888-890.
  173. Block GT, Yeung M. Asthma induced by nickel. *J Am Med Assoc* 1982; 247: 1600-1602.
  174. Malo JL, Cartier A, Doepner M, Nieboer E, Evans S, Dolovich J. Occupational asthma caused by nickel sulfate. *J Allergy Clin Immunol* 1982; 69: 55-59.
  175. Hartmann AL, Walter H, Wuthrich B. Allergisches berufasthma auf pektinase, ein pektolytisches enzym. *Schweiz Med Wschr* 1983; 113: 265-267.
  176. Gheysens B, Auxwerx J, Van Den Eeckhout A, Demedts M. Cobalt-induced bronchial asthma in diamond polishers. *Chest* 1985; 88: 740-744.
  177. Malo JL, Cartier A. Occupational asthma due to fumes of galvanized metal. *Chest* 1987; 92: 375-377.
  178. Vogelmeier C, König G, Bencze K, Fruhmann G. Pulmonary involvement in zinc fume fever. *Chest* 1987; 92: 946-949.
  179. Bruckner HC. Extrinsic asthma in a tungsten carbide worker. *J Occup Med* 1967; 9: 518-519.
  180. Smith AR. Chrome poisoning with manifestations of sensitization. *J Am Med Assoc* 1931; 94: 95-98.
  181. Joules H. Asthma from sensitization to chromium. *Lancet* 1932; ii: 182-183.
  182. Keskinen G, Kalliomaki PL, Alanko K. Occupational asthma due to stainless steel welding fumes. *Clin Allergy* 1980; 10: 151-159.
  183. Novey HS, Habib M, Wells ID. Asthma and IgE antibodies induced by chromium and nickel salts. *J Allergy Clin Immunol* 1983; 72: 407-412.
  184. Shirakawa T, Kusaka Y, Fujimura N, Kato M, Heki S, Morimoto K. Hard metal asthma: cross immunological and respiratory reactivity between cobalt and nickel. *Thorax* 1990; 45: 267-271.
  185. Davies RJ, Hendrick DJ, Pepys J. Asthma due to inhaled chemical agents: ampicillin, benzyl penicillin, 6-amino penicillanic acid and related substances. *Clin Allergy* 1974; 4: 227-247.
  186. Lagier F, Cartier A, Dolovich J, Malo J-L. Occupational asthma in a pharmaceutical worker exposed to penicillamine. *Thorax* 1989; 44: 157-158.
  187. Coutts II, Dally MB, Newman-Taylor AJ, Pickering CAC, Horsfield N. Asthma in workers manufacturing cephalosporins. *Br Med J* 1981; 283: 950.
  188. Briatico-Vangosa G, Beretta F, Bianchi S, et al. Bronchial asthma due to 7-aminocephalosporanic acid (7-ACA) in workers employed in cephalosporin production. *Med Lav* 1981; 72: 488-493.
  189. Kammermeyer JK, Mathews KP. Hypersensitivity to phenylglycine acid chloride. *J Allergy Clin Immunol* 1973; 52: 73-84.
  190. Busse WW, Schoenwetter WF. Asthma from psyllium in laxative manufacture. *Ann Intern Med* 1975; 83: 361-362.
  191. Bardy JD, Malo JL, Séguin P, et al. Occupational asthma and IgE sensitization in a pharmaceutical company processing psyllium. *Am Rev Respir Dis* 1987; 135: 1033-1038.
  192. Cartier A, Malo J-L, Dolovich J. Occupational asthma in nurses handling psyllium. *Clin Allergy* 1987; 17: 1-6.
  193. Malo JL, Cartier A, L'Archevêque J, et al. Prevalence of occupational asthma and immunologic sensitization to psyllium among health personnel in chronic care hospitals. *Am Rev Respir Dis* 1990; 142: 1359-1366.
  194. Harries MG, Newman-Taylor AJ, Wooden J, MacAuslan A. Bronchial asthma due to alpha-methyl dopa. *Br Med J* 1979; 1 (6176): 1461.
  195. Davies RJ, Pepys J. Asthma due to inhaled chemical agents: the macrolide antibiotic spiramycin. *Clin Allergy* 1975; 1: 99-107.
  196. Malo JL, Cartier A. Occupational asthma in workers of a pharmaceutical company processing spiramycin. *Thorax* 1988; 43: 371-377.
  197. Moscato G, Naldi L, Candura F. Bronchial asthma due to spiramycin and adipic acid. *Clin Allergy* 1984; 14: 355-361.
  198. Fawcett IW, Pepys J, Erooga MA. Asthma due to "glycyl compound" powder: an intermediate in production of salbutamol. *Clin Allergy* 1976; 6: 405-409.
  199. Greene SA, Freedman S. Asthma due to inhaled chemical agents: amprolium hydrochloride. *Clin Allergy* 1976; 6: 105-108.
  200. Fawcett IW, Pepys J. Allergy to a tetracycline preparation. *Clin Allergy* 1976; 6: 301-303.
  201. Menon MPS, Das AK. Tetracycline asthma: a case report. *Clin Allergy* 1977; 7: 285-290.
  202. Asai S, Shimoda T, Hara K, Fujiwara K. Occupational



- asthma caused by isonicotinic acid hydrazide (INH) inhalation. *J Allergy Clin Immunol* 1987; 80: 578–582.
203. Perrin B, Malo JL, Cartier A, Evans S, Dolovich J. Occupational asthma in a pharmaceutical worker exposed to hydralazine. *Thorax* 1990; 45: 980–981.
  204. Lee HS, Wang YT, Yeo CT, Tan KT, Ratnam KV. Occupational asthma due to tylosin tartrate. *Br J Ind Med* 1989; 46: 498–499.
  205. Luczynska CM, Marshall PE, Scarisbrick DA, Topping MD. Occupational allergy due to inhalation of ipecacuanha dust. *Clin Allergy* 1984; 14: 169–175.
  206. Coutts II, Lozewicz S, Dally MB, *et al.* Respiratory symptoms related to work in a factory manufacturing cimetidine tablets. *Br Med J* 1984; 288: 14–18.
  207. Biagini RE, Bernstein DM, Klinecicz SL, Mittman R, Bernstein IL, Henningsen GM. Evaluation of cutaneous responses and lung function from exposure to opiate compounds among ethical narcotics-manufacturing workers. *J Allergy Clin Immunol* 1992; 89: 108–117.
  208. Feinberg SM, Watrous RM. Atopy to simple chemical compounds: sulfonechloramides. *J Allergy* 1945; 16: 209–220.
  209. Bourne MS, Flindt MLH, Walker JM. Asthma due to industrial use of chloramine. *Br Med J* 1979; 2: 10–12.
  210. Dijkman JG, Vooren PH, Kramps JA. Occupational asthma due to inhalation of chloramine-T. 1. Clinical observations and inhalation-provocation studies. *Int Arch Allergy Appl Immunol* 1981; 64: 422–427.
  211. Andrasch RH, Bardana EJ, Koster F, Pirofsky B. Clinical and bronchial provocation studies in patients with meat-wrapper's asthma. *J Allergy Clin Immunol* 1976; 58: 291–298.
  212. Sokol WN, Aelony Y, Beall GN. Meat-wrapper's asthma. A new syndrome? *J Am Med Assoc* 1973; 226: 639–641.
  213. Lee HS, Yap J, Wang YT, Lee CS, Tan KT, Poh SC. Occupational asthma due to unheated polyvinylchloride resin dust. *Br J Ind Med* 1989; 46: 820–822.
  214. Kopp SK, McKay RT, Moller DR, Cassidy K, Brooks SM. Asthma and rhinitis due to ethylcyanoacrylate instant glue. *Ann Intern Med* 1985; 102: 613–615.
  215. Weiner A. Bronchial asthma due to the organic phosphate insecticides. *Ann Allergy* 1961; 19: 397–401.
  216. Alanko K, Keskinen H, Byorksten F, Ojanen S. Immediate-type hypersensitivity to reactive dyes. *Clin Allergy* 1978; 8: 25–31.
  217. Park HS, Lee MK, Kim BO, *et al.* Clinical and immunologic evaluations of reactive dye-exposed workers. *J Allergy Clin Immunol* 1991; 87: 639–649.
  218. Romano C, Sulotto F, Pavan I, Chiesa A, Scansetti G. A new case of occupational asthma from reactive dyes with severe anaphylactic response to the specific challenge. *Am J Ind Med* 1992; 21: 209–216.
  219. Pepys J, Hutchcroft BJ, Breslin ABX. Asthma due to inhaled chemical agents-persulphate salts and henna in hairdressers. *Clin Allergy* 1976; 6: 399–404.
  220. Baur X, Fruhmann G, Liebe VV. Occupational asthma and dermatitis after exposure to dusts of persulfate salts in two industrial workers. *Respiration* 1979; 38: 144–150.
  221. Blainey AD, Ollier S, Cundell D, Smith RE, Davies RJ. Occupational asthma in a hairdressing salon. *Thorax* 1986; 41: 42–50.
  222. Pankow W, Hein H, Bittner K, v Wichert P. Asthma in hairdressers induced by persulphate. *Pneumologie* 1989; 43: 173–175.
  223. Gamboa PM, de la Cuesta CG, Garcia BE, Castillo JG, Oehling A. Late asthmatic reaction in a hairdresser, due to the inhalation of ammonium persulphate salts. *Allergol Immunopathol* 1989; 17: 109–111.
  224. Slovak AJM. Occupational asthma caused by a plastics blowing agent, azodicarbonamide. *Thorax* 1981; 36: 906–909.
  225. Malo JL, Pineau L, Cartier A. Occupational asthma due to azobisformamide. *Clin Allergy* 1985; 15: 261–264.
  226. Normand J-C, Grange F, Hernandez C, *et al.* Occupational asthma after exposure to azodicarbonamide: report of four cases. *Br J Ind Med* 1989; 46: 60–62.
  227. Graham V, Coe MJS, Davies RJ. Occupational asthma after exposure to a diazonium salt. *Thorax* 1981; 36: 950–951.
  228. Luczynska CM, Hutchcroft BJ, Harrison MA, Dornan JD, Topping MD. Occupational asthma and specific IgE to diazonium salt intermediate used in the polymer industry. *J Allergy Clin Immunol* 1990; 85: 1076–1082.
  229. Nagy L, Orosz M. Occupational asthma due to hexachlorophene. *Thorax* 1984; 39: 630–631.
  230. Hendrick DJ, Lane DJ. Formalin asthma in hospital staff. *Br Med J* 1975; 1: 607–608.
  231. Burge PS, Harries MG, Lam WK, O'Brien IM, Patchett PA. Occupational asthma due to formaldehyde. *Thorax* 1985; 40: 255–260.
  232. Nordman H, Keskinen H, Tuppurainen M. Formaldehyde asthma: rare or overlooked? *J Allergy Clin Immunol* 1985; 75: 91–99.
  233. Cockcroft DW, Hoepfner VH, Dolovich J. Occupational asthma caused by cedar urea formaldehyde particle board. *Chest* 1982; 82: 49–53.
  234. Frigas E, Filley WV, Reed CE. Asthma induced by dust from urea-formaldehyde foam insulating material. *Chest* 1981; 79: 706–707.
  235. Malo JL, Gagnon G, Cartier A. Occupational asthma due to heated freon. *Thorax* 1984; 39: 628–629.
  236. Cockcroft DW, Cartier A, Jones G, Tarlo SM, Dolovich J, Hargreave FE. Asthma caused by occupational exposure to a furan-based binder system. *J Allergy Clin Immunol* 1980; 66: 458–463.
  237. Topping MD, Forster HW, Ide CW, Kennedy FM, Leach AM, Sorkin S. Respiratory allergy and specific immunoglobulin E and immunoglobulin G antibodies to reactive dyes used in the wool industry. *J Occup Med* 1989; 31: 857–862.
  238. Moscato G, Biscaldi G, Cottica D, Pugliese F, Candura S, Candura F. Occupational asthma due to styrene: two case reports. *J Occup Med* 1987; 29: 957–960.
  239. Jachuck SJ, Bound CL, Steel J, Blain PG. Occupational hazard in hospital staff exposed to 2 percent glutaraldehyde in an endoscopy unit. *J Soc Occup Med* 1989; 39: 69–71.
  240. Lozewicz S, Davison AG, Hopkirk A, *et al.* Occupational asthma due to methyl methacrylate and cyanoacrylates. *Thorax* 1985; 40: 836–839.
  241. Pickering CAC, Bainbridge D, Birtwistle IH, Griffiths DL. Occupational asthma due to methyl methacrylate in an orthopaedic theatre sister. *Br Med J* 1986; 292: 1362–1363.
  242. Hendrick DJ, Connolly MJ, Stenton SC, Bird AG, Winterton IS, Walters EH. Occupational asthma due to sodium isononanoyl oxybenzene sulphonate, a newly developed detergent ingredient. *Thorax* 1988; 43: 501–502.
  243. Waclawski ER, McAlpine LG, Thomson NC. Occupational asthma in nurses caused by Chlorhexidine and alcohol aerosols. *Br Med J* 1989; 298: 929–930.
  244. Burge PS. Occupational asthma, rhinitis and alveolitis due to colophony. *Clin Immunol Allergy* 1984; 4: 55–82.

245. Dugue P, Faraut C, Figueredo M, Bettendorf A, Salvadori JM. Asthme professionnel à l'oxyde d'éthylène chez une infirmière. *Presse Méd* 1991; 20: 1455.
246. Gannon PFG, Sherwood-Burge P, Benfield CFA. Occupational asthma due to polyethylene shrink wrapping (paper wrappers asthma). *Thorax* 1992; 47: 759.
247. Honda I, Kohrogi H, Ando M, *et al.* Occupational asthma induced by the fungicide tetrachloroisophthalonitrile. *Thorax* 1992; 47: 760-761.
248. Shelton D, Urch B, Tarlo SM. Occupational asthma induced by a carpet fungicide, tributyl tin oxide. *J Allergy Clin Immunol* 1992; 90: 274-275.
249. Tarlo SM. Occupational asthma induced by tall oil in the rubber tyre industry. *Clin Exp Allergy* 1991; 22: 99-102.
250. Kennes B, Garcia-Herreros P, Sierckx P. Asthma from plexiglas powders. *Clin Allergy* 1981; 11: 49-54.
251. Tarlo SM, Wong L, Roos J, Booth N. Occupational asthma caused by latex in a surgical glove manufacturing plant. *J Allergy Clin Immunol* 1990; 85: 626-631.
252. Kern DG, Frumkin H. Asthma in respiratory therapists. *Ann Intern Med* 1989; 110: 767-773.
253. Musk AW, Peach S, Ryan G. Occupational asthma in a mineral analysis laboratory. *Br J Ind Med* 1988; 45: 381-386.
254. Hendy MS, Beattie BE, Burge PS. Occupational asthma due to an emulsified oil mist. *Br J Ind Med* 1985; 42: 51-54.
255. Midttun O. Bronchial asthma in the aluminium industry. *Acta Allergol* 1960; 15: 208-221.
256. Saric M, Godnic-Cvar J, Gonzi M, Stilinovic L. The role of atopy in potroom workers' asthma. *Am J Ind Med* 1986; 9: 239-242.
257. Wergeland E, Lund E, Waage JE. Respiratory dysfunction after potroom asthma. *Am J Ind Med* 1987; 11: 627-636.
258. O'Donnell TV, Welford B, Coleman ED. Potroom asthma: New Zealand experience and follow-up. *Am J Ind Med* 1989; 14: 43-49.