

Supplementary information

Title: Smoking exposure is associated with airway eosinophilic activation and autoimmunity in severe asthma

Supplementary methods

Inclusion criteria in SATS and SEPIA:

SATS:

Inclusion criteria: A physician's diagnosis of asthma and high-dose inhaled corticosteroids (ICS) treatment (≥ 1600 μg budesonide or equivalent) with a second controller (long acting beta-agonist (LABA), theophylline or leukotriene-antagonist) for the previous year or OCS for ≥ 50 % of the previous year.

Exclusion criteria: age below 18 years, pregnancy or judged by the investigator to be unable to comply with the study protocol.

SEPIA:

Inclusion criteria: Informed consent, a valid asthma diagnosis either according to patient medical record (positive bronchial provocation test, positive reversibility test or PF-variability) or by positive asthma test at screening (mannitol or reversibility test). Eosinophilic phenotype defined by at least 3% sputum eosinophils, severe asthma according to ERS/ATS guidelines; i.e. treatment with high-dose inhaled corticosteroid (ICS) (≥ 1600 μg budesonide or equivalent) plus a second controller, either long-acting beta-2-agonist (LABA), leukotriene antagonist (LTRA), theophylline, or long-acting muscarinergic antagonist (LAMA).

Exclusion criteria: Current treatment with oral corticosteroids (any dose), current pregnancy, age under 18 years, co-morbidities such as malignant lung disease, severe bronchiectasis, severe emphysema, lung

fibrosis, significant cardiac disease, treatment with anti-asthma biologicals (anti-IgE, anti-IL5, anti-IL5-receptor, anti-IL4/13-receptor) or other immunosuppressants within the last 16 weeks, lower airway infection requiring antibiotics within the last 6 weeks, and change in maintenance ICS dose within the preceding 4 weeks.

Clinical assessments

Dynamic and static lung function measurements (spirometry and body plethysmography) as well as gas diffusion testing was performed on a *MasterScreen Pneumo* spirometer and a *MasterScreen BodyBox* (Jaeger, Würzburg, Germany). Spirometry was conducted in accordance with standard ERS protocol[1] and predicted values were based on NHANES reference data[2]. Fractionated exhaled nitrogen oxide (FeNO) was measured on the *Ecomedics CLD88sp* (Ecomedics AG, Duernten, Switzerland) in the SATS study, while the SIGNATURE study used the *Niox Vero* device (Circassia Limited, Uppsala, Sweden)[3].

Atopy was assessed by IgE to a standard panel of ten aeroallergens: pollen from birch (*betula verrucosa*), grass (*phleum pratense*), or mugwort (*artemisia vulgaris*); dander from horse (*equus caballus*), cat (*felius domesticus*) or dog (*canis familiaris*); house dust mites (*dermatophagoides pteronyssinus* or *dermatophagoides farinae*); or mold (*alternaria alternatia* and *cladosporium herbarum*)[4].

Sputum induction

In the SIGNATURE study, patients with an FEV₁ over 70% were induced with mannitol; if this proved ineffective, incremental doses of saline was used instead. Patients with an FEV₁ under 70% were induced with isotonic saline (0.9%) following a reversibility test. In the SATS study, patients with an FEV₁ over 70% were induced with incrementally increasing doses of hypertonic saline,

while patients with an FEV₁ under 70% were induced with isotonic saline (0.9%)[5, 6]. In both studies, sputum was assessed within an hour of collection, using the “plug selection method”[7].

Supplementary tables

Table S1: Baseline characteristics, patients with severe eosinophilic asthma

Baseline characteristics	< 10 pack years	≥10 pack years	Healthy controls	P-value
Total No., n	31	27	22	-
Sex, females n (%)	14 (45.2%)	6 (22.2%)	13(59%)	0.029
BMI (kg/m ²)	27.1 ± 4	27.4 ± 5	24.4 ± 4	0.47
Mean age (yr), range	50(20-71)	58(40-80)	32(20-63)	<0.001
Adult onset asthma, n (%)	15(48.3)	17(63.0)	NA	0.30
Pack years, total	0(0-1)	17(10-69)	0(0-0)	<0.001
Daily ICS dose, µg	1600(1600-6400)	1600(1600-3200)	NA	0.09
Daily OCS dose, mg	0(0-5)	0(0-12.5)	NA	0.01
ACQ-5 score	1.8 ± 1.0	2.2 ± 0.9	NA	0.08
Exacerbation rate	1.29 ± 1.3	2.63 ± 2.5	NA	0.02
FEV ₁ (%predicted)	72.1 ± 21.6	70.9 ± 19.7	106.7 ± 12.9	<0.001
FVC (%predicted)	93.6 ± 21.4	86.4 ± 17.1	111.7 ± 15.5	<0.01
FEV ₁ /FVC-ratio	0.63 ± 0.12	0.64 ± 0.11	0.82 ± 0.06	<0.001
Atopy ^ε , no. (%)	21(67.7)	14(51.9)	0(0)	<0.001
Total serum IgE (kU/L)	228 (4-13500)	196 (12-2480)	22 (3-137)	<0.001
Total leukocyte count	7.4 ± 2.4	7.8 ± 2.4	6.5 ± 2.3	0.19
Blood eosinophils	0.42 ± 0.43	0.54 ± 0.	0.12 ± 0.07	<0.001
Blood neutrophils	4.4 ± 1.9	4.4 ± 2.0	3.3 ± 1.4	0.18
Blood lymphocytes	1.8 ± 0.7	1.5 ± 0.6	2.3 ± 0.9	0.10
FeNO, ppb	29(9-84)	28(7-219)	12(9.5-16.5)	<0.01
% Sputum eosinophils (range)	7.3(3.0-76.0)	12.5(3.3-92.3)	0 (0-2.8)	<0.001
% Sputum neutrophils (range)	53.3(7.0-88.3)	44.8(2.3-93.3)	38.3(9.0-88.0)	0.45
% Sputum macrophages (range)	20.3(0.5-81.5)	18.3(0.8-65.3)	61.1(8.8-89.0)	<0.01
% Sputum lymphocytes (range)	0(0-4)	0(0-8)	0 (0-2.5)	0.90

Table S1: Baseline characteristics of the study population. Data are shown as mean±SD, median (IQR), numbers (n) or %. ICS: Inhaled corticosteroids; OCS: oral corticosteroids, prednisolone. ^εAtopy defined as a positive skin prick test or positive specific IgE.

Table S2: Baselines characteristics, SEPIA sub-study (2-weeks OCS treatment)

Baseline characteristics	Statistic	≥10 pack years (N=12)	< 10 pack years (N=11)	p-value
Sex (n females, %)	n / N (%)	1 / 11 (9.1%)	6 / 12 (50%)	0.069
Age at inclusion (yr)	Median (Range)	58 (41-68)	51 (28-71)	0.52
BMI (kg/m ²)	Mean, SD	26.7±4.1	25.7±4.8	0.70
Exacerbation rate (events/yr)	Mean, SD	2.36±1.03	1.42±0.67	0.024
Smoking history (pack years)	Median (Range)	15 (12-25)	0 (0-1)	<0.001
Smoking cessation, duration (yr)	Median (Range)	7.0 (2.0-20.0)	NA	-
ACQ5 score	Mean, SD	1.85±0.84	2.37±0.87	0.22
ICS dose (µg/day)	Median (Range)	1.600 (1.600- 3.200)	1.600 (1.600- 3.200)	0.31
Nasal polyposis (n, %)	n / N (%)	5 / 11 (45%)	5 / 12 (42%)	>0.9
Atopy ^ε (n positive, %)	n / N (%)	9 / 11 (82%)	5 / 12 (42%)	0.089
FeNO(ppb)	Median (Range)	39.0 (12.0-67.0)	26.0 (9.0-59.0)	0.21
Total IgE	Median (Range)	160 (4-2480)	289 (50-1260)	0.33
FEV1 (%pred.)	Mean, SD	78.6±22	80.8±21	>0.9
FVC (%pred)	Mean, SD	98.5±19	96.0±25	0.62
FEV1/FVC ratio	Mean, SD	65.0±7.0	66.0±11.0	0.43
Sputum eosinophils (%)	Median (Range)	20 (5-86)	10 (4-90)	0.42
Sputum neutrophils (%)	Median (Range)	37 (4-79)	54 (4-91)	0.11
Sputum macrophages (%)	Median (Range)	42 (4-65)	19 (1-52)	0.052
Sputum lymphocytes (%)	Median (Range)	0 (0-10)	0(0-1.50)	0.20
Blood total leukocyte (x10 ⁹ cells/L)	Mean, SD	6.62±1.70	7.35±2.34	0.40
Blood neutrophils (x10 ⁹ cells/L)	Mean, SD	3.65±0.97	4.04±1.18	0.42
Blood eosinophil (x10 ⁹ cells/L)	Mean, SD	0.38±0.16	0.27±0.64	0.24
Blood lymphocyte (x10 ⁹ cells/L)	Mean, SD	1.85±0.41	1.64±0.91	0.83
CRP (mg/L)	Mean, SD	5.5±6.3	5.2±9.7	0.41
Anti-EPX status (n positive, %)	n / N (%)	4 / 11 (36%)	1 / 12 (8.3%)	0.20
FEG score	Median (Range)	1.00 (1.00-3.00)	0.00 (0.00-3.00)	0.028

Table S2: Baseline characteristics of the study population in the Sepia study. Data are shown as mean±SD, median (IQR), numbers (n) or %. ICS: Inhaled corticosteroids. ^εAtopy defined as a positive skin prick test or positive specific IgE.

1. Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Crapo R, Enright P, van der Grinten CPM, Gustafsson P, Jensen R, Johnson DC, MacIntyre N, McKay R, Navajas D, Pedersen OF, Pellegrino R, Viegi G, Wanger J. Standardisation of spirometry. *Eur. Respir. J.* England; 2005; 26: 319–338.
2. Hankinson JL, Odenkrantz JR, Fedan KB. Spirometric reference values from a sample of the general U.S. Population. *Am. J. Respir. Crit. Care Med.* 1999; 159: 179–187.
3. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide, 2005. *Am. J. Respir. Crit. Care Med.* United States; 2005; 171: 912–930.
4. Bousquet J, Khaltaev N, Cruz AA, Denburg J, Fokkens WJ, Togias A, Zuberbier T, Baena-Cagnani CE, Canonica GW, Van Weel C, Agache I, Ait-Khaled N, Bachert C, Blaiss MS, Bonini S, Boulet L-P, Bousquet P-J, Camargos P, Carlsen K-H, Chen Y, Custovic A, Dahl R, Demoly P, Douagui H, Durham SR, Van Wijk RG, Kalayci O, Kaliner MA, Kim Y-Y, Kowalski ML, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) 2008*. *Allergy* Blackwell Publishing Ltd; 2008; 63: 8–160.
5. Pavord ID, Pizzichini MM, Pizzichini E, Hargreave FE. The use of induced sputum to investigate airway inflammation. *Thorax* [Internet] England; 1997; 52: 498–501 Available from: <http://thorax.bmj.com/cgi/doi/10.1136/thx.52.6.498>.
6. Hamid Q, Kelly MMM, Linden M, Louis R, Pizzichini MMMMM, Pizzichini E, ## CR, Van F, }} O, Djukanovic´z R, Djukanovic´z D, Ronchi C, Van Overveld F, Djukanovic R. Methods of sputum processing for cell counts, immunocytochemistry and in situ hybridisation. *Eur. Respir. J.* 2002; 20: 19S-23s.
7. Pizzichini E, Pizzichini MM, Efthimiadis A, Hargreave FE, Dolovich J. Measurement of inflammatory indices in induced sputum: effects of selection of sputum to minimize salivary contamination. *Eur. Respir. J.* 1996; 9: 1174–1180.