Cystic fibrosis transmembrane conductance regulator protein: What is its role in cystic fibrosis?

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Cystic fibrosis (CF) is a disease involving many organs including sweat glands, pancreas, gastrointestinal tract and airways. Disordered airway function is the main cause of illness and death in the increasing numbers of CF sufferers who survive infancy. At one time it was thought that a circulating CF factor or factors might cause the airway malfunction, but studies of normal lungs transplanted into CF patients have called this into question. Airways in normal lungs transplanted into CF patients have a normal epithelial potential difference and show only the same rate of infection with Pseudomonas aeruginosa and Staphylococcus aureus as do lungs transplanted into non-CF recipients [1]. Thus, the airway lesions in CF seem to depend in large part on the local expression of a disordered gene, probably within the mucosa.

It is now just over three years since the gene locus responsible for CF was found and its gene cloned and sequenced [2, 3, 4]. The gene product is a membranespanning protein, cystic fibrosis transmembrane conductance regulator (CFTR), which acts as a chloride channel regulated via the cyclic AMP (cAMP) pathway [5]. In cells from CF patients, stimulation of cAMP release fails to open the channel. However, introduction of DNA coding for the correct protein into these cells restores their ability to conduct chloride via this channel [6]. This and other observations led to the hypothesis that lack of functional CFTR protein in the apical membrane of the epithelium leads to the abnormalities seen in CF lungs [7]. Surface epithelium reabsorbs sodium ions from the liquid in the airway lumen and can secrete chloride into it. In man Na+ reabsorption is the chief ion movement. Water moves along the osmotic gradient created by ion movements. In patients with CF, chloride movement into the lumen cannot be stimulated by cAMP and sodium reabsorption is greater than normal, though the reason for this is unknown. It has been suggested that these abnormalities would dehydrate mucus, giving it abnormal physical properties, and reduce the layer of periciliary fluid, both of which would slow mucociliary clearance. This in turn might explain why CF airways are liable to infection. This hypothesis is seductive but cannot explain the entire pathophysiology of the CF airway. For example, patients with 'immotile cilia syndrome'

have a more profound slowing of airway mucus transport than that seen in CF but, typically, they suffer only mild airway infections [8]. Also, the hypothesis predicts that preventing sodium reabsorption from the airway with the drug amiloride should prevent the withdrawal of water from the airway lumen and so mitigate the disease. In a trial of amiloride on CF patients with previously stable lung disease, Knowles et al. [9] first treated airway infections vigorously with antibiotics and then withdrew conventional treatments. The patients were split into two groups, one treated with amiloride aerosol and the other receiving placebo. Over the next 6 months, FVC declined in both groups but more slowly in the amiloride-treated group. However, a subsequent 6 month cross-over trial of amiloride aerosol versus placebo given in addition to conventional therapy showed no benefit [10]. Thus, partial correction of the abnormal ion movement seems, at the moment, only to provide a marginal ben-

Could absence of functional CFTR protein at other sites in the airway also be important in the genesis of the CF syndrome? In this issue of the journal, Jacquot et al. publish their finding that airway submucosal gland cells express CFTR protein, some of it in the basal and apical membranes of the secretory cells, some in the membranes of the secretory granules and some in the cell cytoplasm [11]. This independently confirms the recent report of Engelhardt et al. who describe submucosal gland as the main site of CFTR in the airway [12]. Thus, CF patients lack functional CFTR in submucosal gland cells, and this finding suggests alternative explanations for the lung manifestations of the disease, some of which are reviewed below:

1) The role of CFTR as a chloride channel in providing the water for submucosal gland secretions. In airways with submucosal glands, the main bulk of secretion appears to come from these glands. Could chloride secretion via the CFTR protein account for the production of water by the submucosal glands? Under basal conditions human submucosal glands in culture do not secrete Cl but activation of either the Ca⁺⁺ or cAMP second messenger pathway stimulates Cl secretion in non-CF glands [13]. In CF glands, however, stimulation of either pathway fails to produce Cl secretion [14]. Agents acting through these

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second messenger pathways may also have weaker effects on mucin secretion in CF than in normal bronchi [15]. In contrast, Corrales et al. [16] measured the rate at which they could collect fluid from ducts of individual submucosal glands in feline trachea in vitro and tested how this responded to stimulation with phenylephrine. They found that blocking chloride secretion did not alter the flow of mucus in the ducts in response to phenylephrine. This appears to contradict the idea that the CFTR protein is important in providing Cl⁻ and water for submucosal gland secretion. They proposed instead that the secretory granules contain osmotically active particles which, on their release, take up water and cause the granules to swell, an idea supported by Verdugo [17].

A recent study has shown that the CFTR protein introduced into Xenopus oocytes transports water as well as anions in response to cAMP. It is not yet certain that it plays the same role in submucosal glands in the human airway. Thus CFTR protein in human submucosal gland may play a part in conducting Cland water into mucus and failure of this mechanism may be important in CF.

- 2) Plasma membrane recycling. Recently it has been shown that CFTR protein is important in the recycling of plasma membranes [18]. Proteins such as horseradish peroxidase (HRP) can be used as markers to follow the process of endocytosis. In normal cells, raised levels of cAMP within the cytoplasm inhibited endocytosis, but in CF cells raised cAMP failed to influence the process. Transfection of CF cells with the gene encoding for normal CFTR protein restored the ability of cAMP to inhibit endocytosis. This study also showed that CFTR protein regulates exocytosis of macromolecules. In normal cells cAMP stimulates this process, but not in CF cells. Again, transfection of CF cells with the normal CFTR gene produced exocytosis in response to cAMP. We do not know what part CFTR protein has in the normal regulation of endo- and exocytosis, but there is evidence that Cltransport is important in the latter process. Nor is it vet apparent how disordered control of endo- and exocytosis could lead to the airway lesions in CF.
- 3) Acidity of the Golgi apparatus. Some intracellular organelles, such as the Golgi apparatus, are normally more acid than the cell cytoplasm. The process of acidification involves the transfer of a proton (H+) which is accompanied by a Cl- ion, perhaps transported by the CFTR protein. BARASCH et al. [19] have reported that, in nasal polyp tissue, the trans-Golgi and pre-lysosomes from CF tissues are less acid than the equivalent organelles from normals by 0.2-0.3 pH units. They attribute this to the failure of faulty CFTR protein to conduct sufficient Cl to accompany protons in CF. They argue that the relatively alkaline interior of the Golgi apparatus in CF cells may explain the observed lack of sialic acid in the oligosaccharides of CF mucins because sialyl transferase enzymes work most efficiently at an acid pH. This could be

functionally important because sialic acid is one of the terminating groups of the oligosaccharide sidechains of mucins; glycosylating enzymes continue to add further sugars to a sidechain until an end group such as sialic acid is in place. This suggests a possible way in which absence of the CFTR protein could lead to over-glycosylation of mucins in CF and so be responsible for abnormal mucus. Could the modest difference in pH between normal and CF cells really make an important difference? Furthermore, two recent studies now question the role of CFTR in acidifying intracellular organelles [20, 21]. Showing that transfection of CF cells with the CFTR gene increases mucin sialylation would make this hypothesis more persuasive.

- 4) Oversulphation of mucins. CF mucus from both sputum and cell cultures contains an abnormally large amount of sulphate ion in the mucin macromolecules [22]. The authors proposed that this results from an abnormal biochemical process intimately connected with the CF lesion and suggested that the altered mucins resulting from this might explain the susceptibility of CF airways to colonisation by Pseudomonas aeruginosa and Staphylococcus aureus. Increased sulphation in CF might arise from a failure of acidification of the Golgi apparatus (see 3 above) inhibiting sialylation of the oligosaccharide sidechains, thus allowing them to grow abnormally long and providing correspondingly more positions for sulphation. Another hypothesis to explain increased sulphation is that the abnormal anion channel found in CF membranes might cause large quantities of SO₄ to reach sulphation sites in the Golgi apparatus. On the other hand, altered glycosylation and sulphation of mucins with an increase in mucin acidity is a feature of prolonged airway inflammation and so might not be specific to CF [23]. Again, it would be interesting to see whether mucin sulphation returned to normal in cultures of CF airway cells after transfection with the gene for normal CFTR.
- 5) Transport of macromolecules. Finally, it is important to consider the possibility that CFTR protein may act as an active transporter of macromolecules, not just as a Cl⁻ channel. This idea arose because the predicted structure of CFTR protein resembles that of a family of proteins which act as energy requiring transporters of macromolecules. This firmly includes P-glycoprotein, responsible for the drug-resistance in many solid tumours by active cellular extrusion of chemotherapeutic agents. Recently it has become clear that P-glycoprotein can also act as a chloride channel [24]. By analogy CFTR protein might also have a dual role as macromolecular transporter and ion channel.

The idea that the submucosal glands are the site of an important lesion in CF has implications for therapy. The attractions of a topical route for drugs aimed at correcting the consequences of the CF defect, or

for genes to correct the defect itself, are obvious. If the site of the lesion is the submucosal gland, instead of (or as well as) the surface epithelium, directing the therapy to its target will be more difficult. This may delay effective therapy, but should not put

it beyond the scope of human ingenuity.

The way that our knowledge of CF has advanced is instructive. Defective chloride transport was initially predicted by electrophysiologists working on CF tissues. Then molecular biologists identified the faulty gene and, together with electrophysiologists, confirmed that the gene product is a chloride channel. This has not led to an immediate understanding of the disease, but has provided techniques and ideas which are spurring cell biologists, biochemists and physiologists in their attempts to explain CF. We are now approaching the time when physicians will be able to draw together insights provided by the various basic sciences in order to treat CF effectively.

References

- Smyth R, Higenbottam T, Scott J, Wallwork J. The current state of lung transplantation for cystic fibrosis. Thorax 1991; 46: 213-216.
- Rommens J, Ianuzzi M, Kerem B-S, et al. Identification of the cystic fibrosis gene: walking and jumping. Science 1989; 245: 1059-1065.
- Riordan J, Rommens J, Karem B-S, et al. Identification of the cystic fibrosis gene: cloning and characterisation of the complementary DNA. Science 1989; 1066-1073.
- Kerem B-S, Rommens J, Buchanan J, et al. Identification of the cystic fibrosis gene: genetic analysis. Science 1989; 245: 1073-1080.
- 5. Bear C, Li C, Kartner N, Bridges R, Jensen T, Ramjeesingh M, Riordan J. - Purification and functional reconstitution of the cystic fibrosis transmembrane conductance regulator (CFTR). Cell 1992; 68: 809-818.
- Rich D, Anderson M, Gregory R, et al. Expression of cystic fibrosis transmembrane conductance regulator corrects defective chloride channel regulation in cystic fibrosis airway epithelial cells. Nature 1990; 347: 358-363. Quinton P. - Cystic fibrosis: a disease in electrolyte

transport. FASEB J 1990; 4: 2709-2717

Eliason R, Mossberg B, Camner P, Afzelius B. Immotile cilia, chronic airway infections and male sterility. New Engl J Med 1977; 297: 1-6.

Knowles M, Church N, Waltner W, et al. - A pilot

- study of aerosolised amiloride for the treatment of lung disease in cystic fibrosis. New Engl J Med 1990; 322: 1189-
- 10. Graham A, Hasani A, Alton E, Hodson M, Geddes D. Trial of nebulised amiloride in patients with cystic fibrosis. Thorax 1992; 47: 867. (abstract)
- 11. Jacquot J, Puchelle E, Hinnrasky J, et al. Localization of the CFTR in airway secretory glands. Eur Respir J 1993; 6: 169-176.
- 12. Engelhardt J, Yankaskas J, Ernst S, Yang Y, Marino C, Boucher R, Cohn J, Wilson J. - Submucosal glands are the predominant site of CFTR expression in the human bronchus. Nature Genetics 1992; 2: 240-248.
- 13. Yamaya M, Finkbeiner W, Widdicombe J. Ion transport by cultures of human tracheobronchial submucosal glands. Am J Physiol 1991; 261: L485-L490.
- 14. Yamaya M, Finkbeiner W, Widdicombe J. Altered ion transport by tracheal submucosal glands in cystic fibrosis. Am J Physiol 1991; 261: L491-L494.
- 15. Rogers D, Alton E, Dewer A, Lethem M, Barnes P. Impaired stimulus-evoked mucus secretion in cystic fibrosis bronchi. Exp Lung Res 1992; 19: 37-53.
- 16. Corrales R, Nadel J, Widdicombe J. Source of the fluid component from tracheal submucosal glands in cats. J Appl Physiol 1984; 56: 1076-1082.
- 17. Verdugo P. Goblet cell secretion and mucogenesis. Ann Rev Physiol 1990; 52: 157-176.
- 18. Bradbury N, Jilling T, Berta G, Sorscher E, Bridges R, Kirk K. - Regulation of plasma membrane recycling by CFTR. Science 1992; 256: 530-532.
- 19. Barasch J, Kiss B, Prince A, Saiman L, Gruenert D, Al-Aqwati Q. - Defective acidification of intracellular organelles in cystic fibrosis. Nature 1991; 352: 70-73.
- 20. Van Dyke R, Root K, Schrieber J, Wilson J. Role of CFTR in lysosome acidification. Biochem Biophys Res Comm 1992; 184: 300-305.
- 21. Lukacs G, Chang X, Kartner N, Rotstein O, Riordan J, Grinstein S. - The CFTR is present and functional in endosomes. Role as a determinant of endosomal pH. J Biol Chem 1992; 267: 14568-14572.
- 22. Cheng P-W, Boat T, Granfill K, Yankaskas J, Boucher R. - Increased sulfation of glycoconjugates by cultured nasal epithelial cells from patients with cystic fibrosis. J Clin Invest 1989; 84: 68-72.
- 23. Reid L. Animal models in clinical disease. Ciba Found Symp 1978; 54: 297-307.
- 24. Valverde M, Diaz M, Sepulveda F, Gill D, Hyde S, Higgins C. - Volume regulated chloride channels associated with the human multidrug-resistance P-glycoprotein. Nature 1992; 355: 830-833.