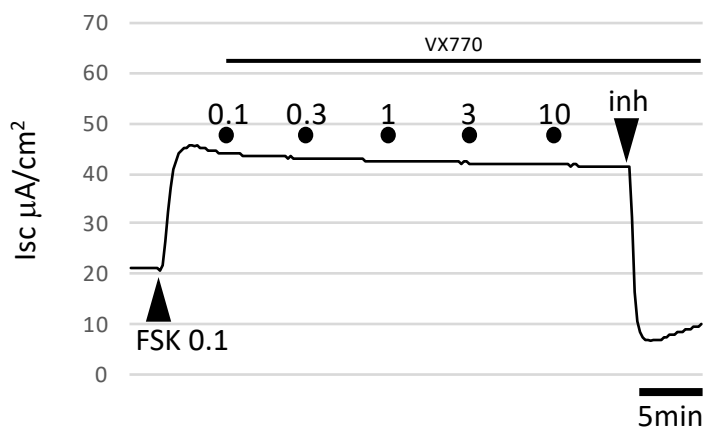


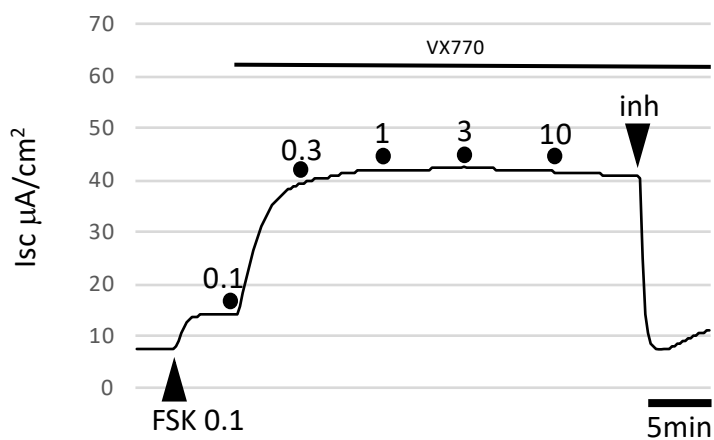
Supplemental Figure S1. Chemicals and protocols A. chemical structures of the therapeutic correctors lumacaftor, tezacaftor and elexacaftor and of the potentiator ivacaftor. B. scheme illustrating the protocols used to record  $I_{sc}$  in airway epithelial cells. POT: genistein or Cact-A1. C. scheme showing the analysis of  $I_{sc}$  to determine basal  $I_{sc}$  (a), forskolin (b) and potentiator (c)-dependent  $\Delta I_{sc}$ , activatable (d) and inhibitable (e) CFTR  $\Delta I_{sc}$ .

Figure S2

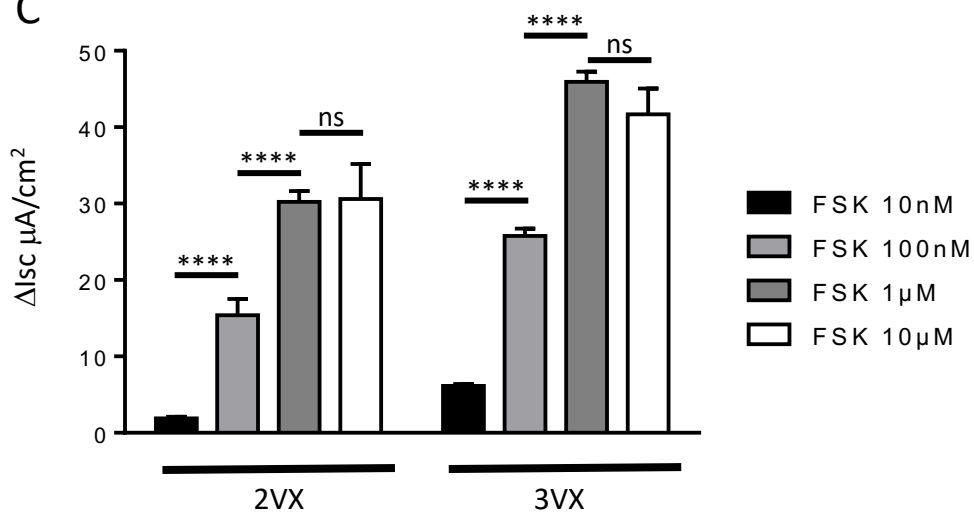
A. 3VX



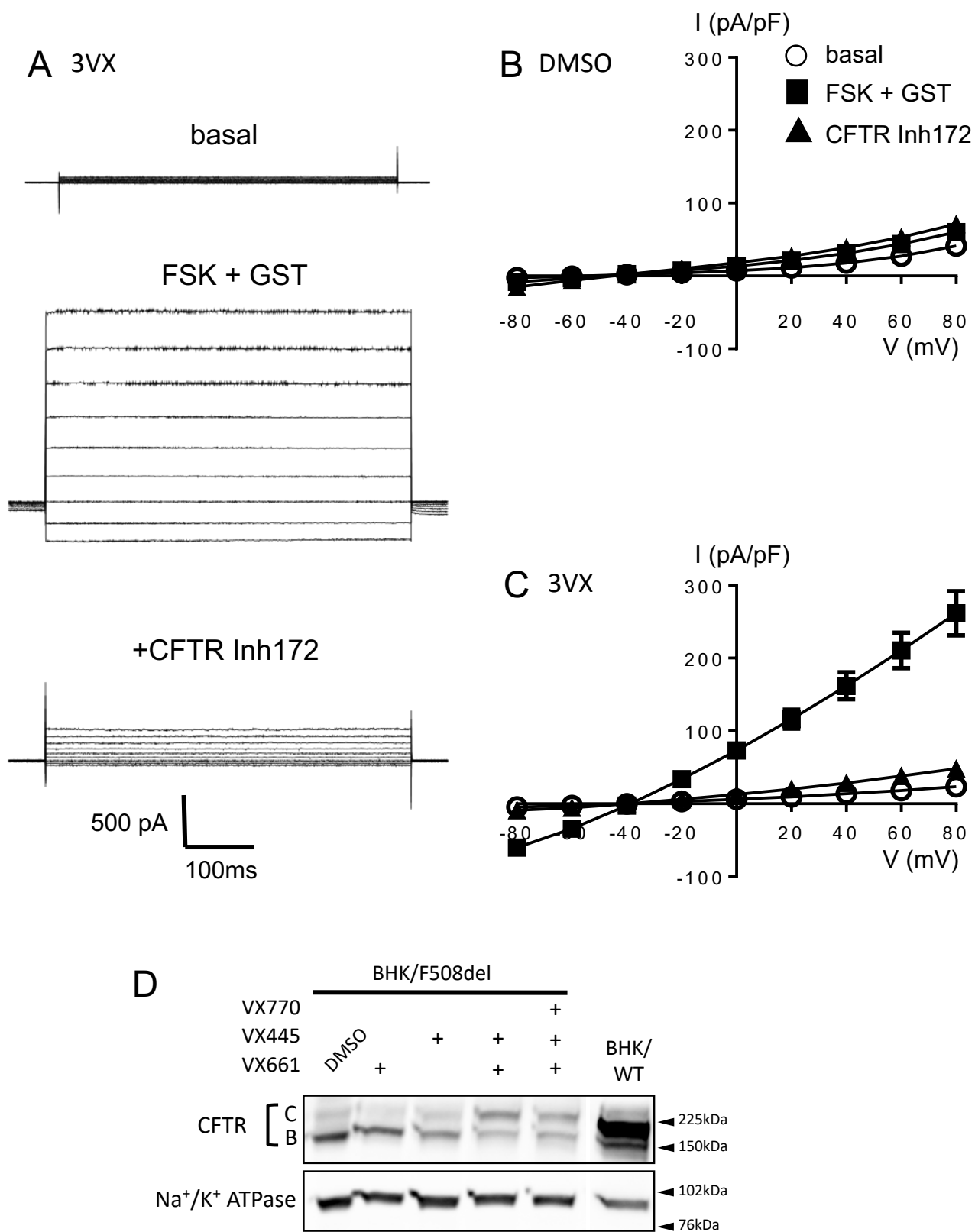
B. 2VX



C



Supplemental Figure S2. Stimulation of F508del-Isc by VX770 and forskolin. a, b) Original tracings showing Isc with CFBE F508del cells incubated by elexacaftor/tezacaftor/ivacaftor (noted 3VX in a) or elexacaftor/tezacaftor (noted 2VX in b). The Isc CFTR current was stimulated by FSK (0.1  $\mu$ M, arrow) then VX770 (increasing concentrations as indicated by the dot on the traces) then inhibited by 10  $\mu$ M CFTRinh172 (inh, second arrow). c) Mean  $\pm$  SEM of  $\Delta$ Isc in response to increasing concentrations of FSK (n=4-8 for each concentration). ns, no significant difference and \*\*\*\* $P < 0.0001$ .



Supplemental Figure S3. Whole-cell patch clamp recordings of F508del chloride currents in BHK cells. a) Original tracings of whole-cell F508del-CFTR chloride currents in basal (upper traces), after adding FSK (10  $\mu$ M) + GST (30  $\mu$ M) (middle traces) and after adding CFTRinh172 (10  $\mu$ M) (bottom traces). b, c) Current density/V relationships for non-treated BHK cells (n=14, b) or BHK cells treated by elexacaftor/tezacaftor/ivacaftor (n=15, c). Basal is indicated by circles, FSK+GST by squares and CFTRinh172 by triangles. d) CFTR expression of the total protein fraction from BHK F508del cells after 24h-incubation with VX661 (18  $\mu$ M), VX445 (3  $\mu$ M) and VX770 (1  $\mu$ M) alone or in combination or DMSO control as indicated above blot and of BHK WT cells grown on dish (representative blot of 2 independent experiments). Equal protein loading was controlled via Na<sup>+</sup>/K<sup>+</sup> ATPase detection.