



Impact and associations of eosinophilic inflammation in COPD: analysis of the AERIS cohort

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Blood eosinophil levels in COPD predict the nature of inflammation at future exacerbations and may guide therapy <http://ow.ly/W10o30dNQiq>

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ABSTRACT Eosinophilic inflammation in chronic obstructive pulmonary disease (COPD) predicts response to treatment, especially corticosteroids. We studied the nature of eosinophilic inflammation in COPD prospectively to examine the stability of this phenotype and its dynamics across exacerbations, and its associations with clinical phenotype, exacerbations and infection.

127 patients aged 40–85 years with moderate to very severe COPD underwent repeated blood and sputum sampling at stable visits and within 72 h of exacerbation for 1 year.

Blood eosinophils $\geq 2\%$ was prevalent at baseline, and predicted both predominantly raised stable-state eosinophils across the year (area under the curve 0.841, 95% CI 0.755–0.928) and increased risk of eosinophilic inflammation at exacerbation (OR 9.16; $p < 0.001$). Eosinophils $\geq 2\%$ at exacerbation and eosinophil predominance at stable visits were associated with a lower risk of bacterial presence at exacerbation (OR 0.49; $p = 0.049$ and OR 0.25; $p = 0.065$, respectively). Bacterial infection at exacerbation was highly seasonal (winter *versus* summer OR 4.74; $p = 0.011$) in predominantly eosinophilic patients.

Eosinophilic inflammation is a common and stable phenotype in COPD. Blood eosinophil counts in the stable state can predict the nature of inflammation at future exacerbations, which when combined with an understanding of seasonal variation provides the basis for the development of new treatment paradigms for this important condition.

Introduction

Chronic obstructive pulmonary disease (COPD) is an established cause of global mortality and morbidity, predicted to be the third leading cause of death by 2030 [1]. Current guidelines for COPD management are based on airway obstruction, symptoms and exacerbation frequency [2]. The guidelines advise on stepwise management, but do not fully account for the biological heterogeneity of this debilitating condition.

Eosinophilic inflammation was historically thought to be a feature of asthma, with neutrophilic inflammation being a classical hallmark of COPD. However, recent studies have demonstrated that eosinophilic inflammation is present in a subset of COPD patients both at exacerbations and during clinical stability [3, 4]. A sputum eosinophil differential count of >3% is an accepted marker of airway eosinophilic inflammation and is derived from the reported enhanced response of this group of patients to corticosteroids [5, 6]. Moreover, a good relationship has been demonstrated between airway and systemic eosinophil counts [3, 4, 7]. It was previously reported that a >2% blood eosinophil cut-off had a high sensitivity in identifying >3% airway eosinophils during acute exacerbations of COPD (AECOPD) [3], suggesting that peripheral eosinophils are a clinically accessible marker to predict inflammatory phenotype.

Using 2% blood eosinophils as a cut-off in the longitudinal ECLIPSE cohort, persistent elevation of blood eosinophils was reported as a common finding [4], especially in milder disease, but exacerbations were not sampled. Reports of relationships between eosinophils and forced expiratory volume in 1 s (FEV₁) and exacerbation frequency vary [4, 8–12], but emerging evidence suggests measurement of eosinophils has clinical relevance. For example, severe AECOPD with eosinophilic inflammation are associated with a shorter length of hospital stay [13]. Furthermore, there is an association between eosinophilic inflammation with a greater response to steroid therapy in both stable COPD and during exacerbations [5, 6, 14–16].

Management of asthma stratified by airway eosinophilic inflammation has led to a reduced rate of exacerbations [17]. SIVA *et al.* [14] demonstrated that COPD treatments targeting airway eosinophils were associated with a significant reduction of severe exacerbations.

Hence, eosinophilic inflammation is an important COPD endotype, but little is known about its stability over time or its relationships to the inflammatory nature and aetiology of exacerbations. To improve our understanding, we investigated these factors in a secondary analysis of the AERIS (Acute Exacerbation and Respiratory InfectionS in COPD) cohort, a prospective, longitudinal study of patients with COPD [18].

Methods

Study design and study population

The AERIS study is a prospective, observational cohort study based at University Hospital Southampton (Southampton, UK) and registered with ClinicalTrials.gov (identifier NCT01360398). The study protocol and full inclusion/exclusion criteria have been previously published [18]. The protocol summary is available at www.gsk-clinicalstudyregister.com (identifier 114378). AERIS was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice, and was approved by the Southampton and South West Hampshire Research Ethics Committee. All participants provided written informed consent. Only patients with a confirmed diagnosis of COPD based on post-bronchodilator spirometry with FEV₁ ≤80% of predicted and FEV₁/forced vital capacity (FVC) <0.7, and ex-smokers or current smokers with significant smoking history (≥10 pack-year history) were included in the cohort, with no patients having diagnosis of asthma at the time of recruitment [18].

Patients were followed monthly in the stable state and reviewed within 72 h of onset of AECOPD symptoms. Exacerbations were detected using daily electronic diary cards. Definitions of AECOPD and severity were as previously described [18, 19].

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Procedures

Blood and sputum analyses were performed as previously described [18, 19]. Further methodological details are provided in the supplementary material.

Criteria for eosinophilic groups and seasonality

Eosinophilic inflammation was defined as sputum eosinophils >3% and blood eosinophils \geq 2%, in line with previous studies [3, 5, 6, 14, 20]. To investigate the stability of blood eosinophilic inflammation over time we divided subjects into three groups: predominantly (PE), intermittent (IE) and rarely (RE) eosinophilic groups. Only those subjects who had at least three (out of five potential) stable visits with valid blood results over 12 months were included in the group analyses (n=99). The PE group was defined as blood eosinophils \geq 2% at either all visits or all but one visit where blood eosinophils were <2%; the RE group was defined as blood eosinophils <2% at all visits or all but one visit where blood eosinophils were \geq 2%; the IE group was defined when none of the above-mentioned criteria were met.

To investigate an impact of seasonality on exacerbations we divided the year into two seasons: winter (October–March) and summer (April–September).

Statistical analysis

Bivariate analyses testing for differences between eosinophilic groups were conducted using two-tailed, Kruskal–Wallis, ANOVA, Chi-squared or Fisher's exact tests, as appropriate. Receiver operator curves were used to assess the predictive ability of different cut-offs to correctly identify presence of sputum eosinophilic inflammation. Intraclass correlation coefficients (ICCs) were used to assess the reliability of measures within individuals over time. As some subjects were represented multiple times in exacerbation analysis, descriptive analyses were conducted only for the first exacerbation occurring for each subject and multivariate analyses with binary outcomes (presence/absence of different conditions at exacerbation) were conducted using conditional logistic regression, including the subject number as a random effect. SPSS version 22 (IBM, Armonk, NY, USA) was used for all analyses with the exception of ICCs and conditional logistic regression, which were analysed using Stata version 14 (StataCorp, College Station, TX, USA). All of these analyses should be considered *post hoc* as they were not pre-specified in the AERIS statistical analysis plan.

Results

General characteristics of the cohort

152 patients were screened, but only 127 patients were included in the AERIS cohort (figure 1). The cohort consisted of patients with moderate to very severe airway obstruction; the majority (87.3%) were receiving inhaled corticosteroids and had a substantial smoking history (mean \pm SD pack-years 50.3 \pm 28.2) (table 1). Only patients who experienced at least one moderate to severe exacerbation in the last 12 months were included in the cohort (median (interquartile range (IQR)) rate 2.00 (2.00)).

Clinical and inflammatory characterisation at enrolment

The prevalence of eosinophilic inflammation \geq 3% in sputum in our cohort at enrolment was 33% (n=27) and in blood \geq 2% was 68% (n=86). Blood eosinophils (percentage and count) displayed moderately strong positive correlations with sputum eosinophil percentage (ρ =0.463 and 0.581, respectively, at enrolment; p <0.001) and were predictive of sputum eosinophils >3% (area under the curve (AUC) 0.851, 95% CI 0.750–0.951 and AUC 0.768, 95% CI 0.651–0.884, respectively) (figure 2). This \geq 2% blood eosinophil cut-point was 95.8% sensitive and 31.8% specific. Using absolute cell numbers (\geq 200 cells- μ L⁻¹) rather than percentages gave similar results (table 1).

Stability of eosinophilic phenotype over time

We categorised our patients into three longitudinal eosinophilic inflammation phenotypic groups based on blood eosinophils \geq 2% and found that, out of the 99 patients with sufficient data, 57 (58%) were in the PE group, 16 (16%) were in the IE group and 26 (26%) were in the RE group over the 12 months examined (table 2). Between the three longitudinal groups, the only significant differences observed at enrolment were age, blood neutrophils and presence of sputum eosinophils >3% (table 2). There were also significant differences in the PE group exacerbation rate over the first year. Significant differences between patients included and excluded from this analysis were seen for C-reactive protein (CRP), 6-min walk test (6MWT) and length of follow-up (supplementary table E2). In order to assess the longitudinal stability of blood eosinophilic inflammation, we analysed the reliability of the marker in defining a longitudinal phenotype and found blood eosinophils to be relatively stable within individuals across the 12 months examined, including enrolment (ICC 0.66, 95% CI 0.58–0.74).

We next examined whether blood eosinophils at a single time-point (enrolment) were a useful predictor of being in the PE group over time (categorisation for the eosinophilic inflammation groups excluding

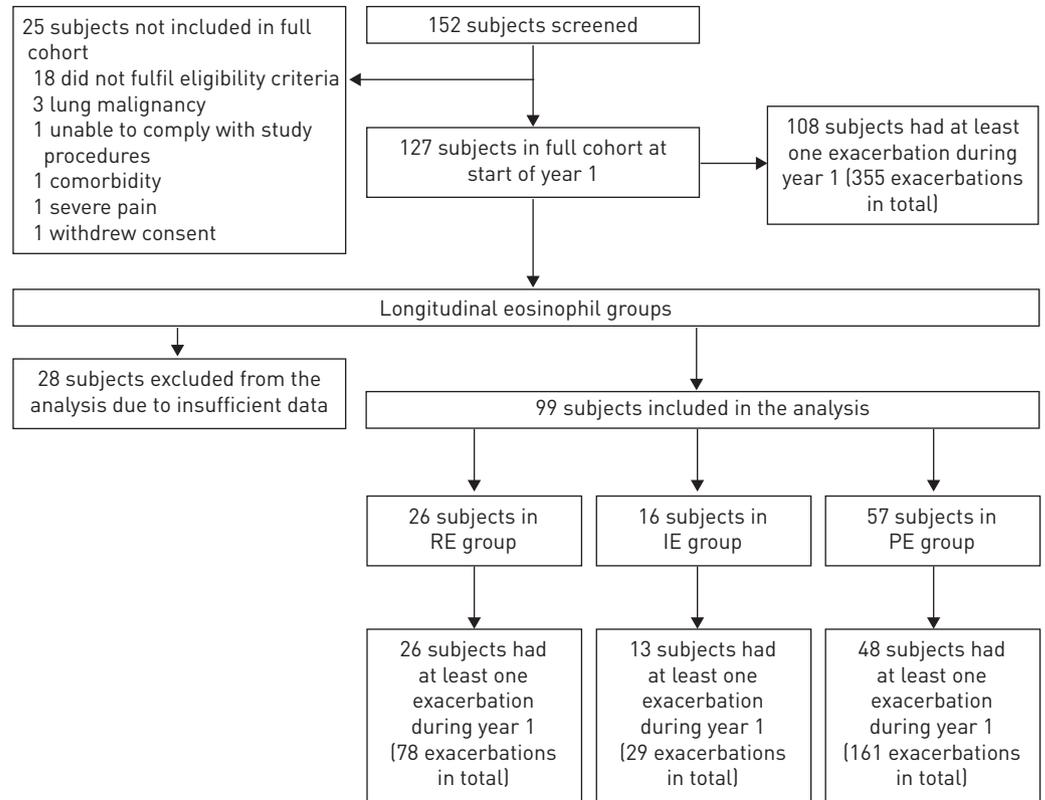


FIGURE 1 Flow diagram of the subjects screened and included in the full cohort at the start of year 1 and the number of exacerbations in year 1. RE: rarely eosinophilic; IE: intermittently eosinophilic; PE: predominantly eosinophilic.

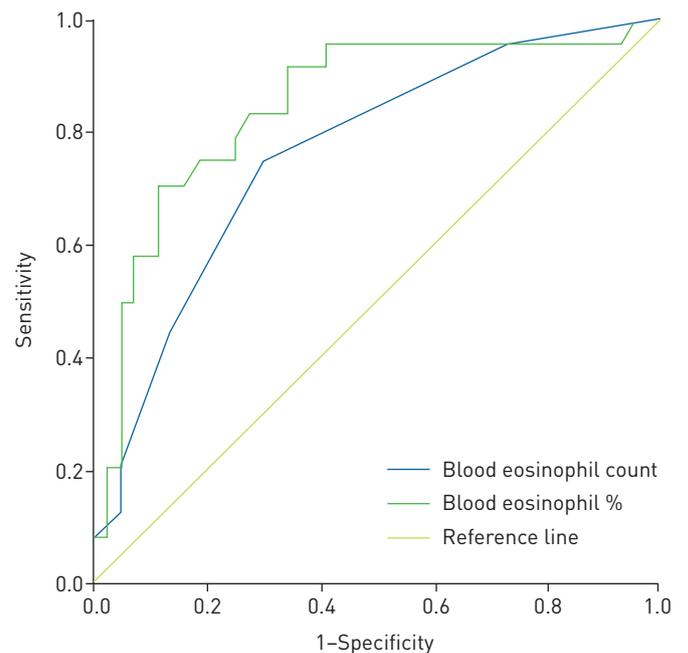


FIGURE 2 Receiver operating characteristic curve with area under the curve (AUC) for blood eosinophils (count and percentage) at enrolment predicting sputum eosinophilia >3% at enrolment (n=68). Blood eosinophil count: AUC 0.768, 95% CI 0.651–0.884; blood eosinophil percentage: AUC 0.851, 95% CI 0.750–0.951.

TABLE 1 Baseline characteristics of the total chronic obstructive pulmonary disease (COPD) cohort

Total patients	127
Continuous variables	
Age years	66.8±8.61
Smoking history pack-years	50.3±28.2
Body mass index kg·m ⁻²	27.0 (6.69)
Fat-free body mass %	48.9 (21.5) (n=125)
White blood cells count ×10 ⁹ L ⁻¹	7.60 (2.20) (n=126)
Eosinophils count ×10 ⁹ L ⁻¹	0.20 (0.20) (n=126)
Eosinophils %	2.94 (3.08) (n=126)
Neutrophils count ×10 ⁹ L ⁻¹	4.80 (1.70) (n=126)
Fibrinogen g·L ⁻¹	4.80 (1.02) (n=114)
C-reactive protein mg·L ⁻¹	5.00 (8.00)
Eosinophils % [#]	1.93 (5.05) (n=83)
Neutrophils % [#]	47.0 (70.4) (n=83)
FEV ₁ % pred	46.7 (25.3) (n=126)
ΔFEV ₁ % of baseline [¶]	4.93 (21.1) (n=85)
FEV ₁ reversibility % of pre-bronchodilator FEV ₁ ⁺	11.2 (17.0) (n=105)
K _{co} %	69.2 (30.7) (n=122)
T _{LCO} %	57.9 (29.5) (n=122)
CAT score	16.0 (10.0) (n=126)
6MWT distance m	300 (170) (n=125)
EXACT-PRO score	37.0 (12.0) (n=101)
Exacerbation rate in year before study	2.00 (2.00)
Exacerbation rate in first year of study	2.94 (3.91)
Eosinophilic exacerbation rate in first year of study	0.99 (2.02)
Follow-up in first year of study years	1.00 (0.02)
Categorical variables	
Sex	
Male	68 (54)
Female	59 (46)
Current smoker [§]	
Yes	54 (43)
No	73 (57)
Frequent exacerbators in year 0	
Yes	99 (78)
No	28 (22)
Use of ICS at enrolment ^f	
Yes	113 (89)
No	14 (11)
Sputum eosinophils >3% [#]	
Yes	27 (33)
No	56 (67)
Blood eosinophils ≥2%	
Yes	86 (68)
No	40 (32)
Bacteria present ^{##}	
Yes	57 (52)
No	53 (48)
Virus present ^{¶¶}	
Yes	18 (17)
No	85 (83)
Blood eosinophils ≥200 cells·μL ⁻¹	
Yes	90 (71.4)
No	36 (28.6)

Data are presented as n for patient numbers, mean±SD or median (interquartile range) for continuous variables, or n (%) for categorical variables. FEV₁: forced expiratory volume in 1 s; K_{co}: transfer coefficient of the lung for carbon monoxide; T_{LCO}: transfer factor of the lung for carbon monoxide; CAT: COPD Assessment Test; 6MWT: 6-min walk test; ICS: inhaled corticosteroid. [#]: sputum eosinophil and neutrophil percentage at baseline is reported ("baseline" is equal to enrolment if good quality data [squamous cell contamination <30%] is present at enrolment or the next (pre-exacerbation) stable visit with quality data within 4 months of enrolment); [¶]: calculated as FEV₁ at month 12×100/FEV₁ at enrolment; ⁺: calculated as (post-bronchodilator FEV₁-pre-bronchodilator FEV₁)/pre-bronchodilator FEV₁×100; [§]: smoking status report derived from American Thoracic Society Division of Lung Disease questionnaire ATS-DLD-78A; ^f: ICS use was coded as "Yes" if one of the following medications/inhalers was on the list: Symbicort, Seretide, QVAR, Fostair, beclomethasone, beclomethasone/formoterol, beclomethasone dipropionate, Clenil, fluticasone/salmeterol or budesonide/formoterol; ^{##}: sputum sampling, measured by culture (includes *Haemophilus influenzae*, *Moraxella catarrhalis*, *Streptococcus pneumoniae*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*); ^{¶¶}: sputum sampling, measured by PCR (includes adenovirus, enterovirus, influenza, coronavirus, metapneumovirus, bocavirus, parainfluenza, respiratory syncytial virus and rhinovirus).

TABLE 2 Baseline characteristics by longitudinal blood eosinophil group over 12 months for the 99 patients with sufficient data

	Rarely eosinophilic	Intermittently eosinophilic	Predominantly eosinophilic	p-value**
Patients	26	19	59	
Continuous variables				
Age	62.7±8.63	67.9±6.87 (n=16)	68.0±9.05 (n=57)	0.034
Smoking history pack-years	46.3±22.8	64.9±43.2 (n=16)	51.0±28.6 (n=57)	0.098
Body mass index kg·m ⁻²	28.2 (8.15)	27.9 (5.69) (n=16)	25.9 (4.73) (n=57)	0.280
Fat-free body mass %	53.5 (21.5) (n=25)	49.3 (18.3) (n=16)	48.8 (22.4) (n=56)	0.886
White blood cells count ×10 ⁹ L ⁻¹	8.30 (3.30) (n=25)	7.65 (1.65) (n=16)	7.30 (2.00) (n=57)	0.060
Blood eosinophils count ×10 ⁹ L ⁻¹	0.10 (0.10) (n=25)	0.20 (0.10) (n=16)	0.30 (0.20) (n=57)	NA
Blood eosinophils %	1.35 (1.45) (n=25)	2.12 (1.09) (n=16)	4.08 (3.05) (n=57)	NA
Blood neutrophils count ×10 ⁹ L ⁻¹	5.20 (2.55) (n=25)	4.80 (0.50) (n=16)	4.30 (1.75) (n=57)	0.030
Fibrinogen g·L ⁻¹	4.70 (1.40) (n=23)	4.75 (0.97) (n=14)	4.80 (0.90) (n=51)	0.905
C-reactive protein mg·L ⁻¹	4.00 (7.50)	5.00 (5.75) (n=16)	5.00 (7.00) (n=57)	0.966
Sputum eosinophils % [#]	0.43 (2.89) (n=18)	0.82 (1.78) (n=7)	2.81 (7.40) (n=40)	0.070
Sputum neutrophils % [#]	9.77 (64.0) (n=18)	62.3 (63.4) (n=7)	38.0 (71.6) (n=40)	0.223
FEV ₁ %	47.7 (25.6)	43.9 (22.4) (n=16)	47.5 (26.0) (n=57)	0.661
ΔFEV ₁ % of baseline [¶]	3.79 (27.3) (n=19)	10.8 (23.2) (n=15)	5.03 (21.4) (n=45)	0.375
FEV ₁ reversibility % of pre-bronchodilator FEV ₁ [*]	7.69 (15.0) (n=24)	17.9 (16.6) (n=13)	12.6 (12.5) (n=46)	0.355
Kco %	74.2 (31.0) (n=25)	68.3 (26.7) (n=16)	69.3 (30.3) (n=53)	0.526
Tlco %	62.0 (41.1) (n=25)	57.8 (28.8) (n=16)	56.3 (27.7) (n=53)	0.407
CAT score	15.5 (10.0)	20.5 (13.0) (n=16)	16.0 (9.00) (n=57)	0.101
6MWT distance m	300 (199) (n=25)	323 (172) (n=16)	327 (164) (n=56)	0.914
EXACT-PRO score	36.0 (14.0) (n=23)	36.5 (7.00) (n=14)	36.0 (15.0) (n=46)	0.607
Exacerbation rate in year before study	2.00 (3.00)	2.00 (3.00) (n=16)	2.00 (2.00) (n=57)	0.941
Exacerbation rate in first year of study	2.47 (3.22)	1.02 (2.00) (n=16)	2.04 (3.95) (n=57)	0.197
Eosinophilic exacerbation rate in first year of study	0.00 (0.99)	0.00 (0.99) (n=16)	1.38 (2.99) (n=57)	<0.001
Follow-up in first year of study years	1.01 (0.02)	1.01 (0.01) (n=16)	1.01 (0.02) (n=57)	0.448
Categorical variables				
Sex				
Male	12 (46)	9 (56)	34 (60)	0.547
Female	14 (54)	7 (44)	23 (40)	
Current smoker [§]				
Yes	15 (58)	4 (25)	24 (42)	0.125
No	11 (42)	12 (75)	33 (58)	
Use of ICS at enrolment ^f				
Yes	21 (81)	15 (94)	51 (89)	0.468
No	5 (19)	1 (6)	6 (11)	
Sputum eosinophils >3% [#]				
Yes	2 (11)	0 (0)	19 (48)	0.003
No	16 (89)	7 (100)	21 (53)	
Blood eosinophils ≥2%				
Yes	7 (28)	9 (56)	55 (96)	NA
No	18 (72)	7 (44)	2 (4)	
Bacteria present ^{###}				
Yes	12 (55)	6 (50)	26 (50)	0.952
No	10 (45)	6 (50)	26 (50)	
Virus present ^{¶¶}				
Yes	4 (19)	0 (0)	10 (20)	0.288
No	17 (81)	12 (100)	40 (80)	

Data are presented as n for patient numbers, mean±SD or median (interquartile range) for continuous variables, or n (%) for categorical variables, unless otherwise stated. FEV₁: forced expiratory volume in 1 s; Kco: transfer coefficient of the lung for carbon monoxide; Tlco: transfer factor of the lung for carbon monoxide; CAT: COPD Assessment Test; 6MWT: 6-min walk test; ICS: inhaled corticosteroid; NA: not appropriate. [#]: sputum eosinophil and neutrophil percentage at baseline is reported ("baseline" is equal to enrolment if good quality data [squamous cell contamination <30%] is present at enrolment or the next [pre-exacerbation] stable visit with quality data within 4 months of enrolment); [¶]: calculated as FEV₁ at month 12×100/FEV₁ at enrolment; ^{*}: calculated as (post-bronchodilator FEV₁-pre-bronchodilator FEV₁)/pre-bronchodilator FEV₁×100; [§]: smoking status report derived from American Thoracic Society Division of Lung Disease questionnaire ATS-DLD-78A; ^f: ICS use was coded as "Yes" if one of the following medications/inhalers was on the list: Symbicort, Seretide, QVAR, Fostair, beclomethasone, beclomethasone/formoterol, beclomethasone dipropionate, Clenil, fluticasone/salmeterol or budesonide/formoterol; ^{###}: sputum sampling, measured by culture (includes *Haemophilus influenzae*, *Moraxella catarrhalis*, *Streptococcus pneumoniae*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*); ^{¶¶}: sputum sampling, measured by PCR (includes adenovirus, enterovirus, influenza, coronavirus, metapneumovirus, bocavirus, parainfluenza, respiratory syncytial virus and rhinovirus); ^{**}: test for differences between the three longitudinal eosinophil groups [Kruskal-Wallis test for continuous variables; Chi-squared test for categorical variables].

enrolment, n=78). Blood eosinophils (percentage and count) at enrolment were predictive of being in the PE group over the next 12 months (AUC 0.841, 95% CI 0.755–0.928 and AUC 0.806, 95% CI 0.710–0.901, respectively) (figure 3). A blood eosinophil cut-off of $\geq 2\%$ was 84.3% sensitive and 48.1% specific in identifying those in the PE group over the next 12 months.

Blood eosinophilic inflammation groups at exacerbation and exacerbation rates

Across all exacerbations with valid data, the prevalence of eosinophilic inflammation in sputum and blood was 23.9% (52 out of 218) and 49.7% (168 out of 338), respectively. Blood eosinophils (percentage and count) were again predictive of sputum eosinophils $\geq 3\%$ at exacerbation (AUC 0.735, 95% CI 0.654–0.817 and AUC 0.729, 95% CI 0.650–0.809, respectively) (supplementary figure E1).

The median (IQR) total exacerbation rate in the first year of the study was similar in the RE and PE groups, but lower in the IE group (PE 2.04 (3.95), IE 1.02 (2.00) and RE 2.47 (3.22)). There was a significant difference in the rate of exacerbations with blood eosinophils $\geq 2\%$, with the PE group having a higher eosinophilic exacerbation rate (PE 1.38 (2.99), IE 0.00 (0.99) and RE 0.00 (0.99); $p < 0.001$).

Longitudinal eosinophilic groups did not differ symptomatically at first exacerbation (COPD Assessment Test (CAT) and EXACT-PRO (www.exactproinitiative.com)), but there was a significant difference in blood neutrophils at exacerbation between these groups ($p = 0.045$) and the PE group was less likely to have bacteria present ($p = 0.044$) (table 3). There was an association between longitudinal eosinophilic phenotype and eosinophils at exacerbation, with the odds of eosinophils $\geq 2\%$ at exacerbation much higher in the PE group compared with the RE group (OR 11.16, 95% CI 5.26–23.68; $p < 0.001$). Similarly, the odds of an exacerbation being eosinophilic were nine times higher in those who had blood eosinophils $\geq 2\%$ than in subjects with blood eosinophils $< 2\%$ at enrolment (OR 9.16, 95% CI 4.10–20.47; $p < 0.001$).

Seasonality of eosinophilic exacerbations

Exacerbations were more common in the winter season than in the summer season (217 and 138, respectively). The proportion of exacerbations with eosinophils $\geq 2\%$ was higher in the summer season than in the winter season; however, the number of eosinophilic exacerbations per season was similar (86 out of 217 in winter and 82 out of 138 in summer) (figure 4). The PE group had similar exacerbation rates in the summer and winter seasons, while the IE and RE groups had higher exacerbation rates in the winter season (supplementary table E3).

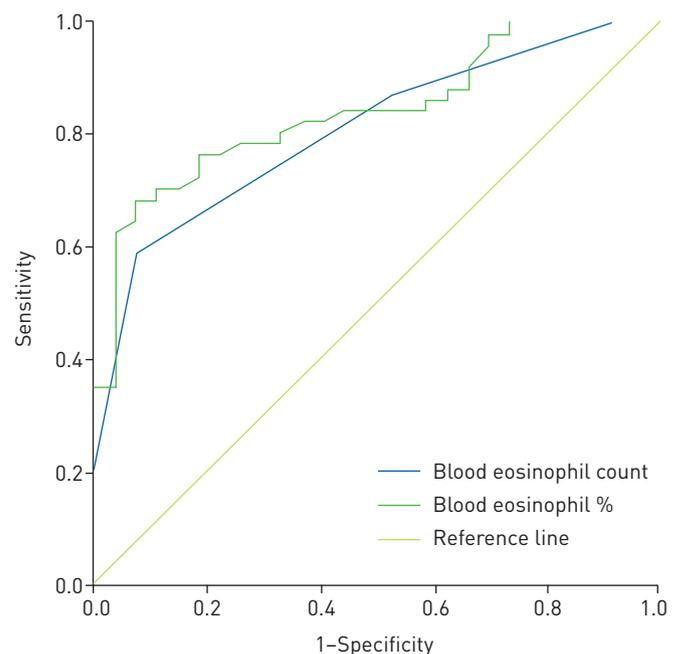


FIGURE 3 Receiver operating characteristic curve with area under the curve [AUC] for blood eosinophils (count and percentage) at enrolment predicting the predominantly eosinophilic group over 12 months following enrolment (n=78). Blood eosinophil count: AUC 0.806, 95% CI 0.710–0.901; blood eosinophil percentage: AUC 0.841, 95% CI 0.755–0.928.

TABLE 3 Characteristics at first exacerbation overall[#] and by longitudinal blood eosinophil groups over 12 months[¶]

	Overall cohort	Rarely eosinophilic	Intermittently eosinophilic	Predominantly eosinophilic	p-value ^f
Patients	108	26	13	48	
Continuous variables					
White blood cells count $\times 10^9 \text{ L}^{-1}$	8.00 [3.53] (n=102)	8.70 [4.25] (n=25)	8.95 [2.55] (n=12)	7.30 [3.23] (n=48)	0.349
Blood eosinophils count $\times 10^9 \text{ L}^{-1}$	0.20 [0.20] (n=102)	0.10 [0.15] (n=25)	0.15 [0.10] (n=12)	0.30 [0.27] (n=48)	<0.001
Blood eosinophils %	2.07 [2.78] (n=102)	1.19 [1.74] (n=25)	1.92 [0.88] (n=12)	3.39 [4.40] (n=48)	<0.001
Blood neutrophils count $\times 10^9 \text{ L}^{-1}$	5.30 [3.20] (n=102)	5.60 [4.05] (n=25)	5.55 [2.57] (n=12)	4.45 [2.98] (n=48)	0.045
Fibrinogen $\text{g}\cdot\text{L}^{-1}$	5.10 [1.40] (n=103)	4.80 [1.80] (n=25)	5.50 [2.25] (n=12)	5.10 [1.60] (n=46)	0.135
C-reactive protein $\text{mg}\cdot\text{L}^{-1}$	8.00 [14.0] (n=103)	7.00 [12.5] (n=25)	26.5 [49.8] (n=12)	7.00 [13.0] (n=47)	0.087
Sputum eosinophils %	1.20 [2.38] (n=61)	0.62 [2.29] (n=15)	1.40 [3.23] (n=9)	1.48 [3.56] (n=28)	0.273
Sputum neutrophils %	74.0 [47.1] (n=61)	76.8 [51.6] (n=15)	86.6 [40.3] (n=9)	74.1 [52.3] (n=28)	0.616
FEV ₁ % pred	42.7 [21.2] (n=97)	47.3 [30.0] (n=22)	41.5 [17.5] (n=12)	41.7 [20.0] (n=44)	0.395
CAT score	21.0 [11.0] (n=106)	22.0 [12.0] (n=26)	22.0 [17.0] (n=13)	19.0 [8.00] (n=47)	0.915
EXACT-PRO score	41.0 [11.0] (n=97)	43.0 [13.0] (n=23)	40.0 [10.0] (n=12)	39.5 [7.00] (n=44)	0.142
Categorical variables					
Bacteria present ⁺					
Yes	60 (61)	19 (86)	6 (55)	27 (57)	0.044
No	38 (39)	3 (14)	5 (45)	20 (43)	
Virus present [§]					
Yes	40 (43)	12 (55)	4 (40)	14 (33)	0.235
No	53 (57)	10 (45)	6 (60)	28 (67)	
Sputum eosinophils >3%					
Yes	23 (30)	2 (11)	0 (0)	17 (47)	0.004
No	54 (70)	16 (89)	7 (100)	19 (53)	
Blood eosinophils $\geq 2\%$					
Yes	73 (68)	7 (28)	7 (54)	47 (98)	<0.001
No	34 (32)	18 (72)	6 (46)	1 (2)	

Data are presented as n for patient numbers, median (interquartile range) for continuous variables or n (%) for categorical variables, unless otherwise stated. FEV₁: forced expiratory volume in 1 s; CAT: COPD Assessment Test; #: n=108; ¶: n=87; +: sputum sampling, measured by culture sputum sampling, measured by culture [includes *Haemophilus influenzae*, *Moraxella catarrhalis*, *Streptococcus pneumoniae*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*]; §: sputum sampling, measured by PCR [includes adenovirus, enterovirus, influenza, coronavirus, metapneumovirus, bocavirus, parainfluenza, respiratory syncytial virus and rhinovirus]; ^f: test for differences between the three longitudinal eosinophil groups (Kruskal-Wallis test for continuous variables; Chi-squared test for categorical variables).

After adjusting for the contribution of multiple exacerbations by some subjects, the odds of an exacerbation having blood eosinophils $\geq 2\%$ were 2.65 times higher in the summer season than in the winter season (supplementary table E4).

Both being in the PE compared with the RE group and summer season compared with winter season were independently associated with the odds of an exacerbation having blood eosinophils $\geq 2\%$ (OR 11.39, 95% CI 4.68–27.72; $p < 0.001$ and OR 2.57, 95% CI 1.37–4.85; $p = 0.003$, respectively).

We then repeated these analyses, stratifying by eosinophilic inflammation longitudinal phenotypes. The odds of an exacerbation having eosinophils $\geq 2\%$ in the summer season compared with the winter season tended to be stronger in the PE group (OR 3.73, 95% CI 1.56–8.91; $p = 0.003$) than in the IE and RE groups (OR 2.00, 95% CI 0.32–12.59; $p = 0.460$ and OR 1.36, 95% CI 0.42–4.44; $p = 0.606$, respectively).

Airway infection and eosinophilic inflammation

The increase in the number of noneosinophilic exacerbations during the winter season may indicate underlying changes in the lung microbiome as a result of the known seasonal effects of viral and bacterial infections [19]. We therefore examined whether the association between seasonality and prevalence of blood eosinophils $\geq 2\%$ at exacerbation persisted when accounting for the presence of potentially pathogenic microorganisms (PPMs) at exacerbation and found that this association did not diminish when accounting for PPMs (OR 2.39, 95% CI 1.29–4.41; $p = 0.005$).

PPMs were present in 59% of all exacerbations with valid data (n=320) and 61% of first exacerbations (n=98). We studied PPM prevalence at first exacerbation in the longitudinal eosinophilic groups and found a significant difference in PPM prevalence; PPMs were more prevalent in the RE group (86%) than in the other two groups (PE and IE: 57% and 55%, respectively; $p = 0.044$) (table 3). There was no such



FIGURE 4 Seasonal distribution of total and eosinophil-associated exacerbations. a) Number of total and exacerbations with blood eosinophils $\geq 2\%$. b) Proportion of exacerbations with blood eosinophils $\geq 2\%$ defined as exacerbations with blood eosinophils $\geq 2\%$ to total exacerbation rates in the predominantly eosinophilic, intermittent eosinophilic and rarely eosinophilic groups.

association in microbiology found between the three groups at enrolment ($n=99$; $p=0.952$) (table 2). Respiratory viruses were detected at 41% of all exacerbations with valid data ($n=305$) and 43.3% of first exacerbations ($n=90$) (table 3). We found no significant difference between the eosinophilic groups in the prevalence of the respiratory viruses either at enrolment or first exacerbation (tables 2 and 3).

The odds of having a PPM present at an exacerbation were 75% lower in the PE group compared with the RE group, but this association was of borderline significance (OR 0.25, 95% CI 0.06–1.09; $p=0.065$). The odds of having a PPM present were 55% lower in eosinophilic AECOPD compared with exacerbations with blood eosinophils $<2\%$, but this was also of borderline significance (OR 0.450, 95% CI 0.240–0.998; $p=0.049$).

The odds of having a PPM present at exacerbation were higher in the winter season than in the summer season (OR 2.51, 95% CI 1.27–4.96; $p=0.008$). This seasonal effect was apparent in the PE group (OR 4.74, 95% CI 1.43–15.71; $p=0.011$), but no statistically significant seasonal variations in PPMs at exacerbation were detected in the IE and RE groups (OR 4.42, 95% CI 0.01–3476.74; $p=0.662$ and OR 1.15, 95% CI 0.29–4.56; $p=0.838$).

Discussion

Eosinophilic inflammation is a stable longitudinal phenotype in a substantial proportion of COPD patients, which can be predicted over 12 months by an initial blood level measurement. We report for the first time that eosinophilic inflammation was more prevalent at exacerbation in those with predominantly raised eosinophils at stable state. In the summer season, a larger proportion of exacerbations were eosinophilic, although this was driven by fewer noneosinophilic exacerbations in the summer season compared with the winter season. We also report evidence that eosinophilic exacerbations are less frequently associated with airway bacterial infection, with the prevalence of airway bacterial infection at exacerbations greater among the group who rarely had raised eosinophils over time. These findings have a

potential implication for future therapeutic clinical trials and eosinophil targeted treatment with a view to stratifying patient care.

To the best of our knowledge, this is the most detailed description of inflammatory phenotype at COPD exacerbations, seasonality and infectious aetiology in longitudinal groups stratified by eosinophil levels. The seasonality of exacerbations has been previously described [21–24]. In our analysis we saw a clear seasonal pattern for all exacerbations; however, when focused on eosinophil-associated exacerbations, we report that these did not appear to show much seasonal variation, resulting in a larger proportion of exacerbations being associated with eosinophilic inflammation in the summer season. The aetiology of eosinophilic inflammation in COPD would appear to be driven by factors other than atopy as, although not formally tested, our patients had no recorded manifestations of atopic disease. Furthermore, the stable incidence of eosinophil-associated AECOPD throughout the year may suggest an intrinsic factor in triggering their incidence and requires further study.

It was previously reported that blood eosinophils $\geq 2\%$ identified sputum eosinophilic inflammation at exacerbation ($>3\%$), and was 90% sensitive and 60% specific [3]. Using the same cut-off for blood eosinophilic inflammation at enrolment in our analysis, it corresponded to the $>3\%$ sputum cut-off with similar sensitivity, but lower specificity (sensitivity 95.8% and specificity 31.8%). One reason for this discrepancy in specificity might be due to the difference in clinical states at the time of analysis; BAFADHEL *et al.* [3] conducted the analysis at exacerbations when sputum capture is greater, whereas in our study we conducted the analysis during clinical stability. We found that at exacerbation blood eosinophils $\geq 2\%$ was 79.6% sensitive and 53.9% specific. In our study the blood eosinophil count was reported up to one decimal and therefore it was not possible to apply the cut-off of $150 \text{ cells} \cdot \mu\text{L}^{-1}$. Using the $200 \text{ cells} \cdot \mu\text{L}^{-1}$ cut-off offered a similar sensitivity (95.8%), but inferior specificity (27.3%). We conducted a sensitivity analysis with the $\geq 200 \text{ cells} \cdot \mu\text{L}^{-1}$ cut-off and found that there was a significant difference in age, smoking history, exacerbation rate with raised eosinophils in the first year and presence of sputum eosinophils $>3\%$ at enrolment.

The rationale of our method to describe the longitudinal eosinophilic phenotype was to focus the analysis on subjects who were predominantly eosinophilic (allowance of one noneosinophilic event) as opposed to persistently eosinophilic (all visits were eosinophilic). This rule, we feel, represents a more pragmatic and “real-world” approach. Applying this rule, we demonstrated that 58% of subjects in our cohort had predominantly raised eosinophils over the period of 12 months. These subjects in the PE group were slightly older than those in the RE group ($p=0.034$) and there were trends towards a lower prevalence of current smokers, but with a greater smoking history in the PE group. SINGH *et al.* [4] studied longitudinal eosinophilic phenotype and reported that 37.4% of subjects had persistently elevated blood eosinophils $\geq 2\%$ at all visits over a period of 3 years. In a previous asthma study, a 90% rule was applied to identify persistent eosinophilic inflammation in sputum [25]. However, this rule was not applied in our cohort due to the limited number of samples available (maximum five samples). A limitation of the longitudinal phenotype method was that individuals with three visits could not be classified as intermittent eosinophilic (20 out of 99 for blood and 14 out of 80 for sputum).

Bacteria play an important role in exacerbations of COPD [26–30]. Bacterial exacerbation had been previously reported to be rarely associated with sputum eosinophilic exacerbation [3]. We investigated the prevalence of PPMs at exacerbation in the prospective eosinophilic groups and found that PPMs were far less common in the eosinophilic group. When we analysed the presence of PPMs at exacerbation in combination with blood eosinophilic inflammation, we found eosinophilic inflammation to be associated with reduced odds of PPM presence; although the magnitude of the difference was large (55% less likely), this was again of borderline significance ($p=0.049$). Therefore, understanding the clinical phenotype of stable inflammation may be a useful tool to stratify bacterial aetiology of exacerbations and hence antibiotic use.

While our detection of respiratory viruses at AECOPD corresponded with previously reported findings [30, 31], we observed no significant difference in the prevalence of respiratory viruses between the three groups at enrolment and at first exacerbation. Previously, PAPI *et al.* [30] reported airway eosinophilic inflammation to be a good predictor of viral infections. However, only severe exacerbations were included in their study, whereas we captured mild, moderate and severe events in our study. Further studies across the disease spectrum are required to ascertain the mechanisms linking infection and inflammatory patterns of disease. However, it is noteworthy that there was no significant difference in use of inhaled corticosteroids and bronchial reversibility across groups, similar to previously reported studies [3, 15].

It is important to recognise that these results are representative of a cohort of moderate to very severe COPD patients with frequent exacerbations receiving a high level of clinical intervention, including inhaled corticosteroids, as part of an intensive study. This might have had an impact on the severity of

exacerbations and recovery, as previous reports indicate that early therapy improves exacerbation outcomes [32]. Thus, the number of potential severe exacerbations might be smaller. In addition, the AERIS study was not originally designed, and therefore was not powered, to specifically investigate longitudinal eosinophilic inflammation and hence the numbers that were included in each eosinophilic group were limited. Larger prospective studies are required to further understand the impact of eosinophilic inflammatory status on a range of clinical outcomes.

In summary, this is the first study to report that eosinophilic inflammation is a stable phenotype in COPD and predictive of eosinophilic exacerbations. These events are seasonal in nature and relate to bacterial aetiology. Our data suggest that stratifying COPD patients into eosinophilic groups to potentially aid management is clinically relevant and potentially important, as is the consideration of season in management of exacerbations. Whether oral corticosteroids should be administered during exacerbations of COPD to predominantly eosinophilic patients, particularly outside the winter season, requires further investigation along with other stratification paradigms through well-designed intervention studies.

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References

- 1 WHO. The global burden of disease: 2004 update. 2004. www.who.int/healthinfo/global_burden_disease/2004_report_update/en/ Date last accessed: June 14, 2016.
- 2 Global Initiative for Chronic Obstructive Pulmonary Disease. Global Strategy for the Diagnosis, Management and Prevention of COPD. 2015. <http://goldcopd.org> Date last accessed: July 9, 2015.
- 3 Bafadhel M, McKenna S, Terry S, *et al.* Acute exacerbations of chronic obstructive pulmonary disease: identification of biologic clusters and their biomarkers. *Am J Respir Crit Care Med* 2011; 184: 662–671.
- 4 Singh D, Kolsum U, Brightling CE, *et al.* Eosinophilic inflammation in COPD: prevalence and clinical characteristics. *Eur Respir J* 2014; 44: 1697–1700.
- 5 Pizzichini E, Pizzichini MM, Gibson P, *et al.* Sputum eosinophilia predicts benefit from prednisone in smokers with chronic obstructive bronchitis. *Am J Respir Crit Care Med* 1998; 158: 1511–1517.
- 6 Brightling CE, Monteiro W, Ward R, *et al.* Sputum eosinophilia and short-term response to prednisolone in chronic obstructive pulmonary disease: a randomised controlled trial. *Lancet* 2000; 356: 1480–1485.
- 7 Eltboli O, Mistry V, Barker B, *et al.* Relationship between blood and bronchial submucosal eosinophilia and reticular basement membrane thickening in chronic obstructive pulmonary disease. *Respirology* 2015; 20: 667–670.
- 8 Vedel-Krogh S, Nielsen SF, Lange P, *et al.* Blood eosinophils and exacerbations in COPD: the Copenhagen General Population Study. *Am J Respir Crit Care Med* 2016; 193: 965–974.
- 9 Balzano G, Stefanelli F, Iorio C, *et al.* Eosinophilic inflammation in stable chronic obstructive pulmonary disease. Relationship with neutrophils and airway function. *Am J Respir Crit Care Med* 1999; 160: 1486–1492.
- 10 O'Shaughnessy TC, Ansari TW, Barnes NC, *et al.* Inflammation in bronchial biopsies of subjects with chronic bronchitis: inverse relationship of CD8+ T lymphocytes with FEV1. *Am J Respir Crit Care Med* 1997; 155: 852–857.
- 11 Hogg JC, Chu F, Utokaparch S, *et al.* The nature of small-airway obstruction in chronic obstructive pulmonary disease. *N Engl J Med* 2004; 350: 2645–2653.
- 12 Ronchi MC, Piragino C, Rosi E, *et al.* Role of sputum differential cell count in detecting airway inflammation in patients with chronic bronchial asthma or COPD. *Thorax* 1996; 51: 1000–1004.
- 13 Bafadhel M, Greening NJ, Harvey-Dunstan TC, *et al.* Blood eosinophils and outcomes in severe hospitalised exacerbations of COPD. *Chest* 2016; 150: 320–328.
- 14 Siva R, Green RH, Brightling CE, *et al.* Eosinophilic airway inflammation and exacerbations of COPD: a randomised controlled trial. *Eur Respir J* 2007; 29: 906–913.
- 15 Bafadhel M, McKenna S, Terry S, *et al.* Blood eosinophils to direct corticosteroid treatment of exacerbations of chronic obstructive pulmonary disease: a randomized placebo-controlled trial. *Am J Respir Crit Care Med* 2012; 186: 48–55.
- 16 Brightling CE, McKenna S, Hargadon B, *et al.* Sputum eosinophilia and the short term response to inhaled mometasone in chronic obstructive pulmonary disease. *Thorax* 2005; 60: 193–198.

- 17 Green RH, Brightling CE, McKenna S, *et al*. Asthma exacerbations and sputum eosinophil counts: a randomised controlled trial. *Lancet* 2002; 360: 1715–1721.
- 18 Bourne S, Cohet C, Kim V, *et al*. Acute Exacerbation and Respiratory InfectionS in COPD (AERIS): protocol for a prospective, observational cohort study. *BMJ Open* 2014; 4: e004546.
- 19 Wilkinson TMA, Aris E, Bourne S, *et al*. A prospective, observational cohort study of the seasonal dynamics of airway pathogens in the aetiology of exacerbations in COPD. *Thorax* 2017; 72: 919–927.
- 20 Pascoe S, Locantore N, Dransfield MT, *et al*. Blood eosinophil counts, exacerbations, and response to the addition of inhaled fluticasone furoate to vilanterol in patients with chronic obstructive pulmonary disease: a secondary analysis of data from two parallel randomised controlled trials. *Lancet Respir Med* 2015; 3: 435–442.
- 21 Hurst JR, Donaldson GC, Quint JK, *et al*. Temporal clustering of exacerbations in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2009; 179: 369–374.
- 22 Jenkins CR, Celli B, Anderson JA, *et al*. Seasonality and determinants of moderate and severe COPD exacerbations in the TORCH study. *Eur Respir J* 2012; 39: 38–45.
- 23 Wilkinson P, Pattenden S, Armstrong B, *et al*. Vulnerability to winter mortality in elderly people in Britain: population based study. *BMJ* 2004; 329: 647.
- 24 Johnston NW, McIvor A, Lambert K, *et al*. The Christmas season as a risk factor for chronic obstructive pulmonary disease exacerbations. *Can Respir J* 2010; 17: 275–281.
- 25 Walsh CJ, Zaihra T, Benedetti A, *et al*. Exacerbation risk in severe asthma is stratified by inflammatory phenotype using longitudinal measures of sputum eosinophils. *Clin Exp Allergy* 2016; 46: 1291–1302.
- 26 Sethi S, Wrona C, Eschberger K, *et al*. Inflammatory profile of new bacterial strain exacerbations of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2008; 177: 491–497.
- 27 Murphy TF, Sethi S, Niederman MS. The role of bacteria in exacerbations of COPD. A constructive view. *Chest* 2000; 118: 204–209.
- 28 Sethi S, Muscarella K, Evans N, *et al*. Airway inflammation and etiology of acute exacerbations of chronic bronchitis. *Chest* 2000; 118: 1557–1565.
- 29 Sethi S, Murphy TF. Bacterial infection in chronic obstructive pulmonary disease in 2000: a state-of-the-art review. *Clin Microbiol Rev* 2001; 14: 336–363.
- 30 Papi A, Bellettato CM, Braccioni F, *et al*. Infections and airway inflammation in chronic obstructive pulmonary disease severe exacerbations. *Am J Respir Crit Care Med* 2006; 173: 1114–1121.
- 31 Seemungal T, Harper-Owen R, Bhowmik A, *et al*. Respiratory viruses, symptoms, and inflammatory markers in acute exacerbations and stable chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2001; 164: 1618–1623.
- 32 Wilkinson TM, Donaldson GC, Hurst JR, *et al*. Early therapy improves outcomes of exacerbations of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2004; 169: 1298–1303.