

# Effect of growth hormone therapy on nitric oxide formation in cystic fibrosis patients

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ABSTRACT: Airway nitric oxide production is decreased in cystic fibrosis. As growth hormone therapy has been shown to increase nitric oxide production in growth hormone-deficient patients, it may also affect nitric oxide production in patients with cystic fibrosis. The objective of the present study was to investigate the effect of growth hormone therapy on systemic and airway nitric oxide formation in patients with cystic fibrosis.

Nitric oxide metabolites in serum and urine, amino acid concentrations in serum and sputum, as well as exhaled nitric oxide, were measured in children with cystic fibrosis before, during and after 1 yr of treatment with human growth hormone.

Nitric oxide metabolite concentrations increased significantly in serum and urine during the treatment period. Serum amino acid concentrations (including L-arginine, the substrate for nitric oxide synthases) also increased during treatment. The systemic bioavailability of L-arginine for nitric oxide synthases, expressed as ratio of L-arginine/L-ornithine+lysine, remained unchanged. In contrast, L-arginine concentrations in sputum decreased significantly during growth hormone treatment, as did exhaled nitric oxide levels.

Treatment with growth hormone in children with cystic fibrosis decreases exhaled nitric oxide by reducing the concentration of L-arginine in the airways.

KEYWORDS: L-Arginine, hormones, inflammatory marker, pulmonary function

ystic fibrosis (CF) is an inherited disease caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene. The clinical course of CF is variable but classically characterised by malabsorption, dystrophy secondary to pancreatic insufficiency, as well as chronic airway infection and inflammation leading to progressive loss of pulmonary function and ultimately respiratory failure [1]. CF airways are deficient for nitric oxide (NO). The mechanisms behind low NO formation in CF airways are incompletely understood but may include deficiency of L-arginine, the substrate of NO synthases (NOS), and decreased expression of NOS [2–5].

There is increasing evidence that low NO concentrations are functionally relevant in CF. Positive correlations between exhaled NO fraction (FeNO) and pulmonary function have been repeatedly observed [6, 7]. In addition, gene variants in neuronal NOS (NOS1), which are associated with low FeNO, seem to predispose to a more rapid decline in pulmonary function in CF children [8]. An increased risk for the colonisation of CF airways with Pseudomonas aeruginosa

was also found to be related to NOS1 and NOS3 gene variants associated with low FeNO [9, 10].

Conversely, L-arginine supplementation exhibits anti-inflammatory effects, as shown in a rat model of chronic Pseudomonas infection [11] and, when added to the airways, has dilatatory effects on airway smooth muscle in CFTR-deficient mice [12]. L-Arginine may also have protective effects on pulmonary oxidative stress and antioxidant defences [13]. In CF patients, L-arginine supplementation resulted in an increase in FeNO when given intravenously or orally [14, 15], and a single inhalation of nebulised L-arginine has been shown to increase FeNO and forced expiratory volume in one second (FEV1) in CF patients [16].

NO formation can be augmented by increasing the availability of L-arginine and also by increasing NOS expression [17]. Endocrine factors, such as oestrogen, insulin-like growth factor-1 and growth hormone have also been found to modify the L-arginine/NO pathway [18, 19]. Growth hormone is capable of transcriptional activation of NOS [20, 21] and may also affect substrate availability for NOS, as it enhances L-arginine

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uptake by the small intestine [22, 23]. Growth hormone deficiency results in decreased systemic NO formation, and replacement of growth hormone in deficient patients leads to increased NO, recovered urinary nitrate and cyclic guanosine monophosphate production, normalisation of elevated vascular peripheral resistance and improved cardiac output [24, 25].

Therapy with human growth hormone (hGH) has been shown to deploy beneficial effects for CF patients, including enhanced growth, weight gain and bone mineralisation [26–28]. Since growth hormone replacement resulted in increased NO metabolism in growth hormone-deficient patients, the current authors hypothesised that growth hormone therapy in CF patients would also result in increased NO formation. Therefore, the effect of hGH therapy on L-arginine metabolism and NO formation was studied in CF patients undergoing hGH therapy as part of a multicentre trial to assess the effect of hGH therapy in CF [26].

## **MATERIALS AND METHODS**

## Study design

Patients included in the present study participated in a randomised placebo-controlled, multicentre study on the effects of hGH therapy in CF (Pharmacia trial 307 MET-9002-026) [26]. Patients were treated with recombinant hGH by daily subcutaneous injection of 0.039 mg·kg<sup>-1</sup> body weight per day (~0.11 IU·kg<sup>-1</sup> body weight per day), 0.07 mg·kg<sup>-1</sup> body weight per day (~0.21 IU·kg<sup>-1</sup> body weight per day) or placebo. The study was conducted as a 24-week double-blind study with a 24-week open-label treatment period. Patients were assigned to one of three treatment arms for 24 weeks. At the end of the double-blind treatment period, patients on growth hormone therapy were maintained on their current growth hormone dosage for an additional 24 weeks, and patients in the placebo group were randomly assigned to either the low- or the highgrowth hormone-dosage treatment regimen. Subjects had to have an established diagnosis of CF, dystrophy defined by a body mass index (BMI) <10th and/or weight <3rd percentile, despite high caloric intake (>120% recommended dietary allowance), and a bone age of 8-18 yrs. Exclusion criteria were acute pulmonary exacerbation, diabetes and liver cirrhosis. Patients were studied before initiation of hGH treatment, every 3 months during therapy, and 6 and 12 months after the treatment period.

## Study population

In total, 15 CF patients from a single centre (Essen, Germany), who had all participated in the multicentre study, were included in the present substudy. Of these, 11 patients were treated with hGH for 12 months; six patients with low dose and five with high dose of hGH. Four patients, who were assigned to the placebo group and therefore did not receive hGH for 12 months, were not included in the analysis. Mean (range) age in the 11 patients was 14 (12-18) yrs, FEV1 was 53 (28-90)% predicted, and BMI SD score was -1.7 (-3.7--0.9); these were not different from the total study population [26]. NO metabolites (NOx) were measured in both serum and urine before, during (at 3, 6, 9 and 12 months) and 12 months after hGH therapy. Serum amino acids were measured before and during (6 and 9 months) hGH therapy. Sputum was primarily collected for microbiology. Amino acid concentrations in sputum were measured in five hGH-treated patients able to provide additional samples during the blinded phase of the study (before and after 3 and 6 months) as well as 6 months after hGH therapy. Exhaled NO was measured in all patients before, during (3, 6, 9 and 12 months) and 6 months after hGH treatment. Feno after 12 months of hGH therapy was only measured in four patients and therefore these data were not analysed.

# Serum and sputum sample processing

Blood was drawn by venous puncture, immediately centrifuged and serum was stored at -20°C until analysed. Spontaneously expectorated sputum was used primarily for microbiology. Additional samples were diluted with buffer 1:1 and vortexed for 2 min, followed by 10 min of centrifugation at  $1,000 \times g$ . Clear supernatant was used for amino acid measurements.

#### NOx measurements

Total NO metabolites (nitrate, nitrite and nitrosothiols) were measured after conversion to NO by using vanadium (III) chloride in hydrochloric acid at 90°C with an NO analyser (NOA 280 and purge vessel; Sievers, Boulder, CO, USA) [29].

## Amino acid measurements

Amino acids were determined by ion exchange chromatography on an amino acid analyser LC 3000 (Eppendorf, Hamburg, Germany) according to the manufacturer's specifications. Serum and sputum samples were deproteinised before analysis.

#### **Exhaled NO measurement**

A chemiluminescence analyser (NOA 280; Sievers) was used to measure  $F_{\rm eNO}$ . Single breath on-line measurements for the assessment of lower airway NO were performed at a constant expiratory flow of 50 mL·min<sup>-1</sup> in accordance with published European Respiratory Society/American Thoracic Society standards [30, 31]. The NO analyser was calibrated before each study with 0 and 185 ppb NO calibration gas (Linde AG, Unterschleissheim, Germany). The mean of three end-expiratory NO concentrations within a variation of 15% was calculated for each subject.

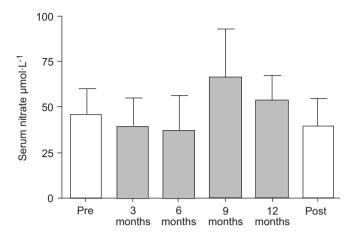
# **Statistics**

Data are shown as median (interquartile range). Comparisons of time courses in NOx and amino acid concentrations, as well as *F*eNO, were performed using the Kruskal–Wallis one-way ANOVA. Comparison of L-arginine bioavailability index (L-arginine/L-ornithine+lysine) in serum *versus* sputum was performed using the Wilcoxon test.

## **RESULTS**

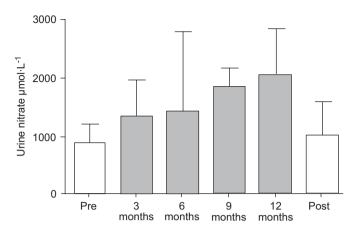
There was no difference between the low- and high-hGH treatment groups in any of the parameters analysed. Serum NOx concentrations increased significantly (p=0.027, ANOVA) during the treatment period and returned to baseline values after hGH was discontinued (fig. 1). Similarly, urine NOx concentrations also increased significantly (p=0.029, ANOVA) during treatment and returned to baseline after hGH was discontinued (fig. 2).

Serum concentrations of 18 amino acids measured before and during hGH therapy are displayed in table 1. With the exception of threonine, mean levels of all amino acids in serum significantly increased during hGH treatment. This



**FIGURE 1.** Nitric oxide metabolite (NOx) concentrations in serum before, during and after 12 months of human growth hormone (hGH) treatment in cystic fibrosis patients (n=11). Serum NOx increased during the treatment period and returned to baseline values after hGH was discontinued.

included L-arginine and L-citrulline, the substrate and the product of NOS activity, respectively, as well as L-ornithine, the product of arginase, and proline, a downstream product of the arginase pathway and precursor of collagen formation (table 1). The ratio of L-arginine/L-ornithine+lysine, which can be used as an index of bioavailability of L-arginine for NOS at a given L-arginine concentration [32], was  $0.45\pm0.08$  in serum before treatment and remained unchanged during hGH treatment (p=0.347, ANOVA).



**FIGURE 2.** Nitric oxide metabolite (NOx) concentrations in urine before, during and after 12 months of human growth hormone (hGH) treatment in cystic fibrosis patients (n=11). Urine NOx concentrations increased during the treatment period and returned to baseline values after hGH was discontinued.

Mean  $F_{\text{eNO}}$  decreased significantly during the treatment period (p=0.005, ANOVA) and returned to baseline after hGH was discontinued (fig. 3).

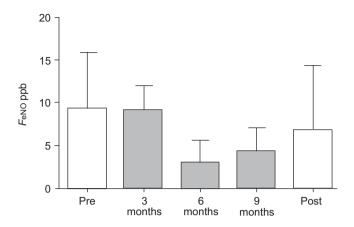
L-Arginine concentration in CF sputum was  $15.5\pm17.3~\mu\text{mol}\cdot\text{L}^{-1}$  at baseline and decreased significantly during hGH treatment (p=0.008, ANOVA; fig. 4a). The L-arginine availability index at baseline was significantly lower in CF sputum than in serum (p<0.001, Wilcoxon test). Individual changes in L-arginine bioavailability index in sputum are displayed in figure 4b.

**TABLE 1** Serum amino acid concentrations in 10 cystic fibrosis patients before and 6 and 9 months after the initiation of therapy with human growth hormone

Amino acid	Concentration μmol·L <sup>-1</sup>			p-value#
	Pre-treatment	6 months	9 months	
L-Arginine	105.6 (92.5–127.6)	110.8 (100.7–122.3)	138.4 (114.1–165.5)	0.027
L-Ornithine	79.3 (54.5–116.9)	75.1 (63.2–94.9)	126.0 (89.5–135.5)	0.038
L-Citrulline	24.5 (18.0–32.4)	22.7 (13.4–25.4)	37.5 (27.1–45.7)	0.006
Threonine	140.5 (102.8–205.3)	128.1 (100.9–167.4)	206.3 (133.8–235.5)	0.090
Serine	132.7 (112.8–171.5)	151.4 (137.6–171.4)	191.3 (168.2–222.3)	0.002
Proline	166.2 (125.9–242.4)	148.6 (115.7–194.6)	224.6 (178.1–254.8)	0.028
Taurine	137.6 (123.9–166.2)	172.3 (136.5–193.8)	175.8 (154.5–203.4)	0.032
Glycine	280.3 (235.7–403.9)	343.9 (312.3–410.7)	436.7 (369.7–485.6)	0.005
Alanine	387.8 (301.7–450.5)	415.4 (365.5–461.0)	469.6 (407.7–573.9)	0.042
Valine	161.1 (148.4–187.1)	165.2 (145.9–200.8)	237.0 (187.2–271.1)	0.006
Methionine	8.3 (3.6–11.6)	5.1 (3.6–8.7)	10.4 (7.5–16.3)	0.016
Isoleucine	47.1 (35.5–88.3)	42.9 (36.3–55.3)	83.0 (65.1–98.8)	0.008
Leucine	79.1 (56.4–107.6)	88.0 (77.8–109.1)	140.6 (114.3–181.5)	0.002
Tyrosine	43.7 (32.2–80.7)	44.4 (40.9–52.3)	67.2 (51.6–76.0)	0.048
Phenylalanine	54.7 (40.2–70.9)	51.8 (44.3–65.4)	77.5 (67.4–94.9)	0.013
Histidine	68.0 (57.8–87.9)	75.0 (66.6–86.3)	94.4 (83.8–109.4)	0.005
Tryptophan	45.3 (35.3–49.6)	40.3 (32.8–55.8)	56.3 (47.3–64.8)	0.031
Lysine	134.6 (125.5–205.3)	131.3 (118.3–161.5)	208.6 (151.1–230.9)	0.018

Data are presented as mean (interquartile range), unless otherwise stated. #: Kruskal-Wallis ANOVA.

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**FIGURE 3.** Fractional exhaled nitric oxide (*F*eNO) in cystic fibrosis patients before, during and after 12 months of human growth hormone (hGH) treatment (n=11). Mean *F*eNO decreased during the treatment period and returned to baseline values after hGH was discontinued.

Although this index decreased in individual patients, the mean sputum L-arginine bioavailability index did not change significantly during hGH treatment (p=0.275, ANOVA).

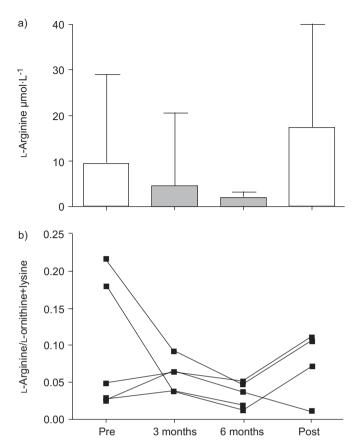
There was no correlation between pulmonary function data and any of the parameters analysed.

## **DISCUSSION**

In the present study, treatment of CF patients with hGH resulted in increased serum amino acid concentrations, including L-arginine, the substrate for NO synthases, as well as increased NOx concentrations in both serum and urine. These findings suggest increased vascular or systemic formation of NO during hGH therapy in CF patients and parallel those in patients deficient for growth hormone, where hGH replacement therapy increased systemic NO formation [25]. In contrast, concentrations of L-arginine in sputum as well as exhaled NO levels decreased during hGH treatment, suggesting decreased substrate availability for NOS and decreased NO formation in airways of CF patients treated with hGH.

Increased NO production during hGH treatment, as evidenced by increased NOx in serum and urine, could result from effects of growth hormone on the expression or activity of constitutive or inducible NOS isoforms. Experiments in cultured human endothelial cells had shown that growth hormone resulted in a dose- and time-dependent increase in endothelial NOS (NOS3) gene and protein expression [20]. In a study in cultured mesangial cells, growth hormone treatment was also found to result in a dose-dependent increase in inducible NOS (iNOS; NOS2) transcript [21]. iNOS is of interest in CF because, despite the inflammatory nature of the disease, its expression is decreased in CF airway epithelial cells [3, 4]. The mechanisms resulting in decreased iNOS expression in CF are not currently understood but even activation with pro-inflammatory cytokines, which results in profound iNOS activation in non-CF cells, has limited to no effect on iNOS expression in CFTR-deficient cells [5]. Translation of NOS2 mRNA can also be downregulated by low concentrations of L-arginine [33, 34].

Another plausible explanation for higher systemic NOx concentrations in CF patients treated with hGH is increased



**FIGURE 4.** a) L-Arginine and b) L-arginine bioavailability index in sputum of cystic fibrosis patients before, during and after 12 months of human growth hormone (hGH) treatment (n=5). L-Arginine concentrations decreased during hGH treatment, while the mean L-arginine availability index did not change.

enzymatic NO formation in response to higher substrate availability. L-Arginine conversion by NOS is known to increase with higher extracellular L-arginine levels, although the intracellular concentrations of L-arginine are usually well above the Michaelis constant for NOS. This phenomenon is referred to as the L-arginine paradox [35] and is thought to be responsible for the increase in  $F_{\rm eNO}$  in healthy subjects and CF patients supplemented with L-arginine [14, 15]. Therefore, the observed increase in NOx levels may be directly related to the circulating concentrations of L-arginine, as both L-arginine and NOx increased in serum.

While systemic L-arginine and NOx production increased in CF patients during growth hormone therapy, at the same time NO in the exhaled air decreased. *F*<sub>e</sub>NO is used as a surrogate marker for NO production in airways; thus, low *F*<sub>e</sub>NO would suggest decreased airway NO production. Alternatively, retention of NO in airway secretions and conversion to metabolites, such as nitrite or nitrate, could result in low *F*<sub>e</sub>NO in the presence of unchanged, or even increased, enzymatic NO production. This mechanism had previously been suggested to be one contributing factor to low *F*<sub>e</sub>NO in CF patients [36, 37]. However, in the present study it was found that treatment with hGH resulted in decreased substrate availability for NOS in CF airways. Although, due to the small sample size, these results need to be interpreted with caution, it was found that not only absolute

sputum L-arginine concentration, but also the relative bioavailability of L-arginine for NOS, decreased in CF sputum during therapy with hGH. The L-arginine bioavailability index (Larginine/L-ornithine+lysine) reflects the availability of L-arginine for NOS at a given L-arginine concentration, as L-arginine, L-ornithine and lysine utilise the same transmembrane transport system, the cationic amino acids transporter (CAT)-2 [32] for cellular internalisation. This index has recently been shown to be decreased in the blood of patients with asthma, sickle cell disease and CF, and is thought to reflect substrate limitation for NOS-mediated smooth muscle relaxation in these conditions [2, 32, 38]. Even at baseline, this index was significantly lower in sputum compared with serum in CF patients. This reduced Larginine availability for CF airway cells may potentially affect the ability of upregulating NO as part of a defence mechanism against airway pathogens sensitive to NO-mediated killing, such as mucoid P. aeruginosa [39].

Chronic infection in CF results in neutrophil-dominated inflammation and increased concentrations of inflammatory markers in the airways [1]. However, activation of signal transducer and activator of transcription (STAT)1 is reduced in CF, and the interferon (IFN)-γ signalling pathway, which involves STAT1 and leads to NOS2 gene induction in airway epithelial cells, is defective in CF [40]. Therefore, inhibitory effects of growth hormone on these pathways may further decrease the expression of NOS2 in CF. Indeed, growth hormone was found to suppress the synergistic induction of iNOS by the cytokines IFN- $\gamma$  and tumour necrosis factor- $\alpha$  in a rat insulin-secreting cell line (INS-1 cells). This effect was thought to be mediated through inhibition of IFN-γ-activated STAT1 by growth hormone [41]. In contrast, iNOS enzyme activity in liver tissue from calves undergoing lipopolysaccharide (LPS) challenges was not affected by pretreatment with growth hormone. However, growth hormone treatment before LPS resulted in an increase in constitutive NOS activity, the phosphorylation of NOS3, plasma NOx levels and liver protein nitration, in growth hormone-treated compared with nongrowth hormone-treated animals. Of interest, the progressive increase in tissue CAT-2 mRNA after LPS was significantly reduced with growth hormone treatment in this model [42], potentially contributing to substrate limitation for NOS. Although not vet studied in CF, these studies may suggest that growth hormone therapy in CF patients could have significant effects on NO formation by affecting the expression of NOS2 and by reducing L-arginine availability for NOS.

Low airway NO concentration may not only reflect decreased production but may also result from the reaction of NO with reactive oxygen species (ROS). Activated NOS can release ROS, such as the superoxide anion and hydrogen peroxide at nonsaturating L-arginine levels [43, 44], and NOS can generate NO and superoxide anion at the same time at low-saturating L-arginine levels [45]. These products may react to generate peroxynitrite and subsequently result in tyrosine nitration. Nitrotyrosine had been previously found to be increased in CF lung tissue, sputum and exhaled breath condensate [46]. Previous studies had suggested that exogenous growth hormone primes human and rat neutrophils, monocytes and macrophages for a respiratory burst and an accompanying increase in superoxide production [47, 48]. In contrast, NO release by cultured endothelial cells treated with growth

hormone was only increased at growth hormone concentrations high enough to also result in a significant reduction of ROS formation. At lower concentrations, the expression of NOS3 was increased but NO was metabolised by reaction with ROS [20]. Therefore, the effect of growth hormone on NO production may be dose dependent, and higher doses than those used in the present study may potentially have different effects on pulmonary NO production.

In summary, the results of the present study suggest that treatment with human growth hormone results in decreased airway nitric oxide formation in patients with cystic fibrosis. Further studies are needed to clarify whether this observation is specific for cystic fibrosis and to specify the pathophysiological consequences of the observed changes in the L-arginine/nitric oxide metabolic pathway.

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