The leukotriene-antagonist ICI-204,219 inhibits the early airway reaction to cumulative bronchial challenge with allergen in atopic asthmatics


ABSTRACT: The hypothesis that cysteinyl-leukotrienes (LTC₄, LTD₄, and LTE₄) are mediators of allergen-induced airway obstruction in asthmatics was tested with the specific receptor antagonist ICI-204,219, in a double-blind, placebo-controlled, randomized, cross-over bronchoprovocation study.

On three occasions, cumulative bronchial challenge with specific allergen was performed in 10 males with mild allergic asthma. The first control session established the baseline provocative dose of allergen producing a decrease in forced expiratory volume in one second (FEV₁) by 20% (PD₂₀FEV₁). The two rechallenges were performed 2 h after oral administration of placebo or 20 mg of ICI-204,219.

The allergen dose-response relations were highly reproducible, producing PD₂₀ values at the control session and after placebo treatment which varied by no more than 0.7–1.3 fold (95% confidence interval (95% CI)). After ICI-204,219, the median cumulated allergen dose was 5.5 fold higher, and the group geometric mean PD₂₀ was increased 2.5 times. Furthermore, the recovery time after the immediate bronchoconstriction was shorter (40 vs 60 min). The wheal and flare responses to intradermally injected LTD₄ were somewhat inhibited by ICI-204,219, whereas responses to histamine were unaffected. However, the findings suggest that skin testing with LTD₄ is unlikely to predict the degree of leukotriene-antagonism in the airways.

The findings confirm and extend the indications that cysteinyl-leukotrienes are important mediators of allergen-induced airway obstruction, and that leukotriene-antagonists should be evaluated as a potential new therapy in allergic asthma.

Eur Respir J., 1994, 7, 324–331.
of allergen-induced bronchoconstriction and excretion of urinary LTE₄ [26], whereas the more potent inhibitor of leukotriene biosynthesis, MK-886, produced substantial inhibition of urinary LTE₄ and of the allergen-induced airway obstruction in asthmatics [27].

The purpose of this study was to confirm earlier indications [23, 24] that the orally active and selective receptor antagonist of cysteinyl-leukotrienes, ICI-204,219 (Accolate) [28, 29], could inhibit allergen-induced airway obstruction in atopic asthmatics. We focused on the early airway reaction (EAR) and tested the effect of ICI-204,219 on the provocative dose (units) required to produce a decrease of forced expiratory volume in one second (FEV₁) by 20% (PD₂₀FEV₁), determined by cumulative challenge with increasing doses of allergen. It is expected that, in the presence of the receptor antagonist, a higher dose of the agonist is required to evoke the 20% decrease in FEV₁, which would result in a displacement of the dose-response relation for allergen to the right. The displacement factor may be used as a quantitative measure of drug potency; enabling the protective effect of the antagonist on allergen-induced airway obstruction to be directly related to its potency as an antagonist of LTD₄-induced bronchoconstriction. This approach has been used in studies with H₁-receptor antagonists [30]. Another objective of the study was to evaluate whether skin responses to LTD₄ could be used to estimate the degree of leukotriene-antagonism achieved by the particular dose (20 mg) of ICI-204,219 used in this study.

Methods

Patients

Ten nonsmoking atopic men, with a history of asthma and with specific allergy to birch pollen, cat or dog dander, participated in the study. The study was approved by the Local Ethics Committee and the Swedish Drug Regulatory Agency (Läkemedelsverket). The patients gave informed consent. For inclusion in the study, bronchial reactivity to histamine was required, with a provocative concentration causing a 20% fall in FEV₁ (PC₂₀) determined as described elsewhere [31].

Table 1. – Patient characteristics

<table>
<thead>
<tr>
<th>Pt No.</th>
<th>Age yrs</th>
<th>% pred</th>
<th>Histamine PC₂₀ mg·ml⁻¹</th>
<th>Allergen</th>
<th>Medication during study</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>28</td>
<td>92</td>
<td>4.00</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>34</td>
<td>105</td>
<td>0.55</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>28</td>
<td>102</td>
<td>1.10</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>28</td>
<td>101</td>
<td>3.20</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>41</td>
<td>109</td>
<td>2.80</td>
<td>Cat</td>
<td>IB, IS</td>
</tr>
<tr>
<td>6</td>
<td>31</td>
<td>90</td>
<td>0.52</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>23</td>
<td>86</td>
<td>0.62</td>
<td>Cat</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>56</td>
<td>88</td>
<td>0.30</td>
<td>Birch</td>
<td>IB, IS</td>
</tr>
<tr>
<td>9</td>
<td>26</td>
<td>86</td>
<td>1.35</td>
<td>Dog</td>
<td>DSCG</td>
</tr>
<tr>
<td>10</td>
<td>42</td>
<td>77</td>
<td>0.62</td>
<td>Dog</td>
<td></td>
</tr>
</tbody>
</table>

FEV₁: forced expiratory volume in one second; PC₂₀: provocative concentration of histamine causing a 20% fall in FEV₁; Pt: patient; IB: inhaled beta-stimulant; IS: inhaled steroid; DSCG: inhaled disodium cromoglycate. *: geometric mean.

Study design

All patients first underwent a control allergen bronchoprovocation, in order to demonstrate sensitivity of the airways to allergen, and to establish their PD₂₀FEV₁. Within four months of the control challenge (average interval 46 days, range 14–104 days) the double-blind, placebo-controlled, cross-over drug trial was started. On two separate days, the patients received either 20 mg of ICI-204,219 or placebo tablets 2 h before allergen challenge. Patients always reported to the clinic at the same time of the day (i.e. 7:30 am). They were allowed a light breakfast while still at home. The two bronchoprovocations were separated by an interval of 2–6 weeks, and the same protocol for allergen dose increments was used at both occasions. For each challenge, the starting allergen dose was selected as 0.5–1 log below the PD₂₀FEV₁ determined with the same protocol at the control session. The patients always returned to the clinic the morning after each treatment day for general follow-up, including physical examination, spirometry, and blood sampling for routine haematology and blood chemistry.

Throughout the study, patients continued their regular asthma therapy (table 1), except for withholding disodium cromoglycate and inhaled β-agonists for 8 h before a study day. Short-acting histamine H₁-receptor antagonists were not allowed for 24 h, and nonsteroidal anti-inflammatory drugs (NSAIDs) were not allowed for 10 days prior to a challenge session.

Allergen provocation and study day procedures

Bronchoprovocation was performed by inhalation of allergen, using a dosimeter-controlled jet nebulizer (Spira Elektro 2, Respiratory Care Center, Finland and Alfax, Sweden [32]), to permit determination of PD₂₀FEV₁. Driven by compressed air at 7.5 l·min⁻¹, the nebulizer generated an aerosol with a mass-mean particle aerodynamic diameter of 4.1 µm. With a 0.5 s nebulization period, the output was 7.1 µl per breath. Pulmonary function was measured as FEV₁ on a spirometer (Vitalograph MDI Compact, Förbandsmaterial, Sweden), and for monitoring of possible late asthmatic reactions (LARs) measurements of peak expiratory flow rate (PEFR) were obtained using a mini-Wright peak flow meter. Three concentrations of allergen extract, 1,000, 10,000 and 100,000 biological units·ml⁻¹, were prepared from powder dissolved in Pharmalgen albumin diluent. The extracts and the diluent were from Pharmacia AB, Uppsala,
Switzerland. By using the three concentrations, and by varying the number of breaths from the nebulizer, a protocol was developed creating approximately half-log increments in the cumulated dose of allergen (7, 21, 71, 213, 710, 2,130 and 7,100 biological units). Baseline FEV$_1$ was defined as the best of three recordings, 5 min apart. The challenge was started by inhalation of the diluent and, providing FEV$_1$ did not change by more than 10%, ICI-204,219 or placebo was ingested. Spirometry was obtained at 30, 60, 90 and 120 min after drug intake, the 120 min FEV$_1$, value being used as baseline for the ensuing allergen challenge. Incremental doses of allergen were inhaled every 20 min until FEV$_1$ had fallen by at least 20%. Dose-response relations for the cumulated dose of allergen were constructed and the PD$_{20}$FEV$_1$ was determined by linear interpolation. After a positive reaction, spirometry was followed every 15 min until FEV$_1$ had returned to within 10% of baseline, and there after hourly for up to 6 h. The patients were asked to measure their PEFR hourly after discharge from hospital, until retiring to bed. The occurrence of a LAR was defined as a 15% drop in PEFR 3–12 h after the EAR.

**Skin tests**

Histamine (10 nmol), LTD$_4$ (0.1, 1 and 10 nmol) and saline were injected at the proximal parts of the volar aspects of the forearms in 50 μl volumes, delivered by 0.3 ml microfine insulin syringes (Benson-Dixon AB, Stockholm). The injections were made after the occurrence of the maximal airway response, i.e. 3–4 h after intake of placebo or active drug in the blinded sessions. The size of the wheal and flare responses were recorded at 5, 10, 20 and 30 min after injection, and delineated with a water-resistant fine-tip pen on a transparency sheet placed above the injected sites. The transparencies have been filed for documentation of the data. The injections and the skin monitoring were conducted by an investigator who was not involved in the bronchial provocation tests. The injection sites were monitored until the patients left the ward, usually about 4 h after the injection, and at the next morning visit (vide supra) the patients were asked about the occurrence of skin reactions during the night. After the experiments, the wheal and flare mean arithmetic diameter was calculated for each time-point, by measurements of the vertical and perpendicular diameters in mm. These measurements were made by an investigator who was unaware of the study code and who had not performed the skin tests. For evaluation, the wheal and flare responses to the injected substances were expressed as the sum of the respective measurements at the four time-points (5, 10, 20 and 30 min). Preliminary experiments indicated that this total index of the reactions gave information which was closely similar to the data obtained by planimetry and calculation of the area under the time-response curve. Histamine dihydrochloride was purchased from Sigma and dissolved in saline. Synthetic LTD$_4$ (GMP-grade) was purchased from Cascade Biochem Ltd (Berkshire, UK) and dilutions from a concentrated stock solution (4.5 mM in ethanol:water 1:1) were made in saline on each experimental day, according to a fixed protocol. The concentration of the stock solution was regularly checked by UV-spectrometry. The concentration of ethanol in the injected solutions was kept below 1%, which had been shown to cause no skin reaction.

**Statistics**

Calculations of group mean and median PD$_{20}$FEV$_1$ values were performed on log transformed raw data. Analysis of variance was applied, and when data were found to be following a normal distribution pattern, the two-tailed paired Student's t-test was used to compare group means after placebo and active drug. Otherwise, the groups were compared with two-tailed Wilcoxon's paired test. Differences were considered to be significant when p was <0.05. The repeatability of the challenge procedure was evaluated by comparing logarithmically transformed PD$_{20}$FEV$_1$ values at control and placebo sessions, using the method described by Bland and Altman [33].

**Results**

**Bronchial challenge testing**

As a representative illustration of the findings in the study, the time-course of the airway response during the two blinded allergen-provocations are shown for subject No. 3 (fig. 1). Firstly, the baseline pulmonary function did not differ significantly between the two challenge sessions. For the group as a whole, the postdiluent FEV$_1$ values, which were used as baseline before drug intake, were 4.1±0.6 and 4.2±0.5 l (mean±SD) on placebo and
ICI-204,219 treated days, respectively. Secondly, there was no significant change in FEV₁ during the 2 h following ingestion of the active drug. For the group, FEV₁ immediately before the start of challenge was 102±3% of baseline on the day active drug was given, as compared with 100±2% after placebo (mean±SD).

Despite these similarities in baseline characteristics, the two challenge sessions were quite different, since one or two half-log increments in the dose of inhaled allergen (fig. 1) was usually required to decrease FEV₁ by at least 20% after ICI-204,219 (median allergen dose 2,130 (range 49–7,100) units) as compared with placebo (median allergen dose 389 (range 21–2,130) units). For the whole group there was a highly significant 5.5 fold increase in the median allergen dose after ICI-204,219.

Seven of the 10 subjects showed a clear-cut rightward shift of the dose-response relation for allergen in the presence of active drug (fig. 2). The mean shift in allergen PD₂₀FEV₁ for these seven subjects was 0.59 (range 0.29–1.00) log units, corresponding to an approximately four fold increase (range 2–10 fold) in the presence of the leukotriene-antagonist. For the remaining three subjects, the difference in PD₂₀ between placebo and active drug was within the expected range of variation between two allergen provocations (±0.12 log units, vide infra).

Nevertheless, for the group as a whole, there was a significant increase (p=0.01; Wilcoxon’s paired test) in the PD₂₀FEV₁ for allergen in the presence of ICI-204,219.

The magnitude of the maximal EAR produced by the cumulative challenge was similar at the two sessions (drop in FEV₁ being 32±11 and 28±6% for active drug and placebo, respectively (mean±SD); p=0.1).

The subjects in this study with respect to disease parameters, such as histamine PC₂₀, allergen PD₂₀FEV₁ and baseline FEV₁% predicted, were quite small (table 1), possibly explaining why there were no clear correlations between the responses to the active drug and any of these parameters or their sum (Pearson’s correlation coefficient r≤0.6; p>0.1).

The magnitude of the maximal EAR produced by the cumulative challenge was similar at the two sessions (drop in FEV₁ being 32±11 and 28±6% for active drug and placebo, respectively (mean±SD); p=0.1).
as the time required for FEV₁ to return from the peak drop to a value within 90% of the prechallenge baseline, was shorter in the group receiving ICI-204,219 (40±11 min for active drug versus 60±26 min for placebo (mean±SD); p<0.05, Student’s paired t-test.). Neither in the presence of placebo nor active drug was rescue treatment required during the EAR.

After being discharged from the clinic, the patients continued to record their PEFR and symptoms for up to 12 h after the provocation. Six LARs occurred in five of the subjects after one or both of the blinded sessions, with an equal distribution between placebo (3 out of 6) and active drug (3 out of 6).

There were no subjective or objective signs of drug-related side-effects after this single 20 mg dose of ICI-204,219.

Repeatability

As indicated in figure 3, there was good agreement for the group as a whole between the PD₂₀FEV₁ values determined at the first control provocation and those obtained during the placebo session. The correlation between the individual PD₂₀FEV₁ values at the two sessions was indeed excellent (fig. 4a). Likewise, the group shift in PD₂₀FEV₁ values between the control and placebo session was negligible (0.04±0.19 log units as compared with 0.40±0.36 log units between ICI-204,219 and placebo (mean±SD)). When tested for repeatability according to Bland and Altman [33], by plotting logarithmically transformed PD₂₀FEV₁ values for the two sessions as the difference in results against their mean, it was further documented that the allergen provocation was highly repeatable (fig. 4b). Thus, in this group of 10 atopic men, the 95% CI obtained for the differences between the first control challenge and the challenge in the presence of placebo was 0.12 log units, corresponding to an expected variation in PD₂₀FEV₁ which was no more than 0.7- to 1.3-fold. This variation is less than previously reported for methacholine, with the same dosimeter [32].

Skin testing

Intradermal injection of LTD₄ produced a wheal and flare response, which was reproducible. For example, the wheal response to 1 nmol of LTD₄ was 40.6±4.0 mm at the control session and 39.8±4.6 mm after placebo (mean±SD), as compared with 57.8±13.3 and 57.7±6.9 for histamine 10 nmol at the same two sessions. The flare evoked by LTD₄ was consistently more pale than that produced by equi-active doses of histamine. The time course for the onset of the responses to histamine and LTD₄ were, however, very similar, and neither compound caused a late cutaneous reaction. Histamine was more potent on a molar basis, but it was not possible to calculate a dose-ratio between the compounds from the present observations, because the dose-response curve for LTD₄ was much shallower than that of histamine. In particular, there was no significant difference between the wheal responses to 0.1, 1 and 10 nmol of LTD₄ (fig. 5a), although all responses were significant compared with saline controls. The flare response to LTD₄, however, showed some dose-dependency (fig. 5b).

The wheal response to LTD₄ was reduced by about 15% at each of the two highest dose levels after ICI-204,219 (fig. 5a). The flare was inhibited by ICI-204,219 to a slightly greater extent (20–30%) (fig. 5b). For the flare, a mean rightward shift amounting to 0.43 log units could be calculated for the group as a whole (table 2). When analysed for the individual subjects, there was no correlation (Pearson correlation coefficient r=0.51; p=0.13) between the shift in the dose-response relation for LTD₄ in the skin after ICI-204,219 and the shift in PD₂₀FEV₁ for allergen in the airways (table 2).

Skin responses to histamine were not inhibited by ICI-204,219. The whealing produced by histamine 10 nmol was 63.1±5.7 mm after ICI-204,219 and 57.7±6.9 mm after placebo (mean±SD; p>0.1; Student’s paired t-test). The corresponding values for the flare were 199.7±35.1 mm and 205.4±25.9 mm (p>0.1; Student’s paired t-test), respectively.

![Figure 4](image-url)  
Fig. 4. – Illustration of the good repeatability of the allergen challenge in the study group. a) Correlation of the PD₂₀FEV₁ values at the control challenge and the placebo treated challenge for each subject. Pearson correlation coefficient r=0.96 (p=0.00001) for two-tailed probability of no correlation. b) The difference in log PD₂₀FEV₁ between the two challenge sessions is plotted against their mean according to the method of Bland and Altman [33].
In this study, the competitive receptor antagonist of cysteinyl-leukotrienes, ICI-204,219 [28] caused substantial inhibition of several aspects of allergen-induced airway obstruction. For the group as a whole, ICI-204,219 caused a significantly higher mean PD20FEV1 value, as well as a higher total dose of allergen required to reach the predetermined level of bronchoconstriction. Despite this higher dose of allergen, the recovery after challenge was faster in the presence of the leukotriene-antagonist, and there was no increase in the occurrence of LARs.

The study thus documents that a lower dose (20 mg) of ICI-204,219 than previously tested (40 mg) [23, 24] can inhibit allergen-induced bronchoconstriction. The reproducible and dose-dependent inhibition of various aspects of the airway responses to allergen-challenge in asthmatics by different second generation anti-leukotriene drugs ([23–25, 27], and this study), strongly supports that leukotrienes are significant mediators of allergen-induced airway obstruction in man.

The finding that ICI-204,219 increased the dose of allergen required to reach a defined response, is consistent with the mode of action of a specific receptor antagonist. In order to surmount the effect of the antagonist at the receptors for leukotrienes in the airways, more mediator must be released by increasing the stimulus for mediator release, i.e., the allergen dose. In fact, we have evidence supporting the fact that the higher doses of allergen after active drug were associated with increased formation of leukotrienes. Thus, in six of the subjects in this study, urinary LTE4 was followed as an index of pulmonary release of cysteinyl-leukotrienes [18]. The postchallenge increase in urinary LTE4 was consistently higher in the presence of ICI-204,219 as compared with placebo [18]. In contrast, neither ICI-204,219 nor another leukotriene-antagonist, MK-0679, by themselves, changed baseline prechallenge levels of urinary LTE4 [18, 34]. Furthermore, when RASMUSSEN et al. [25] gave the same dose of allergen in the presence of placebo or the leukotriene-antagonist, MK-571, the postchallenge increase in urinary LTE4 was the same at both sessions [25]. On the other hand, using cumulative challenge, we recently observed that leukotriene-antagonism increased postchallenge urinary LTE4 and the PD20FEV1 for aspirin in aspirin-sensitive asthmatics [34]. Considered together, the information available, therefore, supports the notion that the extent of urinary excretion of LTE4 is directly dependent upon the intensity of the provocative stimulus.

The magnitude of the shift in allergen-dose after ICI-204,219 corresponded to dose increments between 2.5 and 5.5 times for the group as a whole, depending upon...
how it was calculated (PD₉₀FEV₁ or cumulated dose of inhaled antigen). When determined for the seven responders, the average effect of the antagonist was obviously larger. It is known that the dose of the antagonist used in this study should cause about a ten fold shift of the dose response relation for inhaled LTD₄ at 12 h [35], suggesting a higher degree of antagonism at 2 h. Regrettably, it was not possible to determine the influence of this particular dose of ICI-204,219 on the bronchoprovocation response to LTD₄ at 2 h in this study. However, an indirect attempt to estimate the extent of leukotriene-antagonism, we performed skin tests with LTD₄ (see below).

It is difficult to know the reason(s) for the variability between the subjects in the effect of the leukotriene-antagonist on the airway response. One possibility is that the 20 mg dose of ICI-204,219 was too low to provide significant antagonism of cysteinyl-leukotrienes in all subjects. However, in view of the high repeatability of the procedure employed for allergen-provocations, it is tempting to speculate that the variability in response to the active drug indicates subject heterogeneity with respect to allergen-induced airway obstruction is not conclusively established.

The study also evaluated the possibility that the extent of leukotriene-antagonism could be estimated by performing skin tests with LTD₄. This approach has been useful in studies of antihistamines [30]. It has been reported that the skin response to LTD₄ is exceptionally variable [40], but with the procedure we used, the responses to LTD₄ were as reproducible as those produced by histamine. For the group as a whole, ICI-204,219 caused a 2.5 fold shift in the dose response relation for the LTD₄-flare. There was a tendency that the subjects with the greatest antagonism of the skin response to LTD₄, were those where ICI-204,219 caused the largest displacement of the dose-response relation for allergen in the airways. Nevertheless, the degree of antagonism by ICI-204,219 of the skin response to LTD₄ was not impressive, and was much less than has been reported for its effect on the bronchoprovocation response to LTD₄ [29, 35]. In fact, a much less potent leukotriene-antagonist, LY-171,883, has been reported to cause a similar inhibition of the skin response to LTD₄ [20]. Whether the relative inefficiency of ICI-204,219 on the skin responses was due to differences in drug concentrations between the lung and the skin, or differences in the receptor populations between the two tissues, remains to be determined. In summary, skin testing with LTD₄ cannot be recommended as a means to estimate the degree of antagonism of LTD₄ in the airways by a systemically administered receptor-antagonist.

In conclusion, the results of the present study support further evaluation of leukotriene-antagonism as a new strategy for inhibition of airway obstruction and inflammation in asthma. Future studies are required to evaluate the importance of this leukotriene component relative to other mediators of airway anaphylaxis, but, as discussed above, the role of histamine may have been overestimated by the unselective actions of antihistamines.

Acknowledgements: The authors thank H. Blomqvist and E. Eliasson for dedicated technical assistance, and R. Stark, S. Binks and J. Whitney of ICI (Zeneca) Pharmaceuticals in Alderby, UK and K. Boo of ICI (Zeneca) Pharmaceuticals in Sweden for kind help with different practical matters relating to this study.

References


