Title: CARD15/NOD2 polymorphisms are associated with severe pulmonary sarcoidosis.

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Short title: CARD15 polymorphisms in sarcoidosis.
Abstract

Sarcoidosis and Crohn’s disease (CD) are heterogeneous systemic diseases characterized by granulomatous inflammation. Caspase recruitment domain (CARD) 15 is a major susceptibility gene for CD, and specifically for ileal and fibrostenotic subtypes. The C-C chemokine receptor 5 (CCR5) gene has been associated with both parenchymal pulmonary sarcoidosis and perianal CD. This study explored associations between CARD15 polymorphisms, CCR5 haplotype and distinct pulmonary sarcoidosis subtypes. 185 Caucasian sarcoidosis patients were genotyped for CARD15 and CCR5 polymorphisms. The genetic data were compared to 347 healthy controls, and were examined for associations with serial pulmonary function tests and chest radiographs.

CARD15 genotypes did not differ between the unselected sarcoidosis cohort and controls. However, patients carrying the functional 2104T (702W) polymorphism were more likely to have radiographic stage IV disease at 4 years follow-up. All patients possessing both CARD15 2104T and CCR5 HHC haplotype had stage IV disease at presentation. Carriage of 2104T was associated with worse FEV₁, while carriage of the CARD15 1761G (587R) polymorphism was associated with better lung function.

For the first time, an association between two CARD15 polymorphisms and specific sarcoidosis phenotypes has been demonstrated, as well as an additive effect of possessing CARD15 2104T and CCR5 HHC haplotype.

Key words: CARD15, CCR5, lung function tests, polymorphism, sarcoidosis
Introduction

Sarcoidosis and Crohn’s disease are idiopathic systemic granulomatous disorders of the lung and intestine respectively, with variable involvement of the skin, eyes and joints. There are associations between phenotypic variability and HLA class II alleles in both sarcoidosis (DQB1*0201 with Lofgren’s syndrome and erythema nodosum (1); DQB1*0602 with severe pulmonary disease (1;2)) and Crohn’s disease (DRB1*07 with ileal involvement (3); DRB1*0103 with extraintestinal manifestations (4)). Furthermore, the HHC haplotype of the C-C chemokine receptor 5 (CCR5) gene (which promotes T cell recruitment and activation) is a marker for parenchymal involvement (5) though not for sarcoidosis overall, while the 32 bp deletion (Δ32) polymorphism of CCR5 is associated with the subgroup of Crohn’s patients with perianal disease (6).

The caspase recruitment domain gene CARD15 (also known as NOD2) is a major susceptibility gene for Crohn’s disease (7;8). The CARD15 gene encodes an intracellular protein (CARD15) of the nucleotide oligomerization domain (NOD) family, involved in innate immunity through recognition of bacterial pathogen-associated molecular patterns (PAMPs). CARD15 is composed of a nucleotide binding domain (NBD) and ten leucine rich repeats (LRRs) (7;9), which recognize muramyl dipeptide (MDP), a component of bacterial cell wall peptidoglycan. The three major Crohn’s disease-associated variants, all located in this region, result in either amino acid substitutions (C2104T [R702W] and G2722C [G908R]) or premature truncation of the protein (3020 insertion C [1007fs]). In European and North American cohorts, the risk of disease increases up to threefold with carriage of a single variant allele, and 20-fold with possession of two variant alleles (10).
other CARD15 single nucleotide polymorphisms (SNPs) including T1761G (R587R) and C802T (P268S) are also associated with Crohn’s disease, but in such cohorts are in linkage disequilibrium with the three previously mentioned variants (11). Notably, CARD15 is associated with specific Crohn’s subtypes: ileal and fibrostenotic disease.

CARD15 is the disease gene for Blau syndrome, a rare autosomal dominant granulomatous disorder characterized by arthritis, uveitis and a skin rash (12;13); early onset sarcoidosis has also been associated with CARD15 mutations (14). However, in these diseases the mutations are distinct from those of Crohn’s disease.

The similarities between Crohn’s disease and sarcoidosis, and the genotype-phenotype associations observed in the subtypes of the diseases which highlight their heterogeneity, justify the investigation of CARD15 in sarcoidosis. Previous studies of CARD15 in unselected sarcoidosis cohorts have provided conflicting results, with no association found in three studies (15-17) but an association with the G2722C (G908R) polymorphism (18) in one report. However, specific subgroups of sarcoidosis patients were not investigated in any of these cohorts.

Both CARD15 and CCR5 genes are of importance in innate immunity, and have been associated with specific subtypes of CD and sarcoidosis. Therefore, carefully phenotyped patients with sarcoidosis were genotyped for i. five Crohn’s disease-associated CARD15 polymorphisms and ii. the CCR5 HHC haplotype, to test the hypothesis that polymorphisms in these genes would be associated with specific sarcoidosis phenotypes, and to investigate potential gene-gene interactions.
Methods

Sarcoidosis patients

A total of 185 Caucasian patients were included in this study. All were recruited from the sarcoidosis clinic of the Royal Brompton Hospital, London, a tertiary referral centre with patients mainly from the South East of England. In all patients sarcoidosis was diagnosed histologically, and according to the criteria defined in the ATS/ERS/WASOG consensus statement on sarcoidosis (19).

Written patient consent was obtained from all subjects. The Ethics Committees of the Royal Brompton Hospital gave authorisation for the study.

Controls

The control population comprised 347 white Caucasian subjects from South East England, healthy as judged by checks including medical history, physical examination, and routine laboratory blood testing at regular intervals during a 10-year period before blood was taken for DNA extraction.

Genotyping for \textit{CARD15} and \textit{CCR5} polymorphisms

Single nucleotide polymorphisms (SNPs) were determined using sequence-specific primers (SSPs) and polymerase chain reaction (PCR) (20;21). Five \textit{CARD15} SNPs were studied in all 185 patients (nomenclature according to (11)): 802 C>T (P268S, \textit{exon}4, rs2066842), 1761 T>G (R587R, \textit{exon} 4, rs1861759), 2104C>T (R702W, \textit{exon} 4, rs2066844), 2722G>C (G908R, \textit{exon}8, rs2066845) and 3020insC (1007fs, \textit{exon} 11, rs2066847).
Genotyping for 8 polymorphisms of the *CCR5* gene (−5563(A/G), −3900(C/A), −3458(T/G), −2459(G/A), −2135(T/C), -2086(A/G), -1835(C/T) and Δ32) was also undertaken as previously described and haplotypes assigned, to identify those possessing the HHC haplotype (-2459G/ -2135T/ -2086G / -1835C) (5). 104 of the subjects had been included in the previous study (5) and 56 additional cases were genotyped.

**Radiography**

Chest radiographs for each patient were examined and compared to determine disease severity and course; these were evaluated independently by two experienced pulmonary radiologists (5) at presentation, 2 and 4 years follow-up. Chest radiographic data were available for 177 of the 185 patients at presentation to the Royal Brompton Hospital, 158 at 2 years and 126 at 4 years. These differences were due to the inclusion of a minority of patients with more recent diagnoses who have not been followed up for long enough to be included in the 2 and 4 year analysis. Chest radiograph staging was classified according to the joint ATS/ERS/WASOG consensus statement on sarcoidosis (19).

**Table 1** shows the chest radiograph staging for patients genotyped for *CARD15* polymorphisms only, and for patients genotyped for both *CARD15* and *CCR5* polymorphisms, respectively.

**Pulmonary function testing**

Pulmonary function tests included forced expiratory volume in 1 second (FEV₁) and forced expiratory vital capacity (FVC) assessed by spirometry and carbon monoxide
transfer capacity ($T_{LCO}$) as measured by the single breath technique. Both were expressed as percentage of predicted. These data were available for 174 of the 185 subjects at presentation, 149 at 2 years and 136 at 4 years.

**Data analysis**

Genotype and allele frequencies were determined by direct counting. Haplotypes were deduced by PHASE, version 2, a statistical haplotype reconstruction method (22).

Statistical analyses were performed using the program SPSS version 14 (SPSS Inc., Chicago, Illinois). Categorical data were analysed by chi-square contingency tables or Fisher’s exact test as appropriate. Continuous data were analysed using the Mann-Whitney U test or Kruskal-Wallis test as appropriate. A value of $p<0.05$ was considered significant.
Results

CARD15 polymorphisms: unselected sarcoidosis cohort

In order to determine if there were any associations between sarcoidosis in general and CARD15 polymorphisms or haplotypes, allele and haplotype frequencies were compared for all sarcoidosis patients and the control population. These comparisons are shown in Tables 2 and 3. Table 2 summarises the allele frequencies of the five CARD15 polymorphisms investigated in the sarcoid and control populations. Both populations were in Hardy-Weinberg equilibrium for all genotype frequencies. No significant differences were found between the unselected sarcoidosis cohort and controls. Table 3 shows the CARD15 haplotypes and their frequencies: there were no significant differences in CARD15 haplotypes between the two groups.

Of the functional, CD-associated polymorphisms, 2104T occurred only on haplotype 4, 3020insC on haplotype 5 and 2722C on haplotype 6. 1761G occurred only on haplotype 1, while 802T occurred on haplotypes 3-6 (Table 3).

CARD15, CCR5 and radiographic stage IV disease in sarcoidosis

Associations between CARD15 polymorphisms, the CCR5 HHC haplotype and chest radiographic staging were investigated. Carriage of the CARD15 2104T polymorphism (haplotype 4) was associated with a higher frequency of radiographic stage IV disease compared to non-carriers at 4 years of follow up (76.9% vs. 45.1%, p=0.04, OR=4.1, 95%CI=1.0-15.5). There were no significant differences between carriers and non-carriers of the other CARD15 polymorphisms or haplotypes and either parenchymal disease (stage II-IV) or stage IV disease at presentation, 2 years or 4 years.
We have previously shown an association between carriage of the CCR5 HHC haplotype and parenchymal disease at presentation (radiographic stage ≥ II vs. stages 0 and I) (5). In the current, larger, cohort (56 additional cases) the HHC haplotype was significantly increased in patients with stage IV compared stage 0-III disease at presentation (n=160, 72.1% vs. 46.5%, p=0.003, OR=3.0, 95%CI=1.5-5.9), 2 years (n=144, 68.3% vs. 45.7%, p=0.01, OR=2.6, 95%CI=1.3-5.1 ) and 4 years (n=115, 69.1% vs. 46.7%, p=0.02, OR=2.6, 95%CI=1.2-5.5 ).

Having established the influence of the CCR5 HHC haplotype on radiographic stage IV disease, and having seen for the first time a significant association between one of the functional Crohn’s disease-associated CARD15 SNPs (2104T, haplotype 4) and stage IV disease, we investigated the influence of 2104T in sarcoidosis patients subtyped according to their CCR5 HHC haplotype status. Patients were divided into four groups: i. Group 1 (n=7): carriage of both CCR5 HHC and CARD15 2104T ii. Group 2 (n=83): carriage of only CCR5 HHC iii. Group 3 (n=8): carriage of only CARD15 2104T iv. Group 4 (n=62): carriage of neither.

All seven of the subjects carrying the HHC haplotype and CARD15 2104T (haplotype 4) had radiographic stage IV disease at presentation, 2 years and 4 years (Figure 1).

No differences were found when the subgroups were similarly analysed for change in chest radiograph status (at 4 years follow up vs. at presentation).

The CARD15 802T polymorphism in combination with the HHC haplotype was also significantly associated with radiographic stage IV disease (p=0.002 at presentation, 2 years and 4 years) (Figure 2, groups as described above but for 802T carriage rather than 2104T). Further analysis revealed, however, that this was a consequence of the known linkage disequilibrium (11) between 802T and 2104T (Table 3).
No significant associations were found for 2722C, 3020insC or 1761G, or their respective haplotypes.

**CARD15 polymorphisms and pulmonary function tests**

To clarify whether the carriage of CARD15 polymorphisms and/or haplotypes acts as an index for disease severity, the possession of CARD15 polymorphisms (and hence haplotypes, Table 3) was investigated in relation to lung function tests.

Table 4 shows the carriage of the 802T, 1761G (haplotype 1) and 2104T (haplotype 4) alleles and lung function tests at presentation, 2 years and 4 years. Carriage of the CARD15 1761G allele (haplotype 1) was associated with better lung function, as defined by TLco, compared to non-carriage of the G allele at presentation (p=0.001), 2 years (p=0.006) and 4 years (p=0.002). The differences were significant at all time points, and interestingly a clear gene-dose effect can be seen at presentation: GG homozygotes have better TLco than TG heterozygotes, while TT homozygotes have the worst TLco (Figure 3).

Figure 4 shows the mean comparisons of TLco with CARD15 176I genotypes at presentation, 2 years and 4 years. Carriers of the 1761G allele had a higher FEV1 (p=0.02) at presentation compared to non-carriers. These patients also had a higher FVC at presentation (p=0.01); there was a trend towards significance for FVC at 2 years (p=0.07); and at 4 years the difference was again significant (p=0.02) (Table 4). Of note, this cohort with better lung function did not possess haplotype 4 which includes the 2104T allele.

Consistent with the worse radiographic stage associations, patients carrying the 2104T allele (haplotype 4) had a worse FEV1 at presentation (median; 79.3% vs. 92.3%,
p=0.04), 2 years (median; 73.8% vs. 93.3%, p=0.02) and 4 years (median; 77.6% vs. 92.4%, p=0.05), and a worse FVC at 4 years follow-up (median; 89.3% vs. 100.3%, p=0.04), compared to patients not carrying the 2104T allele.

No significant differences were seen between the carriage of 802T, 3020insC or 2722C, and pulmonary function tests.
DISCUSSION

Sarcoidosis and Crohn’s disease are both characterised by inflammation, granuloma formation and their systemic nature, with frequent extrapulmonary and extraintestinal manifestations respectively. They both display considerable phenotypic heterogeneity and degrees of disease severity, and important genetic associations have been identified for both conditions. The similarities between sarcoidosis and Crohn’s disease have prompted several studies of the Crohn’s susceptibility gene $CARD15/NOD2$ in unselected sarcoidosis cohorts (15-18). Of these, one has reported an association with the $2722C \ (G908R)$ variant (18), and one (17) found an increased transmission of two of the alleles but concluded that these mutations played no major role in their study population.

The $CARD15$ polymorphisms studied are specifically associated with Crohn’s disease subtypes, and it was hypothesised that they may be associated with subtypes of sarcoidosis. The $CCR5$ gene has also been associated with both diseases (5;6), and this gene was further investigated to allow subclassification of the cohort on the basis of their $CCR5$ haplotype.

Significant associations have been demonstrated between both a functional $CARD15$ SNP ($2104T$, haplotype 4) and a synonymous $CARD15$ SNP ($1761G$, haplotype 1), and severe pulmonary sarcoidosis phenotypes. An association between the $CCR5$ $HHIC$ haplotype and chest radiographic stage IV disease has been established. Of note, all patients possessing both the $CCR5$ $HHIC$ haplotype and $CARD15 \ 2104T$ allele had radiographic stage IV disease at presentation.

Patients possessing the functional $2104T$ allele (haplotype 4) had worse FEV$_1$ values at presentation, 2 years and 4 years, as well as worse FVC at 4 years. The carriage of
this polymorphism was also associated with an increased risk of stage IV disease at 4 years.

The \textit{CARD15} gene product plays an important role in innate immunity by influencing recognition of specific bacterial patterns, and this is impaired in those possessing the CD-associated functional variants. The precise mechanism through which these variants result in increased disease susceptibility remains uncertain. Transient transfection experiments have suggested that the impaired binding of MDP decreases NF\(\kappa\)B activation (23), with consequent impaired microbial clearance. In \textit{CARD15} mutant mice, however, increased downstream NF\(\kappa\)B signalling has been demonstrated (24) with subsequent inflammation. The association of one of the functional mutations with a severe pulmonary sarcoidosis phenotype offers an intriguing insight into possible dysregulated responses to bacteria in this cohort.

We have previously found associations between the \textit{CCR5 HHC} haplotype and parenchymal lung disease in a cohort of 104 British patients (5). In this larger cohort, we have elucidated a significant association with pulmonary radiographic stage IV disease. These findings are consistent with our previous results, suggesting that dysfunction of this gene, of major importance in T cell activation, could result in abnormal trafficking of T cells to the lung and hence parenchymal abnormalities culminating in fibrosis (5).

All patients carrying the \textit{CCR5 HHC} haplotype and the \textit{CARD15 2104T} polymorphism had pulmonary stage IV disease at presentation. It is of interest that these two genes are both involved in innate immunity as discussed above. \textit{CARD15}-deficient mice have been shown to lack the normal inhibition of the Toll Like Receptor 2-mediated NF\(\kappa\)B response, with consequent \(T_{H1}\) overactivation (25). \textit{CCR5}}
also enhances T cell activation (26). It is hypothesised that the possession of $CARD15$ $2104T$ contributes to an impaired response to bacteria, whether commensal or pathogenic, with subsequent abnormal T cell activation which is further enhanced by the possession of $CCR5$ $HHC$ in this cohort of patients.

The $CARD15$ $1761G$ allele, present in 40% of the sarcoidosis cohort, was associated with significantly better lung function parameters (not only TLco but also FVC) than the wild-type allele at all time-points. This suggests that carriage of this polymorphism is associated with less diffuse lung disease. A significant gene-dose effect was also observed (Figure 3). In Crohn’s disease this polymorphism is protective: there is a lower frequency of Crohn’s disease in those possessing the $CARD15$ $1761G$ allele (11). This may suggest that the polymorphism confers a favourable phenotype. It is, however, a synonymous SNP.

Possible explanations for the association of this synonymous SNP with a specific disease phenotype may include linkage disequilibrium with another, functional SNP as yet unidentified. This hypothesis would suggest that it is acting as a marker. Alternatively, there has been considerable interest in ‘silent’ polymorphisms, and a functional study has shown that the product of a synonymous polymorphism can differ significantly from the wildtype protein (27). It was hypothesised that the synonymous polymorphism may affect the timing of cotranslational folding and so alter the structure of binding sites. Further functional studies of the $1761G$ SNP will be required to investigate this.

In this study, chest radiographs were evaluated independently by two experienced pulmonary radiologists and chest radiographic appearances remain the accepted means of staging sarcoidosis (19). However, pulmonary computed tomography (CT) scoring may be a more sensitive means of assessing lung disease pattern including
fibrosis and disease severity (28), and further studies investigating carriage of these genetic polymorphisms in relation to CT score may serve to confirm the association between lung fibrosis and these genes. Indeed, the relative insensitivity of chest radiographs may explain the lack of association found for CARD15 1761G and radiographic stage, when significant associations with lung function were found. Quantification of lung function change with continuous variables may be more precise than the categorical, descriptive chest radiographic staging system, and thus more likely to be able to identify associations (29).

A recent study using blood mononuclear cells has shown that a combination of NOD2 and Toll-like receptor (TLR)-2 ligand stimulation induced a higher secretion of the pro-inflammatory cytokines TNF-α and IL-1β in sarcoidosis patients compared to healthy controls (30). These cytokines are important in determining the inflammatory response in active sarcoidosis, and indeed granuloma formation. It will be informative to perform further such studies in individuals sub-classified according to their CARD15 genotype. Specifically, it would be of interest to investigate whether those possessing the 1761G allele have a reduced predisposition to TNF-α and IL-1β secretion compared to those with the wild-type allele, and whether those with the 2104T polymorphism have significantly greater secretion of these cytokines.

In conclusion, neither the CARD15 nor CCR5 variants were more prevalent overall in the sarcoidosis cohort than the controls, confirming the hypothesis that these are not disease susceptibility genes but rather are associated with disease modification and/or progression after sarcoidosis is established. These results provide further evidence for genetic heterogeneity in determining the phenotype in sarcoidosis.
Acknowledgements

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Conflict of Interest

The authors declare that they have no conflicts of interest.
References


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Figure legends

Figure 1. Frequencies of chest radiograph stage IV disease related to *CCR5 HHC* and *CARD15 2104T* (haplotype 4) status. Group 1 (n=7 at presentation and 2 years and n=6 at 4 years): carriage of both *CCR5 HHC* and *CARD15 2104T*. Group 2 (n=83 at presentation, n=73 at 2 years and n=60 at 4 years): carriage of only *CCR5 HHC*. Group 3 (n=8 at presentation and 2 years and n=6 at 4 years): carriage of only *CARD15 2104T*. Group 4 (n=62 at presentation, n=56 at 2 years and n=43 at 4 years): carriage of neither.

All of the subjects carrying *CCR5 HHC* and *CARD15 2104T* had stage IV disease at presentation (p=0.0003, Kruskal-Wallis test), and so at 2 years and 4 years.

![Figure 1](image)

Figure 2. Frequencies of chest radiograph stage IV disease related to *CCR5 HHC* haplotype and *CARD15 802T* (haplotypes 3-6) status. Group 1 (n=28 at
presentation, n=27 at 2 years and n= 25 at 4 years): carriage of both CCR5 HHC and CARD15 802T. Group 2 (n=62 at presentation, n=53 at 2 years and n=41 at 4 years): carriage of only CCR5 HHC. Group 3 (n=31 at presentation, n=30 at 2 years and n=24 at 4 years): carriage of only CARD15 802T. Group 4 (n=39 at presentation, n=34 at 2 years and n=25 at 4 years): carriage of neither.

The CARD15 802T polymorphism in combination with the CCR5 HHC haplotype was also significantly associated with stage IV disease at presentation (p<0.005), 2 years (p<0.005), and 4 years (p<0.005), Kruskal-Wallis test.

Figure 3. Influence of CARD15 1761 genotype on TLCO. There was a significant gene-dose effect between CARD15 1761 genotype (GG > GT > TT) and the level of TLCO at presentation (p=0.001). Median and interquartile ranges are shown.
Figure 4. Mean (± Standard Error) comparisons of TL\textsubscript{CO} with \textit{CARD15 1761} genotypes at presentation, 2 years and 4 years. Individuals with the GG genotype (>TG>TT) had significantly higher mean TL\textsubscript{CO} levels at presentation (p=0.001), 2 years (p=0.009) and 4 years (p=0.003).
Figure 4

![Graph showing changes in mean TLco over years with different genotypes (TT, TG, and GG).]
Table 1. Chest radiograph staging of sarcoidosis patients genotyped for the *CARD15*, and both *CARD15* and *CCR5* SNPs, at presentation, 2 years and 4 years

**For CARD15**

<table>
<thead>
<tr>
<th>Stage</th>
<th>At presentation (N=177)</th>
<th>At 2 years (N=158)</th>
<th>At 4 years (N=126)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>22 12.4%</td>
<td>33 20.9%</td>
<td>37 29.4%</td>
</tr>
<tr>
<td>I</td>
<td>33 18.6%</td>
<td>24 15.2%</td>
<td>12 9.5%</td>
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<td>II</td>
<td>39 22.0%</td>
<td>21 13.3%</td>
<td>10 7.9%</td>
</tr>
<tr>
<td>III</td>
<td>16 9.0%</td>
<td>11 7.0%</td>
<td>6 4.8%</td>
</tr>
<tr>
<td>IV</td>
<td>67 37.9%</td>
<td>69 43.7%</td>
<td>61 48.4%</td>
</tr>
</tbody>
</table>

**For CARD15 and CCR5**

<table>
<thead>
<tr>
<th>Stage</th>
<th>At presentation (N=160)</th>
<th>At 2 years (N=144)</th>
<th>At 4 years (N=115)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>21 13.1%</td>
<td>31 21.5%</td>
<td>35 30.4%</td>
</tr>
<tr>
<td>I</td>
<td>29 18.1%</td>
<td>22 15.3%</td>
<td>9 7.8%</td>
</tr>
<tr>
<td>II</td>
<td>35 21.9%</td>
<td>17 11.8%</td>
<td>10 8.7%</td>
</tr>
<tr>
<td>III</td>
<td>14 8.8%</td>
<td>11 7.6%</td>
<td>6 5.2%</td>
</tr>
<tr>
<td>IV</td>
<td>61 38.1%</td>
<td>63 43.8%</td>
<td>55 47.8%</td>
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Table 2. *CARD15* allele frequencies in sarcoidosis and controls

<table>
<thead>
<tr>
<th>Polymorphism</th>
<th>Amino Acid</th>
<th>Sarcoidosis</th>
<th>Controls</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>( n = 185 )</td>
<td>%</td>
<td>( n = 347 )</td>
</tr>
<tr>
<td><em>802</em> ( (rs2066842) )</td>
<td>( C ) Pro</td>
<td>289</td>
<td>78.1</td>
</tr>
<tr>
<td></td>
<td>( T ) Ser</td>
<td>81</td>
<td>21.9</td>
</tr>
<tr>
<td><em>1761</em> ( (rs1861759) )</td>
<td>( T ) Arg</td>
<td>222</td>
<td>60.0</td>
</tr>
<tr>
<td></td>
<td>( G ) Arg</td>
<td>148</td>
<td>40.0</td>
</tr>
<tr>
<td><em>2104</em> ( (rs2066844) )</td>
<td>( C ) Arg</td>
<td>351</td>
<td>94.9</td>
</tr>
<tr>
<td></td>
<td>( T ) Trp</td>
<td>19</td>
<td>5.1</td>
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<tr>
<td><em>2722</em> ( (rs2066845) )</td>
<td>( G ) Gly</td>
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<td></td>
<td>( C ) Arg</td>
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<td>0.5</td>
</tr>
<tr>
<td><em>3020</em> ( (rs2066847) )</td>
<td>( W )</td>
<td>367</td>
<td>99.2</td>
</tr>
<tr>
<td></td>
<td>( ins C ) Fs</td>
<td>3</td>
<td>0.8</td>
</tr>
</tbody>
</table>

*ins C*: insertion C

\( W \): wild-type

Fs: frameshift
**Table 3.** *CARD15* haplotypes and their carriage frequencies in sarcoidosis and controls

<table>
<thead>
<tr>
<th></th>
<th>802</th>
<th>1761</th>
<th>2104</th>
<th>2722</th>
<th>3020</th>
<th>Sarcoidosis</th>
<th>Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haplotype 1</td>
<td>C</td>
<td>G</td>
<td>C</td>
<td>G</td>
<td>W</td>
<td>123 (n = 185) 66.5</td>
<td>212 (n = 347) 61.1</td>
</tr>
<tr>
<td>Haplotype 2</td>
<td>C</td>
<td>T</td>
<td>C</td>
<td>G</td>
<td>W</td>
<td>116 (n = 185) 62.7</td>
<td>205 (n = 347) 59.1</td>
</tr>
<tr>
<td>Haplotype 3</td>
<td>T</td>
<td>T</td>
<td>C</td>
<td>G</td>
<td>W</td>
<td>52 (n = 185) 28.1</td>
<td>122 (n = 347) 35.2</td>
</tr>
<tr>
<td>Haplotype 4</td>
<td>T</td>
<td>T</td>
<td>T</td>
<td>G</td>
<td>W</td>
<td>17 (n = 185) 9.2</td>
<td>35 (n = 347) 10.1</td>
</tr>
<tr>
<td>Haplotype 5</td>
<td>T</td>
<td>T</td>
<td>C</td>
<td>G</td>
<td>ins C</td>
<td>3 (n = 185) 1.6</td>
<td>11 (n = 347) 3.2</td>
</tr>
<tr>
<td>Haplotype 6</td>
<td>T</td>
<td>T</td>
<td>C</td>
<td>C</td>
<td>W</td>
<td>2 (n = 185) 1.1</td>
<td>10 (n = 347) 2.9</td>
</tr>
</tbody>
</table>

**Bold:** variant allele.

W: wild-type

ins C: insertion C
Table 4. Carriage of *CARD15* 802T, 1761G (haplotype 1) and 2104T (haplotype 4) alleles in relation to lung function tests at presentation, 2 years and 4 years (median values shown, % predicted).

<table>
<thead>
<tr>
<th></th>
<th>802T</th>
<th>1761G (haplotype 1)</th>
<th>2104T (haplotype 4)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>FEV₁</td>
<td>FVC</td>
</tr>
<tr>
<td><strong>Presentation</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carrier</td>
<td>64</td>
<td>93.05</td>
<td>98.35</td>
</tr>
<tr>
<td>Non carrier</td>
<td>110</td>
<td>90.95</td>
<td>98</td>
</tr>
<tr>
<td><strong>p-value</strong></td>
<td></td>
<td>0.42</td>
<td>0.5</td>
</tr>
<tr>
<td><strong>2 years</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carrier</td>
<td>59</td>
<td>94.3</td>
<td>101.1</td>
</tr>
<tr>
<td>Non carrier</td>
<td>89</td>
<td>91.2</td>
<td>98.7</td>
</tr>
<tr>
<td><strong>p-value</strong></td>
<td></td>
<td>0.6</td>
<td>0.9</td>
</tr>
<tr>
<td><strong>4 years</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carrier</td>
<td>53</td>
<td>91.3</td>
<td>96.8</td>
</tr>
<tr>
<td>Non carrier</td>
<td>83</td>
<td>92.4</td>
<td>100.2</td>
</tr>
<tr>
<td><strong>p-value</strong></td>
<td></td>
<td>0.3</td>
<td>0.2</td>
</tr>
</tbody>
</table>

N=number; FEV₁=Forced expiratory volume in 1 second; FVC=Forced vital capacity; TL<sub>CO</sub>=Carbon monoxide transfer capacity.

Total numbers: 174 at presentation, 148 at 2 years and 136 at 4 years.