Effects of cigarette smoke condensate on pneumococcal biofilm formation and pneumolysin

N.D. Mutepe¹, R. Cockeran¹*, H.C. Steel¹, A.J. Theron¹, T.J. Mitchell², C. Feldman³, R. Anderson¹.

¹Medical Research Council (MRC) Unit for Inflammation and Immunity, Department of Immunology, Faculty of Health Sciences, University of Pretoria, and Tshwane Academic Division of the National Health Laboratory Service, South Africa; ²Institute of Infection, Immunity and Inflammation, Sir Graeme Davies Building, University of Glasgow, UK; ³Division of Pulmonology, Department of Internal Medicine, Charlotte Maxeke Johannesburg Academic Hospital and Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa.

Correspondence: Dr R. Cockeran
Department of Immunology
Private Bag x323
Pretoria 0001
South Africa
Telephone: +27-12-319-2624
Telefax: +27-12-323-0732
E-mail: riana.cockeran@up.ac.za

ERJ Express. Published on June 27, 2012 as doi: 10.1183/09031936.00213211

Copyright 2012 by the European Respiratory Society.
Abstract:

**Introduction and Aim:** Although the well-recognised predisposition of cigarette smokers for development of severe pneumococcal disease may be attributable to impairment of local host defences, less is known about the direct effects of smoke exposure on airway pathogens, or their virulence factors. In the current study, we have investigated the effects of cigarette smoke condensate (CSC) on biofilm formation by *Streptococcus pneumoniae*, as well as on the pore-forming activity of its major toxin, pneumolysin.

**Materials and Methods:** Biofilm formation following exposure of the pneumococcus to CSC (20-160 µg/ml) was measured using a crystal violet-based spectrophotometric procedure, while the pore-forming activity of recombinant pneumolysin was determined by a fura-2/AM-based spectrofluorimetric procedure to monitor the uptake of extracellular Ca²⁺ by isolated human neutrophils.

**Results:** Exposure of the pneumococcus or pneumolysin to CSC resulted in significant dose-related augmentation of biofilm formation (*P*≤0.05 at 80 and 160 µg/ml) and substantial attenuation of the pore-forming interactions of pneumolysin, respectively.

**Conclusion:** Augmentation of biofilm formation and inactivation of pneumolysin as a consequence of smoking, are likely to favour microbial colonization and persistence, both being essential precursors of pneumococcal disease.

Keywords: cigarette smoking; pneumococcus; severe pneumococcal disease
**Introduction:**

*Streptococcus pneumoniae* is a major cause of morbidity and mortality worldwide. It commonly causes otitis media, meningitis and pneumonia, with the young and the elderly being particularly vulnerable, while in those infected with the human immunodeficiency virus (HIV), all age groups are affected. Notwithstanding the threats posed by emerging antibiotic resistance, a significant percentage of patients with severe pneumococcal disease who receive appropriate antimicrobial chemotherapy, still die [1]. Importantly, cigarette smoking is the strongest independent risk factor for invasive pneumococcal pneumonia, affecting the immunocompetent non-elderly, and is responsible for one-half of the disease burden, apparently by promoting colonization, a prerequisite for invasive disease [2].

Although the exact causes of the predisposing effect of the smoking habit on development of severe pneumococcal disease are incompletely understood, smoke-mediated impairment of airway host defenses has been implicated. However, less is known about the direct effects of cigarette smoke on the pneumococcus. In the current study, we have investigated the effects of exposure to cigarette smoke condensate on biofilm formation by the pneumococcus, as well as on the bioactivity of its pore-forming, pro-inflammatory toxin, pneumolysin. Biofilm is a self-generated polymer matrix which insulates the pneumococcus, and other microbial pathogens from host defenses and antibiotics, promoting bacterial persistence [3]. Pneumolysin, on the other hand, considered by many to be one of the most important virulence factors of the pneumococcus, initiates, via its interactions with airway epithelial cells, an early inflammatory response primarily involving neutrophils, which controls colonization [4].

**Materials and Methods:**

**Bacterial strains:**

An antibiotic-sensitive, clinical isolate of *S. pneumoniae*, strain 172 (serotype 23F, MLST 81), was kindly provided by the National Institute of Communicable Diseases (NICD), Johannesburg, South Africa. The strain was grown overnight to the mid-log phase, at 37°C in an atmosphere of 5% CO₂ in tryptone soy broth (TSB, Merck, Darmstadt, Germany) followed by turbidometric numerical standardisation of the culture, representing 6.14 x 10⁶ colony forming units (cfu)/ml,
before treatment with or without cigarette smoke condensate (CSC). A limited number of experiments was also performed with strain 3328 (serotype 14, MLST: ST230), an \textit{erm}(B)-expressing macrolide-resistant strain of the pneumococcus.

\textit{Chemicals and reagents}

All chemicals and reagents, unless otherwise indicated, were obtained from Sigma Chemical Co (St Louis, MO, USA).

\textit{Cigarette Smoke Condensate (CSC)}

CSC was purchased from Murty Pharmaceuticals (Lexington, KY, USA) and used at final concentrations of 20, 40, 80, and 160 µg/ml. Solvent controls (DMSO) were included in all experiments. The total amount of condensate generated during the combustion of one cigarette is 26.3 mg [5]; therefore the concentrations of CSC used in this study are relevant in the context of the smoking habit.

\textit{Effects of CSC on the growth of S. pneumoniae}

\textit{S. pneumoniae} was exposed to CSC (20-160 µg/ml) or the solvent control for 16h at 37°C, in an atmosphere of 5% CO$_2$. The growth of the planktonic bacteria was determined by standard colony forming unit (cfu) enumeration procedures.

\textit{Effects of CSC on biofilm formation by S. pneumoniae}

The bacteria in growth medium were exposed to CSC (20-160 µg/ml) or the solvent control for 16h at 37°C, 5% CO$_2$, in a 6-well tissue culture plate to facilitate adherence and biofilm formation. Following incubation, the non-adherent bacteria and media were removed, and the wells washed three times with phosphate-buffered saline (PBS 0.15M, Beckton Dickinson and Company, Sparks, MD, USA). Following the removal of all the unbound bacteria, the biofilm was stained with 0.1% crystal violet, the excess dye was removed, and the wells washed five times with PBS. The crystal violet was released from the adherent bacteria by the addition of 96% ethanol and the amount of biofilm formed determined spectrophotometrically at a wavelength of 570nm using the PowerwaveX (Bio-Tec Instruments Inc., Winooski, Vermont, USA) plate reader.

\textit{Neutrophils}

The study was approved by the Research Ethics Committee, Faculty of Health Sciences, University of Pretoria (Protocol no S1/2011).
Purified neutrophils were prepared from heparinized (5 U/ml of preservative–free heparin) venous blood from healthy adult volunteers. These were routinely of high purity (>90%) and viability (>95%) [6].

Spectrofluorimetric measurement of cytosolic calcium (Ca\(^{2+}\))

For these experiments, recombinant pneumolysin (20 µg/ml) [6] was treated with either 20 or 40 µg/ml CSC or DMSO control for a period of 10 min at room temperature. The effect of CSC on the pore-forming activity of pneumolysin was then assessed according to the magnitude of uptake of extracellular Ca\(^{2+}\) by neutrophils, using the fluorescent Ca\(^{2+}\)–sensitive dye fura-2/AM [6]. Neutrophils (1 x 10\(^7\)/ml) were pre-loaded with fura-2/AM (2 µM, final) for 30 min at 37°C in PBS, washed twice and resuspended in indicator-free Hanks Balanced Salt Solution (HBSS, Highveld Biological, Johannesburg, South Africa). The fura-2/AM-loaded neutrophils (2 x 10\(^6\)/ml) were preincubated for 10 min at 37°C after which they were transferred to disposable cuvettes, which were maintained at 37°C in a Hitachi 650 10S Fluorescence Spectrophotometer (Hitachi High-Technologies Corporation, Tokyo , Japan) with excitation and emission wavelengths set at 340 and 500nm, respectively. After a stable baseline was obtained (± 1 min), the neutrophils were exposed to 3µl of CSC-treated or -untreated (control) pneumolysin and alterations in cytosolic Ca\(^{2+}\) concentrations were monitored over a 7 min time course. The final volume in each cuvette was 3ml, containing a total of 6 x 10\(^6\) neutrophils, and the final concentrations of pneumolysin and CSC were 20 ng/ml and 20 or 40 ng/ml, respectively. At these concentrations (20 and 40 ng/ml), CSC has no effects on neutrophils.

Statistical analysis

Five and 6 separate experiments were performed for measurement of the effects of CSC on growth and biofilm formation (strain 172) and for pneumolysin bioactivity respectively, while 3 experiments were performed using strain 3328 of the pneumococcus. The results are expressed as median values with 95% confidence limits or as Fura-2/AM fluorescence traces for the biofilm and pneumolysin experiments respectively. Statistical significance was calculated by the Mann-Whitney \(U\) test (2 tailed). \(P\) values of \(< 0.05\) were considered significant.

Results:

Effects of CSC on the growth and viability of S. pneumoniae
The effects of CSC (20-160 µg/ml) on the growth of planktonic *S. pneumoniae* strain 172 are shown in Figure 1A. The condensate did not significantly affect the viability of strain 172, as was the case with strain 3328 (results not shown).

**Effects of CSC on biofilm formation by *S. pneumoniae***

The effects of CSC (20-160 µg/ml) on biofilm formation by strain 172 of *S. pneumoniae* are shown in Figure 1B. Exposure of the pneumococcus to CSC resulted in a dose-dependent increase in biofilm formation, which reached statistical significance at 80 and 160 µg/ml. In the case of strain 3328, exposure to CSC at 80 and 160 µg/ml caused a significant (*P* < 0.001) increase in biofilm formation, the median OD values with the upper 95% confidence limit for the control, 80 and 160 µg/ml CSC-treated systems being 0.074 ± 0.0778; 0.092 ± 0.0982; and 0.117 ± 0.1595 respectively.

**Effects of CSC on the bioactivity of pneumolysin***

The effects of exposure of neutrophils to untreated and CSC-treated pneumolysin on the pore-forming activity of the toxin measured according to the influx of extracellular Ca²⁺ are shown in Figure 2. Exposure of neutrophils to control pneumolysin at 20 ng/ml was followed by a lag phase of approximate duration 20sec and by an abrupt increase in fluorescence intensity thereafter due to the influx of extracellular Ca²⁺, which reached a plateau after approximately 3 min. Addition of CSC-treated pneumolysin to neutrophils was associated with a prolongation of the lag phase (1.5-2.5 min), followed by a significant (*P*<0.0028 – <0.0002) attenuation of influx of cytosolic Ca²⁺, compatible with loss of pore-forming activity.

**Discussion:**

The results of this study clearly demonstrate that exposure of the pneumococcus to CSC for 16 hours, at concentrations which are representative of the smoking habit, results not only in the augmentation of biofilm formation in the absence of detectable levels of growth, but also substantial inactivation of the pore-forming, pro-inflammatory activity of its major toxin, pneumolysin. CSC, possibly by acting as a stressor, may activate quorum sensing mechanisms, which in turn, initiate biofilm formation, enabling the pneumococcus to enter a quiescent, persistent phase from which it can re-emerge when host defences are transiently compromised [4]. We cannot exclude the possibility that prolonged exposure to CSC at higher concentrations than those used in the current study may affect the growth of the pneumococcus.
We do concede, however, that brief exposure of the pneumococcus to CSC *in vitro* is not entirely representative of exposure to inhaled cigarette smoke in the airways. Nonetheless, several other groups have recently reported on the effects of cigarette smoke exposure on biofilm formation by various microbial pathogens. Baboni *et al.* reported that exposure of both *Streptococcus mutans* and *Candida albicans* to CSC increased the adherence of these oral pathogens to orthodontic material in the setting of increased biofilm formation [7], while Bagaitkar *et al.* also observed increased biofilm formation following exposure of another oral pathogen, *Porphyromonas gingivalis* to CSC [8]. Even more recently, Goldstein-Daurach *et al.* reported that exposure of a range of bacterial pathogens, isolated from the sinonasal cavities of smokers, to smoke generated by combustion of 5 reference cigarettes resulted in activation of biofilm formation [9].

To our knowledge, the effects of cigarette smoke on the bioactivity of the pneumococcal toxin, pneumolysin, have not been described previously. When released at high concentrations during severe pneumococcal disease, pneumolysin, as a consequence of its cytolytic and pro-inflammatory activities, causes acute lung injury following extracellular dissemination of the pneumococcus [1]. However, during the early stages of colonization when numbers of the pneumococci are low, pneumolysin via its pore-forming interactions with airway epithelial cells promotes production of interleukin-8 and early influx of neutrophils which contribute to the early control of colonization [4,10]. In the current study, exposure to very low concentrations of CSC resulted in attenuation of the pore-forming activity of pneumolysin, probably by oxidative inactivation, which is likely to favour colonization.

In conclusion, these effects of cigarette smoke on both biofilm formation and the bioactivity of pneumolysin may promote both colonization and persistence, important precursors of invasive pneumococcal disease.

Acknowledgement:
CF and RC are supported by the National Research Foundation of South Africa.
References:


Legends to Figures:

Figure 1:
Effects of exposure of *S. pneumoniae* to cigarette smoke condensate (CSC) at 20, 40, 80 and 160 μg/ml on viability, measured as colony forming units (cfu)/ml (A), and biofilm formation, measured spectrophotometrically by crystal violet staining at a wavelength of 570nm (B). The results are presented as the median values ± 95% confidence limits of 5 different experiments, with 3 replicates for each system in each experiment. *P ≤ 0.05 when compared to the solvent control.*
Figure 2:
Fura-2/AM fluorescence responses of neutrophils following exposure to cigarette smoke condensate (CSC, final concentration of 20 or 40 ng/ml)-treated or untreated pneumolysin (at a fixed, final concentration of 20 ng/ml). Pneumolysin (with or without CSC pre-treatment) was added as indicated (↑) after a stable baseline was achieved (± 1 min). Data from 3 experiments, using cells from 3 different donors are shown. A total of six different experiments, using neutrophils from 6 different donors were performed, all showing comparable results. *$P<0.0028-<0.0002$ for comparison of the areas under the curve for the control system with those of the CSC-treated systems for all 6 experiments.