

## References

- 1 World Health Organization. Global tuberculosis report 2015. 20th edition. Geneva, WHO, 2015.
- 2 Olayemi A, Nicolas V, Hulsemans J, *et al.* Taxonomy of the African giant pouched rats (Nesomyidae: *Cricetomys*): molecular and craniometric evidence support an unexpected high species diversity. *Zool J Linnean Soc* 2012; 165: 700–719.
- 3 Poling A, Weetjens B, Cox C, *et al.* Tuberculosis detection by giant African pouched rats. *Behav Anal* 2011; 34: 47–54.
- 4 Steingart KR, Ng V, Henry M, *et al.* Sputum processing methods to improve the sensitivity of smear microscopy for tuberculosis: a systematic review. *Lancet Infect Dis* 2006; 6: 664–674.
- 5 Uddin MK, Chowdhury MR, Ahmed S, *et al.* Comparison of direct *versus* concentrated smear microscopy in detection of pulmonary tuberculosis. *BMC Res Notes* 2013; 6: 1–6.
- 6 Getahun H, Harrington M, O'Brien R, *et al.* Diagnosis of smear-negative pulmonary tuberculosis in people with HIV infection or AIDS in resource-constrained settings: informing urgent policy changes. *Lancet* 2007; 369: 2042–2049.
- 7 Cooper RG. Care, husbandry and diseases of the African giant rat (*Cricetomys gambianus*). *J S Afr Vet Assoc* 2008; 79: 62–66.
- 8 Weetjens BJ, Mgone GF, Machang'u RS, *et al.* African pouched rats for the detection of pulmonary tuberculosis in sputum samples. *Int J Tuberc Lung Dis* 2009; 13: 737–743.
- 9 Mahoney A, Weetjens BJ, Cox C, *et al.* Pouched rats' detection of tuberculosis in human sputum: comparison to culturing and polymerase chain reaction. *Tuberc Res Treat* 2012; 2012: 716989.
- 10 Reither K, Jugheli L, Glass TR, *et al.* Evaluation of giant African pouched rats for detection of pulmonary tuberculosis in patients from a high-endemic setting. *PloS One* 2015; 10: e0135877.
- 11 Flahault A, Cadilhac M, Thomas G. Sample size calculation should be performed for design accuracy in diagnostic test studies. *J Clin Epidemiol* 2005; 58: 859–862.
- 12 Mahoney A, Edwards TL, Weetjens BJ, *et al.* Giant African pouched rats (*Cricetomys gambianus*) as detectors of tuberculosis in human sputum: two operational improvements. *Psychol Rec* 2013; 63: 583–593.
- 13 Lönnroth K, Migliori GB, Abubakar I, *et al.* Towards tuberculosis elimination: an action framework for low-incidence countries. *Eur Respir J* 2015; 45: 928–952.
- 14 Stuckler D, Basu S, McKee M, *et al.* Mining and risk of tuberculosis in sub-Saharan Africa. *Am J Public Health* 2011; 101: 524–530.
- 15 Kik SV, Denkinger CM, Casenghi M, *et al.* Tuberculosis diagnostics: which target product profiles should be prioritised? *Eur Respir J* 2014; 44: 537–540.

Eur Respir J 2016; 48: 579–582 | DOI: 10.1183/13993003.00264-2016 | Copyright ©ERS 2016

# Is bedaquiline as effective as fluoroquinolones in the treatment of multidrug-resistant tuberculosis?



To the Editor:

Bedaquiline (Bdq) is approved for the treatment of multidrug-resistant (MDR) tuberculosis (TB). In a phase IIb trial, Bdq allowed a significant reduction in time to culture conversion and improved outcome in MDR-TB patients [1, 2]. Preliminary reports of Bdq compassionate use have shown promising results [3–5]. However, in an early bactericidal activity (EBA) study, the association of moxifloxacin (Mfx) with PA-824 and pyrazinamide showed better activity than Bdq-based associations [6]. In addition, resistance to fluoroquinolones (Fq) has been associated with poorer outcome in MDR-TB before Bdq use [7]. These data reinforce the pivotal role of Fq. Comparing Bdq to Fq in interventional studies is challenging. Indeed, the paucity of drugs available for MDR-TB treatment, and the need for combination therapy, often impose the need to use all available drugs.

In an interim analysis of a Bdq-treated MDR-TB cohort, we showed that culture conversion reached 96% with 6-month Bdq-containing treatment regimens [8]. Following these encouraging results, we sought to compare the microbiological efficacy of Bdq- and Fq-containing regimens.

A retrospective study comparing microbiological outcome in Bdq-treated and Fq-treated patients was designed using the 2006–2014 cohort of MDR-TB patients hospitalised at Bligny's Sanatorium (Briis-sous-Forges), a French referral TB centre. The first group included patients treated for  $\geq 30$  days with Bdq, who either had not received any Fq or had received Fq but harboured *Mycobacterium tuberculosis* isolates with high-level phenotypical Fq resistance. The second group comprised patients treated for  $\geq 30$  days with any Fq but without Bdq, with isolates susceptible to ofloxacin (Ofx) and Mfx. All patients received any second-line injectable drug and linezolid for  $\geq 30$  days, and had positive sputum cultures at treatment start.

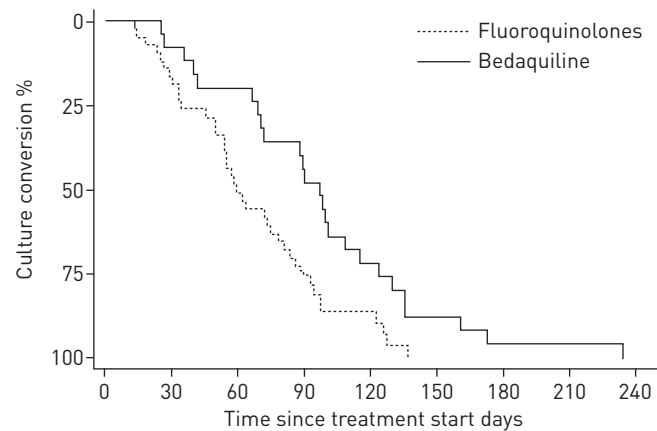


FIGURE 1 Kaplan-Meier curves of sputum time to culture conversion in the bedaquiline- and fluoroquinolone-treated patients.

All regimens were individually tailored according to the drug susceptibility test (DST) results and were started at the hospital where the diagnosis was made or at the Bligny Sanatorium. All drugs were administered under direct observation and according to international guidelines [9]. Sputum cultures were repeated every 2 weeks up to culture conversion, and monthly thereafter. Time to culture conversion was measured from treatment start to the first of two consecutive negative culture results.

The DST was performed at the French National Reference Center for Mycobacteria (Paris) on Löwenstein-Jensen medium by the critical proportion method [10]. Resistance to Ofx was defined as mycobacterial growth at a concentration  $\geq 2 \text{ mg}\cdot\text{L}^{-1}$ . High-level Fq resistance was defined as mycobacterial growth at a concentration  $\geq 2 \text{ mg}\cdot\text{L}^{-1}$  of Mfx.

Statistical analysis was performed with STATA (StataCorp, College Station, TX, USA). Categorical variables were compared using Chi-squared or Fisher's exact test, and continuous variables by the Wilcoxon-Mann-Whitney test. Kaplan-Meier curves for culture conversion were estimated. The Mantel-Cox test was used to compare time to culture conversion between the two groups. A Cox proportional hazards model was used to estimate the association between explanatory variables and time to culture conversion. Variables associated in univariate analysis ( $p < 0.20$ ) were considered for backward multivariable analysis.  $p$ -values  $< 0.05$  were considered as significant.

Bdq was provided under the national compassionate use programme, and patients received information regarding this programme and the safety profile of all drugs. The Institutional Review Board of Bligny's Hospital granted ethical approval.

The cohort included 119 MDR-TB patients with a positive sputum culture at the beginning of anti-TB treatment. Among them, 86 received both linezolid and any second-line injectable drug for  $\geq 30$  days, and 25 Bdq-treated and 42 Fq-treated patients were finally included. The median age of the 67 patients was 33 years (interquartile range (IQR) 27–40 years). A majority was male ( $n=50$ ; 75%) and foreign-born ( $n=60$ ; 90%). 35 (52%) patients harboured isolates susceptible to any Fq and second-line injectable drug, 16 (24%) had isolates with additional resistance to only one of these two drug classes and 16 (24%) to both. Among the 25 Bdq-treated patients, 17 (68%) never received Fq and eight (32%) received levofloxacin (Lfx), Mfx or both successively. Among the 42 Fq-treated patients, 36 (86%) received Mfx, four Lfx and two both successively. Compared with Fq-treated patients, Bdq-treated patients were more likely to be male (96% *versus* 62%;  $p=0.001$ ), born in Eastern Europe (84% *versus* 33%;  $p<0.001$ ), to have received prior TB treatment (92% *versus* 55%;  $p=0.002$ ) and to have bilateral pulmonary involvement (100% *versus* 76%;  $p=0.010$ ). There was no difference regarding the presence of lung cavities and sputum smear status at treatment start. Three patients were HIV positive, all in the Fq-treated group. The median number of drugs to which isolates were susceptible was five in the Bdq group and eight in the Fq group ( $p<0.001$ ). Bdq-treated patients were less likely to receive ethambutol (28% *versus* 59%;  $p=0.022$ ) and ethionamide (20% *versus* 48%;  $p=0.036$ ) but more likely to receive clofazimine (32% *versus* 5%;  $p=0.004$ ) and carbapenem-clavulanate (48% *versus* 2%;  $p<0.001$ ). No difference was found in the proportion of patients treated with pyrazinamide, streptomycin, cycloserine and para-aminosalicylic acid.

The 3-month culture conversion rate was higher in Fq-treated than in Bdq-treated patients (74% *versus* 44%;  $p=0.02$ ), while no statistical difference was found at 6 months (93% *versus* 96%, respectively). The median

(IQR) time to culture conversion was shorter for Fq-treated than for Bdq-treated patients (60 (35–89) days *versus* 98 (70–124) days;  $p=0.005$ ) (figure 1).

In a multivariate proportional hazard model, variables remaining associated with faster time to culture conversion were absence of lung cavities (hazard ratio (HR) 6.60, 95% CI 3.21–13.56;  $p<0.001$ ), negative sputum smear at treatment start (HR 4.73, 95% CI 1.01–22.08;  $p=0.048$ ) and female sex (HR 3.22, 95% CI 1.65–6.30;  $p=0.001$ ). Other variables, including the treatment group, were not significantly associated with time to culture conversion in the multivariate model.

Our study showed no difference in culture conversion rate between Bdq-treated and Fq-treated patients at 6 months. This is promising for Bdq-treated patients, as the 6-month end-point has been linked to successful outcome [11]. Moreover, these results were observed while the characteristics of Bdq-treated patients (bilateral pulmonary involvement, number of drugs for which susceptibility was demonstrated) suggest that they are more difficult to treat than the others.

Nevertheless, time to culture conversion was slower in the Bdq-treated than in the Fq-treated group. This is consistent with previous EBA studies [6, 12]. However, the difference may be due to companion drugs [13]. Indeed, Bdq-treated patients were less likely to receive ethambutol and ethionamide. Interestingly, in the multivariate analysis, Fq-containing regimens were not associated with faster time to culture conversion, while TB characteristics (absence of lung cavitations and smear-negative TB) were the most determinant factors.

Female sex was linked to faster time to culture conversion, but all but one female patient were in the Fq group. Nevertheless, male sex was reported as an independent risk factor for mortality in MDR-TB [14].

In the Bdq-treated group, the achievement of culture conversion closer to the 6-month end-point may suggest continuing Bdq after 24 weeks of treatment [9] for late converters, to prevent culture reversion.

Our study is limited by its observational nature. Moreover, the fact that one third of the Bdq patients with Mfx-resistant isolates received Fq during the treatment course may have affected the results. However, Mfx resistance is almost constantly associated with high-level resistance to other Fq [15]. Hence, it is unlikely that Fq treatment had an impact on culture conversion in the Mfx-resistant Bdq-treated group.

Our study suggests that the 6-month culture conversion rate is similar with Bdq- and Fq-containing regimens. The slower time to culture conversion in the Bdq-treated group could be explained by the patient case mix and differences in the background regimen. Further studies are needed to relate the difference in time to culture conversion with treatment outcome.



@ERSpublications

**Bedaquiline and fluoroquinolone treatments give similar culture conversion rates at 6 months in MDR-TB patients** <http://ow.ly/oqVY300mJCy>

**Lorenzo Guglielmetti<sup>1,2</sup>, Damien Le Dû<sup>1</sup>, Nicolas Veziris<sup>2,3</sup>, Eric Caumes<sup>4</sup>, Dhiba Marigot-Outtandy<sup>1,5</sup>, Yazdan Yazdanpanah<sup>6</sup>, Jérôme Robert<sup>2,3,7</sup> and Mathilde Fréchet-Jachym<sup>1,7</sup> for the MDR-TB Management Group of the French National Reference Center for Mycobacteria and the Physicians of the French MDR-TB Cohort<sup>8</sup>**

<sup>1</sup>Sanatorium, Centre Hospitalier de Bligny, Briis-sous-Forges, France. <sup>2</sup>Sorbonne Universités, UPMC Univ Paris 06, CR7, INSERM, U1135, Centre d'Immunologie et des Maladies Infectieuses, CIMI, Team E13 (Bactériologie), Paris, France. <sup>3</sup>APHP, Centre National de Référence des Mycobactéries et de la Résistance des Mycobactéries aux Antituberculeux (CNR-MyRMA), Bactériologie-Hygiène, Hôpitaux Universitaires Pitié Salpêtrière-Charles Foix, Paris, France. <sup>4</sup>APHP, Service des Maladies Infectieuses et Tropicales, Hôpitaux Universitaires Pitié Salpêtrière-Charles Foix, Paris, France. <sup>5</sup>APHP, Service des Maladies Infectieuses et Tropicales, CHU Raymond Poincaré, Garches, France. <sup>6</sup>APHP, Service des Maladies Infectieuses et Tropicales, Hôpital Bichat Claude Bernard, Paris, France. <sup>7</sup>Both authors contributed equally. <sup>8</sup>A full list of the members of the MDR-TB Management Group of the French National Reference Center for Mycobacteria and of the Physicians of the French MDR-TB Cohort can be found in the Acknowledgements section.

Correspondence: Jérôme Robert, Laboratoire de Bactériologie-Hygiène, Faculté de Médecine Pierre et Marie Curie (UPMC Paris 6), 91 Boulevard de l'hôpital, 75634 Paris Cedex 13, France. E-mail: [jerome.robert@psl.aphp.fr](mailto:jerome.robert@psl.aphp.fr)

Received: Feb 25 2016 | Accepted after revision: April 28 2016 | First published online: June 23 2016

Conflict of interest: Disclosures can be found alongside this article at [erj.ersjournals.com](http://erj.ersjournals.com)

Acknowledgements: The members of the MDR-TB Management Group of the French National Reference Center for Mycobacteria (Paris) are as follows: C. Andrejak, A. Aubry, C. Bernard, F. Brossier, K. Chadelat, B. Dautzenberg, B. Henry, V. Jarlier, M. Jaspard, L. Raskine and B. Rivoire.

The members of the Physicians of the French MDR-TB Cohort are as follows: N. Amiot (Orleans), E. Aslangul (Paris-Hôtel Dieu), P. Assouline (Longjumeau), E. Bergot (Caen), J.F. Boitier (Lagny), A. Bourgarit (Paris-Saint-Louis), A.S. Carrie (Avicenne), M. Caseris (Paris-Bichat), L. Colombain (Perpignan), H. Cordel (Saint Denis), N. De Castro (Paris-Lariboisière), C. Delanoe (Paris-Bichat), V. Delcey (Paris-Lariboisière), M. De Menthon (Paris-Saint-Louis), R. De Meyer-Cristiani (Bastia),

B. Duchemann (Avicenne), J. Dumoulin (Boulogne-Billancourt), C. Duval (Juvisy-sur-Orge), L.I. Escaut (Paris-Kremlin Bicêtre), H. Ferrand (Paris-Bichat), R. Flicoteaux (Paris-Bichat), P. Fraisse (Strasbourg), S. Gallien (Paris-Saint-Louis), S. Girard (Le Mans), C. Godet (Poitiers), M. Gousseff (Avicenne), D. Herman (Nevers), S. Jaureguierry (Paris-Pitié-Salpêtrière), V. Joly (Paris-Bichat), J. Le Grusse (Toulouse), N. Lerolle (Paris-Saint-Louis), E. Leroy-Terquem (Meulan), D. Liné (Soissons), A. Lopes (Paris-Lariboisière), J. Macey (Paris-Cochin), G. Mellon (Paris-Bichat), J.L. Meynard (Paris-Saint-Antoine), J.C. Mouries (Bastia), J.M. Naccache (Paris-Tenon), E. Ngwem (Créteil), G. Oliviero (Longjumeau), B. Philippe (Pontoise), C. Richaud (Paris-Necker), C. Rioux (Paris-Bichat), Z. Saakashvili (Créteil), T.A. Szwabel (Paris-Hôtel Dieu) and P. Vaillant (Nancy).

The authors thank all physicians and microbiologists who provided data for the study.

## References

- 1 Pontali E, Sotgiu G, D'Ambrosio L, *et al.* Bedaquiline and multidrug-resistant tuberculosis: a systematic and critical analysis of the evidence. *Eur Respir J* 2016; 47: 394–402.
- 2 Diacon AH, Pym A, Grobusch MP, *et al.* Multidrug-resistant tuberculosis and culture conversion with bedaquiline. *N Engl J Med* 2014; 371: 723–732.
- 3 Tiberi S, De Lorenzo S, Centis R, *et al.* Bedaquiline in MDR/XDR-TB cases: first experience on compassionate use. *Eur Respir J* 2014; 43: 289–292.
- 4 van Halsema C, Humphreys S, Bonington A. Extensively drug-resistant tuberculosis: early access to bedaquiline for a UK patient. *Eur Respir J* 2014; 43: 292–294.
- 5 Ndjeka N, Conradie F, Schnippel K, *et al.* Treatment of drug-resistant tuberculosis with bedaquiline in a high HIV prevalence setting: an interim cohort analysis. *Int J Tuberc Lung Dis* 2015; 19: 979–985.
- 6 Diacon AH, Dawson R, von Groote-Bidlingmaier F, *et al.* 14-day bactericidal activity of PA-824, bedaquiline, pyrazinamide, and moxifloxacin combinations: a randomised trial. *Lancet* 2012; 380: 986–993.
- 7 Falzon D, Gandhi N, Migliori GB, *et al.* Resistance to fluoroquinolones and second-line injectable drugs: impact on multidrug-resistant TB outcomes. *Eur Respir J* 2013; 42: 156–168.
- 8 Guglielmetti L, Le Dù D, Jachym M, *et al.* Compassionate use of bedaquiline for the treatment of multidrug-resistant and extensively drug-resistant tuberculosis: interim analysis of a French cohort. *Clin Infect Dis* 2015; 60: 188–194.
- 9 World Health Organization. The Use of Bedaquiline in the Treatment of Multidrug-resistant Tuberculosis: Interim Policy Guidance. WHO/HTM/TB/2013.6. Geneva, World Health Organization, 2013.
- 10 Canetti G, Rist N, Grosset J. Mesure de la sensibilité du bacille tuberculeux aux drogues antibacillaires par la méthode des proportions [Measurement of sensitivity of the tuberculous bacillus to antibacillary drugs by the method of proportions. Methodology, resistance criteria, results and interpretation]. *Rev Tuberc Pneumol* 1963; 27: 217–272.
- 11 Kurbatova EV, Cegielski JP, Lienhardt C, *et al.* Sputum culture conversion as a prognostic marker for end-of-treatment outcome in patients with multidrug-resistant tuberculosis: a secondary analysis of data from two observational cohort studies. *Lancet Respir Med* 2015; 3: 201–209.
- 12 Rustomjee R, Diacon AH, Allen J, *et al.* Early bactericidal activity and pharmacokinetics of the diarylquinoline TMC207 in treatment of pulmonary tuberculosis. *Antimicrob Agents Chemother* 2008; 52: 2831–2835.
- 13 Migliori GB, Sotgiu G, Gandhi NR, *et al.* Drug resistance beyond extensively drug-resistant tuberculosis: individual patient data meta-analysis. *Eur Respir J* 2013; 42: 169–179.
- 14 Balabanova Y, Ignatyeva O, Fiebig L, *et al.* Survival of patients with multidrug-resistant TB in Eastern Europe: what makes a difference? *Thorax* 2016; in press [DOI: 10.1136/thoraxjnl-2015-207638].
- 15 Bernard C, Veziris N, Brossier F, *et al.* Molecular diagnosis of fluoroquinolone resistance in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2015; 59: 1519–1524.

Eur Respir J 2016; 48: 582–585 | DOI: 10.1183/13993003.00411-2016 | Copyright ©ERS 2016



# Respiratory decline is integral to disease progression in Huntington's disease



CrossMark

## To the Editor:

Huntington's disease is an autosomal inherited monogenetic condition in which the mutation is an expansion of the cytosine-adenine-guanine (CAG) repeat sequence at the N-terminal end of the huntingtin gene [1]. More than 40 repeats are associated with neuronal dysfunction and death, predominantly within the striatum resulting in a triad of movement, behaviour and cognitive impairment; other symptoms include weight loss, sleep disturbance and respiratory dysfunction, which may or may not be of primary neurological origin [1–3]. Death occurs 15–30 years after onset of symptoms [1], usually due to pneumonia [4], yet it is not known whether respiratory dysfunction is a feature of late stage disease or whether it appears earlier in the disease evolution. Previous research suggests that dysregulation within the respiratory centre results in irregular breathing patterns [5, 6]; decreased respiratory muscle strength and lung volumes have also been identified [7] which, alongside swallow dysfunction [4], could precipitate respiratory failure. Huntington's disease is a complex long-term condition and contributing factors such as swallow dysfunction, posture,