

Tachykinins and airway microvascular leakage induced by HCl intra-oesophageal instillation

S. Daoui*, B. D'Agostino[#], L. Gallelli[#], X. Emonds Alt[†], F. Rossi[#], C. Advenier*

Tachykinins and airway microvascular leakage induced by HCl intra-oesophageal instillation. S. Daoui, B. D'Agostino, L. Gallelli, X. Emonds Alt, F. Rossi, C. Advenier. ©ERS Journals Ltd 2002.

ABSTRACT: Gastro-oesophageal reflux is a common clinical disorder associated with a variety of respiratory symptoms, including chronic cough and exacerbation of asthma. In this study, the potential role of acid-induced tachykinin release was examined in guinea pigs and rabbits, by examining the effects of the tachykinin NK₁ and NK₃ receptors antagonists (SR 140333 and SR 142801, respectively) (1–10 mg·kg⁻¹) on plasma protein extravasation induced in airways by hydrochloric acid (HCl) infusion in the oesophagus.

Guinea pigs were anaesthetised with urethane, while rabbits were subject to neuroleptoanalgesia with hypnorm. Airway vascular leakage was evaluated by measuring extravasation of Evans blue dye. All animals were pretreated with atropine (1 mg·kg⁻¹ *i.p.*), propranolol (1 mg·kg⁻¹ *i.p.*), phosphoramidon (2.5 mg·kg⁻¹ *i.v.*) and saline or tachykinin receptor antagonists (1–10 mg·kg⁻¹ *i.p.*).

Infusion of 1 N HCl into the oesophagus led to a three- and five-fold increase in plasma extravasation in the main bronchi and trachea, respectively. This increase was largely prevented by the tachykinin NK₁ and NK₃ receptor antagonists SR 140333 and SR 142801 (1–10 mg·kg⁻¹).

These results suggest that protein extravasation in the airways, as induced by intra-oesophageal HCl infusion, is mainly dependent on the release of tachykinins, and that both NK₁ and NK₃ tachykinin receptors are involved. The results suggest that HCl-induced sensory nerve stimulation may act in the periphery on intermediate neurons and/or ganglia where NK₃ receptors have been shown to play an important role.

Eur Respir J 2002; 20: 268–273.

*Dept of Pharmacology, University Paris V, Paris, France, [#]Dept of Experimental Medicine, Faculty of Medicine and Surgery, Naples, Italy, [†]Sanofi Synthelabo Research, Montpellier, France.

Correspondence: C. Advenier
Université Paris V
UPRES EA220, Pharmacologie
45 rue des Saints Pères
F-75006 Paris
France
Fax: 33 142863810
E-mail: charles.advenier@wanadoo.fr

Keywords: Airways
oesophageal reflux
tachykinins

Received: June 6 2001
Accepted after revision: January 16 2002

Gastro-oesophageal reflux is a common clinical disorder associated with a variety of respiratory symptoms, including chronic cough and bronchoconstriction [1–4]. It has been shown to be more frequent in people with asthma [4]. Experimental studies have shown that oesophageal hydrochloric acid (HCl) perfusion induces a slight bronchoconstriction in dogs [5], cats [6] and in asthmatic patients [3, 4], and that it potentiates bronchoconstriction induced by isocapnic hyperventilation of dry air or metacholine in asthmatic subjects [4].

The mechanism by which gastro-oesophageal reflux might aggravate asthmatic symptoms remains unclear. Although instillation of acid in airways produces bronchospasm in animals [6–7], pulmonary aspiration of refluxed stomach content has not been convincingly demonstrated in clinical or scintigraphical studies of large groups of asthmatic patients with gastro-oesophageal reflux [8, 9].

Many reports have suggested that gastro-oesophageal reflux could worsen bronchoconstriction in asthmatics via a vagally-mediated reflex triggered by stimulation of the oesophageal acid-sensitive receptors [4, 10, 11]. However, the involvement of C-fibres and tachykinins has recently been reported in a model of airway

neurogenic inflammation, induced by intra-oesophageal HCl stimulation in guinea pigs [12].

In the airways, the activation of C-fibre afferent nerves leads to local release of tachykinins, a group of neuropeptides, including substance P (SP), neurokinin A (NKA) and neurokinin B (NKB). Tachykinins are present in several types of peripheral tissues, including gastro-intestinal tract, lung and bladder [13]. They are particularly responsible for several biological effects in the airways, including: bronchospasm, increase in microvascular permeability, vasodilatation, stimulation of glandular secretions, facilitation of cholinergic neurotransmission, and the recruitment and activation of inflammatory cells. Moreover, sensory nerves also mediate respiratory defence reflexes, such as coughing and sneezing [13–15].

The biological actions of tachykinins are mediated via three types of receptors, denoted tachykinin NK₁, NK₂ and NK₃, which have the highest affinity for SP, NKA and NKB, respectively [16]. This receptor classification has been established from receptor-binding and functional studies using selective agonists or antagonists for tachykinin receptors [16]. It has now been recognised that the expression of the tachykinin NK₃ receptor is confined mainly to the

central and peripheral nervous system, whilst tachykinin NK₁ and NK₂ receptors are expressed both in the nervous system and target organs, including airways [17, 18].

HAMAMOTO *et al.* [12] have shown that HCl infusion in guinea-pig oesophagus causes plasma extravasation in the airways. This effect is abolished in animals treated with capsaicin, a drug that induces a depletion of tachykinins from sensory nerves, or by the tachykinin NK₁ receptor antagonist FK 888. As these effects were observed in animals pretreated with atropine and propranolol and were abolished by bilateral vagotomy, HAMAMOTO *et al.* [12] suggested that HCl effects in that airway involve nerve pathways in the periphery.

It has been suggested that tachykinin NK₃ receptors play an important role in parasympathetic ganglia and nerve transmission in the periphery [18–20]. Thus, the current authors postulated that the tachykinin NK₃ receptor antagonist SR 142801 [21] might prevent HCl-induced neurogenic inflammation in the airways. To test this hypothesis, the airway plasma extravasation in the guinea pig was examined by stimulating the oesophagus using HCl, in the presence of SR 142801 or SR 140333, an NK₁ receptor antagonist [22]. To extend these observations to another species, a similar set of experiments were performed on rabbits.

Materials and methods

Oesophageal stimulation by hydrochloric acid

New Zealand white rabbits (2.5–2.8 kg) or tricolour guinea pigs (300–400 g) of either sex were used throughout the study. The guinea pigs were anaesthetised with urethane (1.25 g·kg⁻¹ *i.p.*; Prolabo, Paris, France). Rabbits were subjected to neuroleptoanalgesia. They were premedicated with an intraperitoneal injection of diazepam (5 mg·mL⁻¹, 5 mg·kg⁻¹; Roussel Pharma SpA, Milan, Italy) and subsequently administered hypnorm (0.4 mL·kg⁻¹; a mixture of fentanyl citrate 0.315 mg·mL⁻¹ and fluanisone 10 mg·mL⁻¹ *i.m.*; Janssen Pharmaceutical Ltd, Grove, Oxfordshire, UK). Guinea pig jugular veins or rabbit-ear marginal veins were cannulated for injection of drugs. Thirty min before experimentation, all animals were pretreated with atropine (1 mg·kg⁻¹ *i.p.*; Sigma, St Louis, MO, USA) and propranolol (1 mg·kg⁻¹ *i.p.*; Sigma) to block muscarinic and β -adrenergic receptors, and phosphoramidon (2.5 mg·kg⁻¹ *i.v.*, 10 min before exposure to HCl (Prolabo); Sigma) to inhibit tachykinin metabolism.

The oesophageal wall was partly sectioned at the level of the 3rd–5th tracheal cartilage ring. In guinea pigs, a catheter was placed in the midoesophagus. Thus, the oesophagus was ligated at the upper portion to inhibit HCl leakage. The lower end of the oesophagus from the abdomen was then exposed and ligated to block communication between the oesophagus and the stomach. In rabbits, a second catheter with an attached latex balloon was inflated to block communication between the oesophagus and

the stomach. Thus, there was no leakage of fluid from the oesophagus to the stomach.

Measurement of airway microvascular leakage

The vascular permeability was quantified by the extravasation of Evans blue dye [23]. Evans blue dye (30 mg·kg⁻¹) was injected into the rabbit's ear marginal vein or the guinea pig's jugular vein, followed 1-min later by intra-oesophageal HCl 1 N or saline (0.4 mL). After the induction of leakage (10 min after completion of the intra-oesophageal HCl infusion), the thorax was opened and a blunt-ended 13–14-gauge needle was passed through a left ventriculotomy into the aorta. The ventricle was cross clamped and blood was expelled through an incision into the right atrium at 80 mmHg pressure with ~100 mL (guinea pig) or ~700 mL (rabbit) of phosphate buffer. The lung was then removed. The connective tissues, vasculature and parenchyma were gently scraped away and the airways were divided into four components: trachea, main bronchi and proximal and distal intrapulmonary airways [23]. The tissues were blotted dry, placed in preweighed tubes and reweighed, and their dye content was extracted in formamide at 37°C for 18 h. Dye concentration was quantified by light absorbance at 620 nm (DCP spectrophotometer; Vital, Dieren, the Netherlands), and tissue content (ng dye·mg wet weight tissue⁻¹) was calculated from a standard curve of dye concentrations in the 0.5–10 μ g·mL⁻¹ range.

Pretreatment with drugs

Guinea pigs or rabbits received a single dose (1–10 mg·kg⁻¹ *i.p.*) of the NK₃ (SR 142801; Sanofi Recherche, Montpellier, France) or NK₁ (SR 140333; Sanofi Recherche) receptor antagonist, or vehicle 30 min before exposure to HCl.

Capsaicin pretreatment. A group of rabbits was treated with capsaicin (total dose 80 mg·kg⁻¹ *s.c.*; Pierce, Rockford IL, USA) or vehicle (10% ethanol, 10% tween 80 and 80% NaCl 0.9%) administered over a 3-day period (5 mg·kg⁻¹ *s.c.* on day 1, 50 mg·kg⁻¹ *s.c.* on day 2 and 25 mg·kg⁻¹ *s.c.* on day 3). The volume of injection was 1 mL·kg⁻¹.

Statistical analysis of results

Data are expressed as mean \pm SEM. Statistical analysis of the results was performed using the unpaired t-test. *p*-Values of <0.05 were considered significant.

Results

Effect of phosphoramidon on airway plasma extravasation induced by oesophageal stimulation with hydrochloric acid in guinea pigs and rabbits

There were no significant differences at any airway level in extravasation of Evans blue dye after sham

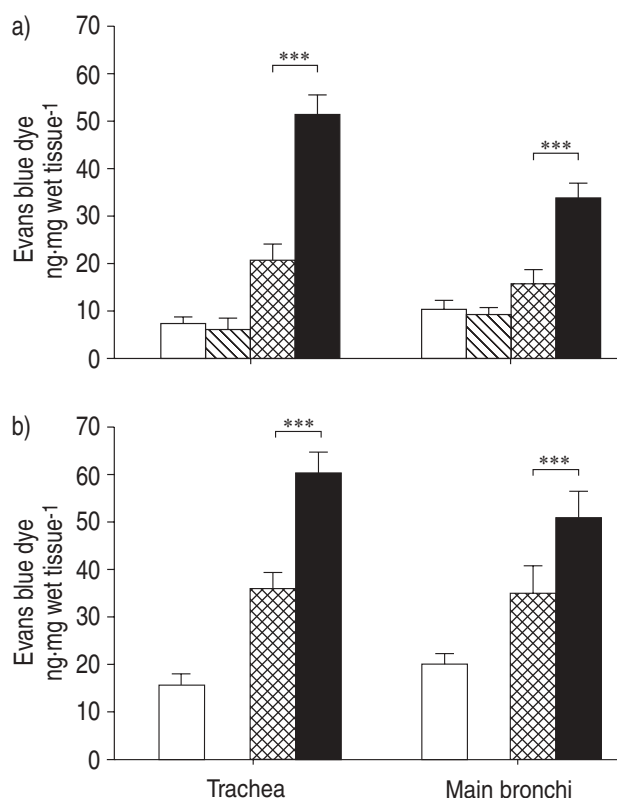


Fig. 1.—Histograms illustrating the effects of intra-oesophageal hydrochloric acid (HCl) (1 N, 10 min) on microvascular leakage in a) guinea-pig or b) rabbit airways 30 min after intraperitoneal administration of atropine and propranolol (1 mg·kg⁻¹) and 10 min after intravenous administration of phosphoramidon (2.5 mg·kg⁻¹) or saline (NaCl 0.9%). Data are presented as means±SE, n=5–6. □: saline and saline; ▨: phosphoramidon and saline; ▤: saline and HCl; ■: phosphoramidon and HCl. ***: p<0.001.

stimulation with intra-oesophageal saline (0.9%) infusion in guinea pigs pretreated with either saline or phosphoramidon (fig. 1). In both species, infusion of HCl (1 N) into the oesophagus without phosphoramidon significantly increased (p<0.01) plasma extravasation in the trachea and the main bronchi, and this effect was potentiated by phosphoramidon (p<0.001) (fig. 1). No effects were observed in the proximal and distal intrapulmonary airways (results not shown).

Influence of hydrochloric acid concentration on airway plasma extravasation in guinea pigs

Only infusion of HCl at 1 N into the oesophagus in the presence of phosphoramidon induced a significant plasma extravasation in guinea pigs pretreated with atropine, propranolol and phosphoramidon, whereas 0.3 and 3 N HCl did not (fig. 2).

Effects of SR 140333 and SR 142801 on plasma extravasation in guinea-pig airways

The plasma extravasation was significantly inhibited in guinea pigs by the tachykinin receptor antagonists

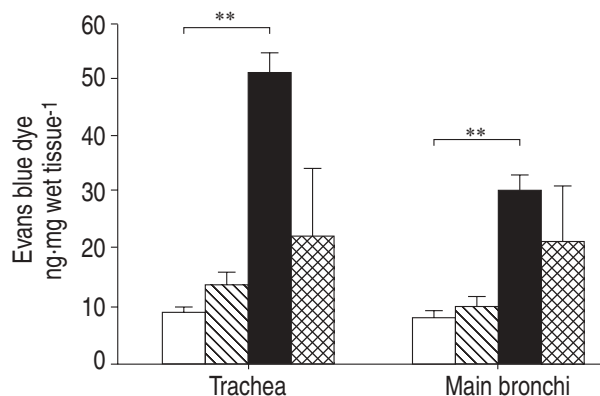


Fig. 2.—Histograms illustrating the effects of intra-oesophageal hydrochloric acid (HCl) concentration (0.3–3 N, 10 min) on airway plasma exudation in airways in guinea pigs pretreated with atropine, propranolol and phosphoramidon. Data are presented as means±SE, n=6. □: saline; ▨: HCl 0.3 N; ▤: HCl 1 N; ■: HCl 3 N. **: p<0.01.

SR 140333 and SR 142801 (3–10 mg·kg⁻¹) (fig. 3). When the two tachykinin receptor antagonists were administered together (3 mg·kg⁻¹ of each), the inhibition

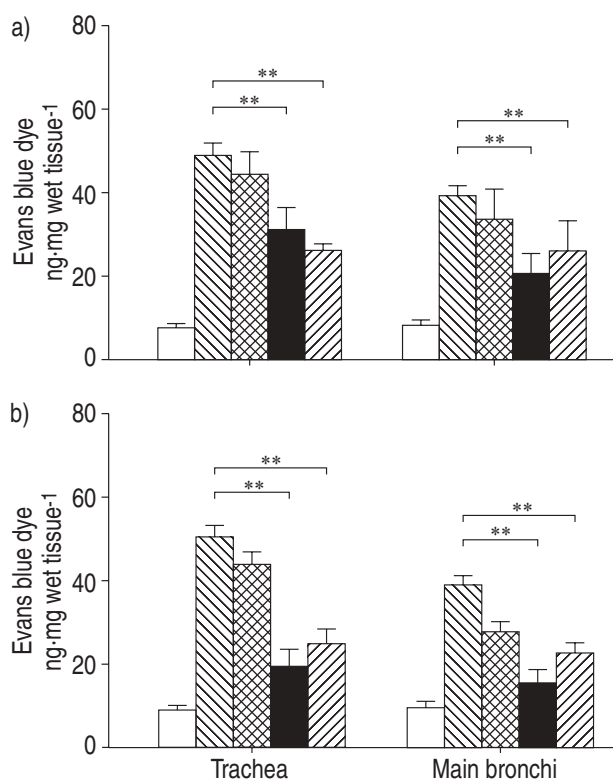


Fig. 3.—Effects of the tachykinin NK₁ or NK₃ receptor antagonists, a) SR 140333 or b) SR 142801, on the vascular leakage induced in guinea-pig trachea and main bronchi by intra-oesophageal hydrochloric acid (HCl) infusion. Tachykinin receptor antagonists (1–10 mg·kg⁻¹) were given 30 min before HCl infusion. Guinea pigs were pretreated with atropine, propranolol and phosphoramidon. Data are presented as means±SE, n=6/group. □: control without HCl; ▨: control and HCl (1 N); ▤: 1 mg·kg⁻¹; ■: 3 mg·kg⁻¹; ▩: 10 mg·kg⁻¹. **: p<0.01.

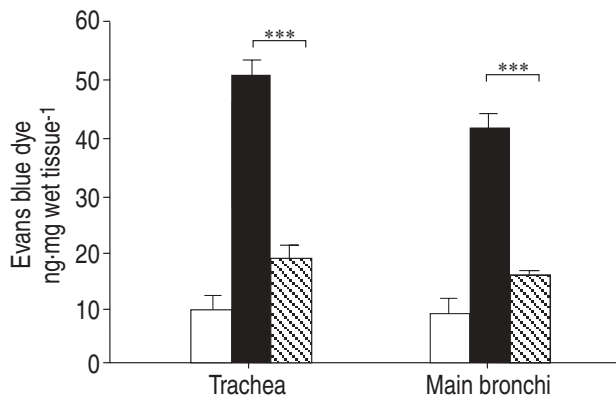


Fig. 4.—Effects of SR 140333 and SR 142801 on the vascular leakage induced by intra-oesophageal hydrochloric acid (HCl) infusion in the guinea-pig trachea and main bronchi. Doses of tachykinin receptor antagonists were given 30 min before HCl intra-oesophageal infusion. Guinea pigs were pretreated with atropine, propranolol and phosphoramidon. Data are presented as means \pm SE, $n=6$. □: control without HCl; ■: control and HCl (1 N); ▨: SR 140333 and SR 142801 (3 mg·kg⁻¹ i.p.). ***: $p<0.001$.

of HCl-induced plasma extravasation in the airways was also significant ($p<0.001$) (fig. 4), but no additional effect or synergy was observed.

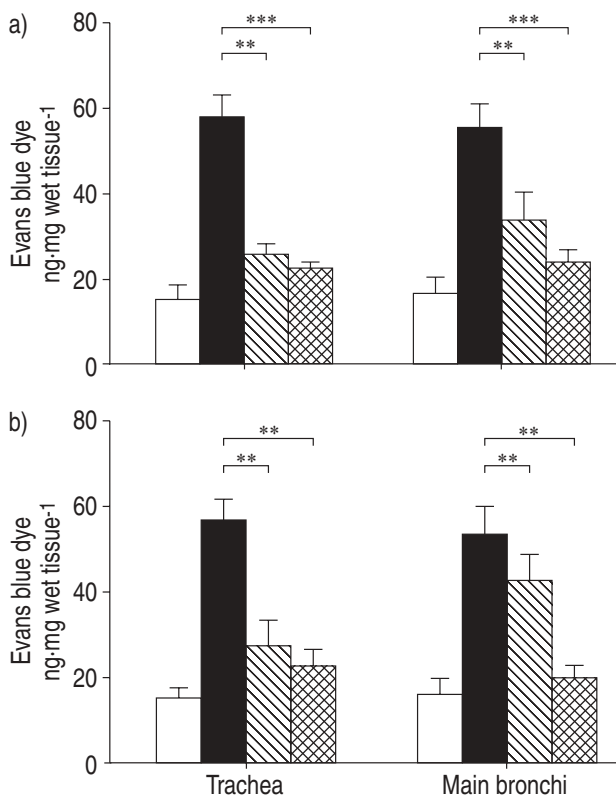


Fig. 5.—Effects of the tachykinin NK₁ or NK₃ receptor antagonists, a) SR 140333 or b) SR 14281, on the vascular leakage induced in rabbit trachea and main bronchi by intra-oesophageal hydrochloric acid (HCl) infusion. Doses of tachykinin receptor antagonists were given 30 min before intra-oesophageal HCl (1 N, 10 min) infusion. Rabbits were pretreated with atropine, propranolol and phosphoramidon. Data are presented as means \pm SE, $n=5$. □: control without HCl; ■: control and HCl; ▨: 1 mg·kg⁻¹; ▩: 3 mg·kg⁻¹. **: $p<0.01$; ***: $p<0.001$.

Effects of SR 140333 and SR 142801 on plasma extravasation in rabbit airways

Figure 5 shows that both SR 140333 and SR 142801 (1 mg·kg⁻¹) induced a strong inhibition ($p<0.01$ – $p<0.001$) of plasma extravasation in rabbit airways following HCl instillation in the oesophagus. The effects were more pronounced in trachea than in main bronchi.

Effects of systemic capsaicin treatment on hydrochloric acid-induced airway plasma extravasation in rabbits

The plasma extravasation in airways induced by HCl was significantly abolished in the capsaicin-treated rabbits (fig. 6).

Discussion

In the present study, it has been shown that in both guinea pigs and rabbits pretreated with atropine and propranolol to block muscarinic and adrenergic receptors, oesophageal stimulation by HCl causes neurogenic inflammation in the trachea and main bronchi. Airway plasma extravasation was abolished by capsaicin in rabbits, in agreement with previous results obtained in guinea pigs by HAMAMOTO *et al.* [12]. These authors also demonstrated that this effect was inhibited in the guinea pig by the tachykinin NK₁ receptor antagonist, FK 888. The current authors provide evidence that, in addition to the tachykinin NK₁ receptor antagonist SR 140333, the NK₃ receptor antagonist SR 142801 was able to prevent neurogenic inflammation in airways both in guinea pigs and rabbits. These results suggest that nonadrenergic, noncholinergic (NANC) nerves, communicating between the oesophagus and airways, are implied in this response and that, in addition to NK₁

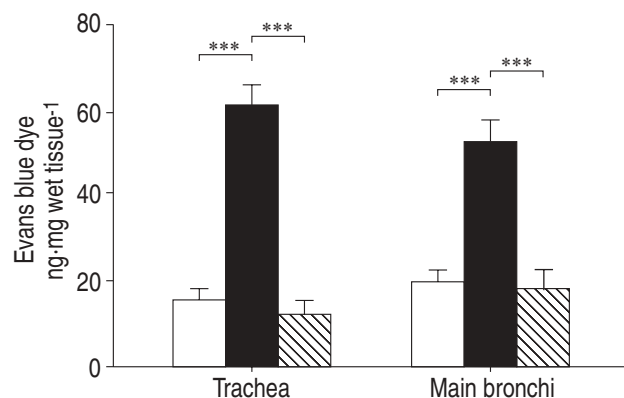


Fig. 6.—Effects of capsaicin pretreatment on plasma extravasation induced by intra-oesophageal stimulation by hydrochloric acid (HCl) (1 N, 10 min) in the rabbit. Animals were pretreated with atropine, propranolol and phosphoramidon. Data are presented as means \pm SE, $n=5$. □: control without HCl; ■: control and HCl (1 N); ▨: capsaicin and HCl (1 N). ***: $p<0.001$.

receptors, NK₃ receptors are involved in transmission of the neurogenic response.

Neurogenic inflammation, induced by oesophageal instillation of HCl in the airways of guinea pigs or rabbits, is not associated with leakage from intra-oesophageal HCl [12], but involves nerve pathways communicating between the oesophagus and airways. A vagally-mediated autonomic pathway may be involved in the experimental models described here, as HAMAMOTO *et al.* [12] have shown that airway leakage induced by HCl was abolished in guinea pigs by bilateral vagotomy.

Nerve pathways communicating between the oesophagus and airway may also be in the periphery, *via* autonomic ganglia [18, 19]. Anatomical studies have demonstrated the presence of axon collaterals from visceral afferent fibres (gastro-intestinal tract, bladder, airways) in autonomic ganglia [19, 24, 25]. Furthermore, studies carried out in isolated ganglia have demonstrated effects of these afferent nerves on the electrophysiological properties of the autonomic ganglion neurons after selective stimulation of their sensory/afferent nerve terminals [19]. Interestingly, CANNING *et al.* [20] have reported that, *in vitro*, in a preparation of guinea-pig trachea and oesophagus with associated nerves, and in the presence of propranolol, atropine, indomethacin and the tachykinin NK₂ receptor antagonist SR 48968, capsaicin elicited concentration-dependent relaxations of the trachea only when the adjacent oesophagus was intact. The effect of capsaicin was abolished by tetrodotoxin. On the basis of these results and of additional immunochemistry studies, the authors concluded that the NANC responses of the guinea pig trachea were mediated by nerve fibres emanating from parasympathetic ganglia that are intrinsic to the adjacent oesophagus [20, 26, 27]. In a similar manner, the current authors suggest that stimulation by HCl of capsaicin-sensitive fibres in the oesophagus may elicit neurogenic inflammation in airways through intermediate neurons, including parasympathetic ganglion neurons.

The results from the current study demonstrate that, in rabbits as in guinea pigs, both tachykinin NK₁ and NK₃ receptors antagonists SR 140333 and SR 142801 were able to inhibit airway extravasation induced by HCl. The inhibitory effect of NK₁ receptor antagonists has previously been shown by HAMAMOTO *et al.* [12] in guinea pigs with the compound FK 888. Inhibition by FK 888 or SR 140333 may be caused by direct inhibition of endogenous tachykinin effects on vascular cells, since NK₁ receptors play a predominant role in microvascular leakage induced by tachykinins [23, 28], although a role in nerve transmission cannot be excluded.

In contrast, the inhibitory effect of the tachykinin NK₃ receptor antagonist, SR 142801, was less anticipated, since the NK₃ receptor does not seem to be involved in the impairment of vessels and endothelial cells leading to microvascular leakage [17]. An effect of SR 142801 on NK₁ receptors can also be excluded, since the selectivity of this compound has clearly been demonstrated by radioligand binding and well characterised *in vitro* by functional

assays for tachykinin receptors [21, 29, 30]. *In vivo*, the selectivity of SR 142801 is clearly suggested by two assays: in contrast to SR 48968, SR 142801 (1 mg·kg⁻¹) did not inhibit bronchoconstriction induced by [Nle¹⁰] neurokinin A [4–10] in anaesthetised guinea-pigs [29]; and unlike SR 140333, SR 142801 (1 mg·kg⁻¹) failed to inhibit the hypotension induced by [Sar⁹, Met (O₂)¹¹] substance P in guinea pigs and dogs [22, 31]. It has also been shown that SR 142801 failed to inhibit tachykinin and capsaicin-induced plasma extravasation in the ear skin of mice, an effect that could be suppressed by SR 140333 [32].

These results are in agreement with those of CANNING *et al.* [20] who demonstrated in their preparations of guinea pig trachea and oesophagus with associated nerves, that SR 142801 was able to inhibit tracheal responses to capsaicin. This suggests that, in the current *in vivo* model, HCl-induced sensory nerve stimulation may act on intermediate neurons and parasympathetic neurons where NK₃ receptors play a predominant role [18, 19].

An effect of the NK₃ receptor antagonist SR 142801 on nonadrenergic, noncholinergic nerves in periphery has also been reported in the guinea pig in other experimental models of airway hyperresponsiveness. Indeed, SR 142801 is able to prevent airway hyperresponsiveness to methacholine and the increased response to histamine on microvascular leakage in airways of guinea pigs exposed 24 h earlier to an aerosol of substance P or of citric acid [29, 33]. In this paper, it has been shown that SR 142801 has additional potential in inhibiting airway responses linked to acidity and gastro-oesophageal reflux.

References

1. Ing AJ, Ngu MC, Breslin AB. Chronic persistent cough and gastro-oesophageal reflux. *Thorax* 1991; 46: 479–483.
2. Irwing RS, Zawacki JK, Curley FJ, French CL, Hoffman PJ. Chronic cough as the sole presenting manifestation of gastroesophageal reflux. *Am Rev Respir Dis* 1989; 140: 1294–1300.
3. Spaulding HS, Mansfield LE, Stein MR, Sellner JC, Cremillon DE. Further investigation of the association between gastroesophageal reflux and bronchoconstriction. *J Allergy Clin Immunol* 1982; 69: 516–521.
4. Hervé P, Denjean A, Jian R, Simonneau G, Duroux P. Intraesophageal perfusion of acid increases the bronchomotor response to metacholine and to isocapnic hyperventilation in asthmatic subjects. *Am Rev Respir Dis* 1986; 134: 986–989.
5. Mansfield LE, Harneister HH, Spaulding HS, Smith NJ, Glab N. The role of the vagus nerve in airway narrowing caused by intraesophageal hydrochloric acid provocation and esophageal distension. *Ann Allergy* 1981; 47: 431–434.
6. Tuchman DN, Boyle JT, Pack AI, *et al.* Comparison of airway responses following tracheal or esophageal acidification in the cat. *Gastroenterology* 1984; 87: 872–881.
7. Ishikawa T, Sekizawa SI, Sant'Ambrogio FB, Sant'Ambrogio G. Larynx versus esophagus as

- reflexogenic sites for acid-induced bronchoconstriction in dogs. *J App Physiol* 1999; 86: 1226–1230.
8. Berquist JD, Rachelefsky GS, Kadden M, *et al.* Gastroesophageal reflux associated recurrent pneumonia and chronic asthma in children. *Pediatrics* 1981; 68: 29–35.
9. Ghaed N, Stein MR. Assessment of a technique for scintigraphic monitoring of pulmonary aspiration of gastric contents in asthmatics with gastroesophageal reflux. *Ann Allergy* 1979; 42: 306–308.
10. Mansfield LE, Stein MR. Gastroesophageal reflux and asthma: possible reflex mechanism. *Ann Allergy* 1978; 41: 224–226.
11. Kjellen G, Tibbling L, Wranne B. Bronchial obstruction after acid perfusion in asthmatics. *Clin Physiol* 1981; 1: 285–292.
12. Hamamoto J, Kohrogi H, Kawano O, *et al.* Esophageal stimulation by hydrochloric acid causes neurogenic inflammation in the airways in guinea-pigs. *J Appl Physiol* 1997; 82: 738–745.
13. Maggi CA, Patacchini R, Rovero P, Giachetti A. Tachykinin receptors and tachykinin receptor antagonists. *J Auton Pharmacol* 1993; 13: 23–93.
14. Ellis JL, Undem BJ. Pharmacology of nonadrenergic, noncholinergic nerves in airway smooth muscle. *Pulm Pharmacol* 1994; 7: 205–223.
15. Widdicombe JG. Neurophysiology of the cough reflex. *Eur Respir J* 1995; 8: 1193–1202.
16. Regoli D, Boudon A, Fauchère JL. Receptors and antagonists for substance P and related peptides. *Pharmacol Rev* 1994; 46: 551–599.
17. Baluk P, McDonald DM. Proinflammatory peptides in sensory nerves of the airways. In: Said SS, ed. *Proinflammatory peptides*. New York, Marcel Dekker, 1998; pp. 45–85.
18. Myers A, Undem B. Electrophysiological effects of tachykinins and capsaicin on guinea-pig bronchial parasympathetic ganglion neurones. *J Physiol* 1993; 470: 665–679.
19. Myers A, Undem B, Kummer W. Anatomical and electrophysiological comparison of the sensory innervation of bronchial and tracheal parasympathetic ganglion neurons. *J Auton Nerv System* 1996; 61: 162–168.
20. Canning BJ, Fischer A, Undem J. Pharmacological analysis of the tachykinin receptors that mediate activation of nonadrenergic, noncholinergic relaxant nerves that innervate guinea-pig trachealis. *J Pharmacol Exp Ther* 1998; 284: 370–377.
21. Emonds-Alt X, Bichon D, Ducoux JP, *et al.* SR 142801, the first potent non-peptide antagonist of the tachykinin NK₃ receptor. *Life Sci* 1995; 56: L27–L32.
22. Emonds-Alt X, Doutremepuich JD, Heaulme M *et al.* *In vitro* and *vivo* biological activities of SR 140333, a novel potent non-peptide tachykinin NK₁ receptor antagonist. *Eur J Pharmacol* 1993; 250: 403–413.
23. Rogers DF, Belvisi MG, Aursudkij B, Evans TW, Barnes PJ. Effects and interaction of sensory neuropeptides on airway microvascular leakage in guinea-pigs. *Br J Pharmacol* 1988; 95: 1109–1116.
24. Coleridge HM, Coleridge JCG, Schultz HD. Afferent pathways involved in reflex regulation of airway smooth muscle. *Pharmacol Ther* 1989; 42: 1–63.
25. Mawe GM. Tachykinins as mediators of slow EPSPs in guinea-pig gall-bladder ganglia: involvement of neurokinin-3 receptors. *J Physiol (Lond)* 1995; 485: 513–524.
26. Canning BJ, Undem BJ. Evidence that distinct neural pathways mediate parasympathetic contractions and relaxations of guinea-pig trachealis. *J Physiol* 1993; 471: 25–40.
27. Canning BJ, Undem BJ, Karakousis PC, Dey RD. Effects of organotypic culture on parasympathetic innervation of guinea-pig trachealis. *Am J Physiol* 1996; 271: L698–L706.
28. Lei YHL, Barnes PJ, Rogers DF. Inhibition of neurogenic plasma exudation in guinea-pig airways by CP-96,345, a new non-peptide NK₁ receptor antagonist. *Br J Pharmacol* 1992; 105: 261–262.
29. Daoui S, Cui YY, Lagente V, Emonds-Alt X, Advenier C. A tachykinin NK₃ receptor antagonist, SR 142801 (Osanetant), prevents substance P-induced bronchial hyperactivity in guinea-pigs. *Pulm Pharmacol Ther* 1997; 10: 261–270.
30. Nguyen-Le XK, Nguyen QT, Gobeil F, *et al.* Pharmacological characterization of SR 142801: A non-peptide antagonist of the neurokinin NK₃ receptor. *Pharmacology* 1996; 52: 283–291.
31. Roccon A, Marchionni D, Nisato D. Study of SR 142801, a new potent non-peptide NK₃ receptor antagonist on cardiovascular responses in conscious guinea-pig. *Br J Pharmacol* 1996; 118: 1095–1102.
32. Inoue H, Nagata N, Koshihara Y. Involvement of tachykinin receptors in oedema formation and plasma extravasation induced by substance P, neurokinin A, and neurokinin B in mouse ear. *Inflamm Res* 1996; 45: 316–323.
33. Daoui S, Cognon C, Naline E, Emonds-Alt X, Advenier C. Involvement of tachykinin NK₃ receptors in citric acid-induced cough and bronchial responses in guinea-pigs. *Am J Respir Crit Care Med* 1998; 158: 42–48.